Chapter 5

5.1 Introduction

Modeling of chemical parameters in the CES

The present structure and behavior patterns of estuaries are a result of the various biogeo and physicochemical processes. The level of complexity of the system depends on the number of interacting or superimposed processes and increases generally with time. The knowledge on the dynamics of evolution and the present state of affairs is essential for the development of a working model. Models should not only explain the experimental data but should predict future probability. This is an attempt to predict the overall reactivity of the Cochin Estuarine System in terms of the behaviour of boron.

The estuarine systems are having a special significance in the hydrosphere because of its role as a link between land and sea. The impact of human activities is at its maximum in the estuarine waters. The agricultural drains, industrial effluents sewage and sullage etc deliver considerably to this system modifying the chemical characteristics. Compared to the rivers and oceans, the estuarine waters are the most vulnerable points in terms of pollution. A special significance is attributed in the hydrosphere to the estuarine systems because of the multitude and complexity of the processes; the system can not be defined by interpreting any of the individual properties. The various independent but interdependent processes include geochemical processes such as sedimentation, resuspension, precipitation and

modification etc, and biochemical process such as uptake, depuration, death and decay. The seawater undergoes various chemical and physical modifications during its mixing with fresh water from the land. The dilution of seawater, which is the major phenomena during mixing, is associated with a re speciation of the elements present in these waters also. The speciation and residence time of any species in an estuary is dependent on fresh water discharge, density gradients, atmospheric input, tidal and water current pattern. No comprehensive model is available which can be applied to an estuarine system in its totality.

The various approaches adopted for the interpretation of estuarine system can be grouped in to two and primarily utilises only the physical parameters. In the first group the averaging of tidal flow (Tee, 1976; Sinha et al., 1995, 1998, Prandle, 1978) and in the second group time averaged equations of motions (Nihoul and Ronday, 1987) were

5.1 Introduction and Raney, 1991): The model developed in this study, though

The present structure and behavior patterns of estuaries are a result of the various biogeo and physicochemical processes. The level of complexity of the system depends on the number of interacting or superimposed processes and increases generally with time. The knowledge on the dynamics of evolution and the present state of affairs is essential for the development of a working model. Models should not only explain the experimental data but should predict future probability. This is an attempt to predict the overall reactivity of the Cochin Estuarine System in terms of the behaviour of boron.

The estuarine systems are having a special significance in the hydrosphere because of its role as a link between land and sea. The impact of human activities is at its maximum in the estuarine waters. The agricultural drains, industrial effluents sewage and sullage etc deliver considerably to this system modifying the chemical characteristics. Compared to the rivers and oceans, the estuarine waters are the most vulnerable points in terms of pollution. A special significance is attributed in the hydrosphere to the estuarine systems because of the multitude and complexity of the processes; the system can not be defined by interpreting any of the individual properties. The various independent but interdependent processes include geochemical processes such as sedimentation, resuspension, precipitation and

dissolution, chemical transformation such as complexation, exchange, species modification etc, and biochemical process such as uptake, depuration, death and decay. The seawater undergoes various chemical and physical modifications during its mixing with fresh water from the land. The dilution of seawater, which is the major phenomena during mixing, is associated with a re speciation of the elements present in these waters also. The speciation and residence time of any species in an estuary is dependent on fresh water discharge, density gradients, atmospheric input, tidal and water current pattern. No comprehensive model is available which can be applied to an estuarine system in its totality.

The various approaches adopted for the interpretation of estuarine system can be grouped in to two and primarily utilises only the physical parameters. In the first group the averaging of tidal flow (Tee, 1976; Sinha et al., 1995, 1998; Prandle, 1978) and in the second group time averaged equations of motions (Nihoul and Ronday, 1987) were used. A few theoretical models using the baraclinic effects are also available (Smith and Chang, 1987; Jin and Raney, 1991). The model developed in this study, though considers both tidal contribution and time averaged flow, and basically is different from these two. In the first phase, because of the regular and continuous pattern of the physical and chemical processes, the system is considered to be at a steady state and so the initial state will not have much significance in defining the system. Secondly chemical reactivity of the estuary is assumed to regulate the species concentration of any element in the system than the physical processes. Thirdly, in positive estuarine system like CES, where regulate flow of fresh water and tidal intrusion of sea water are present, the mixing time and the flushing time can be considered as better representative of the physical processes rather than the tidal flow.

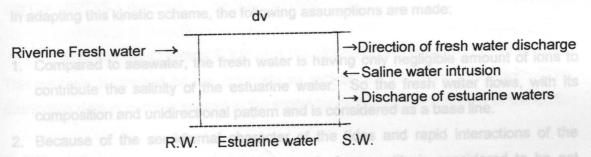
5.2 The System

- 1. As mentioned in earlier chapters, the CES is a tropical positive estuarine system with two distinct arms through the rivers Periyar at the North and Muvattupuzha at the South. These rivers which flows through the adjoining land masses of agriculture, are the major contributors of fresh water to the system and so contains considerable vegetative matter.
- 2. The Barmouth, ca 450m wide, is the only permanent opening to the sea.
- The average depth of the estuary is around 3m and the maximum tidal height is about 1m. The tidal processes thus have profound influence in this system.

- 4. During monsoon months, the river and estuary get flooded with fresh water and will show only fresh water characteristics.
- 5. During postmonsoon and premonsoon the riverine flow is considerably slow and the estuarine system up to the river mouths will give saline characteristics.
- 6. Though the system is highly positive with a flushing time of 7 days and the depths are very low, significant vertical salinity gradient is observed in the entire estuarine region.
- 7. Though the average depth is very low, the high organic content in the sediments results in an anoxic condition in the sediments.

5.3 The Model

The estuarine system with its riverine discharge and saline water intrusion can be considered in a broader sense a flow reactor.



The reactor start from the river mouth where salinity is zero or near to zero and ends at the Barmouth where salinity is equivalent to the salinity of seawater. Through the reactor the reactant 1, the fresh water, flows at a flow rate of V m/s. The reactant 2, the seawater, enters the reactor at a velocity that can be indicated by the time average of high tide and leaves the reactor by the time average of low tide.

If U liter/ S is considered as the rate of flow of the chemical entity to the reactor and C the concentration of the entity, then during a time interval dt the amount of the species that is transferred to the reactor will be UC dt. If the order of the reaction in the rate transformation

$$V = -dc/dt = k C^n$$
.

If dv is the volume of the reactor of reaction that will take place in a time interval dt, the rate $v = -dc/dt = kc^n dv$.

After a period of time the component leaves the compartment at the rate of U (C + dc)dt. Because of the regular cycling feature, the reactor is assumed to be in a steady state: or the rate of addition and the rate of removal are equal.

The steady state contains contributions from

- (1) the amount of species that enters the reactor, i.e., U C dt.
- (2) the amount of species that leaves the compartment i.e., U (C+dc) dt.
- (3) the amount of species that undergoes reaction i.e., k Cⁿ dv dt.

At the steady state UC dt = U (C+dc) dt + kCⁿ dv dt.

Rearranging -U dc dt = kC^n dv dt or $dc/C^n = -k dv/U$

In adapting this kinetic scheme, the following assumptions are made:

- Compared to seawater, the fresh water is having only negligible amount of ions to contribute the salinity of the estuarine water. So the fresh water flows, with its composition and unidirectional pattern and is considered as a base line.
- Because of the semidiurnal character of the tides and rapid interactions of the chemical entities, the biochemical removal of any entity is considered to be not affecting the chemical environment of the system.
- 3. The sedimentary incorporation and exchange is regarded as the major phenomena in the regulation of availability of a particular species of the element in the estuarine waters. Considering the concentration changes associated with the mixing processes and possible magnitude of the unidentifiable interdependent multitude of the various transformations, the overall order of the reaction is considered to be one.
- 4. The tidal distribution of salinity is more or less regularised by the riverine flow and can be considered as in a steady state. The horizontal salinity gradient is a function of the ratio of flow rate to tide height.
- 5. The major ions, which constitute the salinity because of its conservative character generally, will exhibit a normalised distribution pattern with the addition of fresh water.
- 6. In a natural estuarine system, though the river flow can be measured precisely, the salinity intrusion, or the seawater influx, can not be measured. At the same time, the dilution patterns can be taken as an index of the rate of flow of seawater to the

- estuary or from the estuary. As mixing will lead to a general dilution pattern, the volume element can be replaced by the salinity element.
- 7. Boron is a major element, which is conservative in seawater, but susceptible to biogeochemical modifications. So a comparison of the dilution patterns of boron with salinity can give an idea on the biogeochemical reactivity of the system.

The equation with this modification takes the form

dC/C = - Kds/T where ds is the salinity element and T time averaged tidal height during the addition of seawater. T can be considered as a function of the physical processes operating in the system and K the geochemical reactivity. The ratio K/T can be considered as an index of the total estuarine reactivity that varies with changes on the physical characteristics processes and is a constant at a particular period. This on integration

log C = - (K/T) S + constant. = - Ke S + Constant

A plot of log C versus S will give the slope Ke, the reactivity of the estuary.

The reactivity is depend on the substrate as well as the estuarine character. The distribution data of boron is applied to the model for estimating the reactivity of the CES.

The concept of constantancy of major ions becomes a misnomer when one

5.4 Boron tant ratio with chlorinity in sceanic waters. The behave studies has been studied detailed by several workers and the result

Boron is one of the most important elements, which is significantly involved in the marine biogeochemical cycle. Boron, a metalloid, occurs in more than 100 minerals, the most common being tourmalins, a complex silicate mineral present in igneous rocks and sedimentary rocks. Boron in this form is largely inert and is released in to the environment at an extremely slow rate through natural weathering processes. The building block for the borates is BO₃ triangles linked together with Na or Ca ions. Natural weathering releases approximately 360 X 10³ metric tons of elemental boron worldwide every year (Westall et al., 1980). Although the amount of boron mobilised annually from anthropogenic sources has not been accurately determined, it is widely assumed that the most important source to surface waters are sewage, sewage sludge, and user industries such as detergent manufactures.

About 76 % of the total boron are present as boric acid, B (OH)₃, about 13% as borate, B(OH)₄ and nearly 11% is complexed with cations (Na, Mg, Ca, Co, Ni, Pb etc.)

(Robert R. Bryne, (Jr) et al., 1974). Polysaccharides and high molecular weight polyhydroxy compounds complex with boric acid to form macromolecular structure (Deuel & Nuekom., 1949).

Boron is one of the major elements that have long been regarded as characterising the seawater and marine sediments. Soon after the fundamental work by Goldschmidt and Peters (1932) on the geochemistry of boron, an attempt was made to classify using the boron content whether a sediment had been deposited in marine or lacustrine environment (saline or fresh water environment). Boron was used as a geochemical indicator of salinity and temperature of the depositional medium and there will be relation between the boron content in the water and boron content in the deposited (Harder, 1970).

The concept of constantancy of major ions becomes a misnomer when one considers an estuarine environment. The rapid response of the estuarine environment to the time dependent physico-chemical parameters such as land runoff, precipitation and evaporation, brings about changes in the behavior of most of the elements of these waters (Shirodkar & Anand., 1985; Rajagopal et al., 1981; Liss & Pointon., 1973; Narvelkar., 1980;). Exchange of solid solution components under varying chlorinity conditions in estuaries often result in nonconservative behavior of several major ions which normally bear a constant ratio with chlorinity in oceanic waters. The behavior of boron in estuaries has been studied detailed by several workers and the results indicated that some estuaries it was conservative (Liddicoat et al., 1983) while in many estuaries boron showed removal and addition processes (Shirodkar & Anand., 1985; Zingde et al., 1987; Rajagopal et al., 1981; Liss & Pointon., 1973;

Although, boron is an essential micro nutrient, low levels of born in phytoplankton indicate that the uptake of boron may not be the primary factor responsible for lower boron levels at the surface. The global average of boron and chlorinity in river water is 0.013 mg/l and 0.01 respectively (Livingston, 1963). This gives a B/Cl of 1.3. The ratio observed for surface water however is not indicative of any increase due to riverine influence.

evaporation loss (Sevenko, 1977). In the south-western Indian Ocean region of the

The interesting features that can be seen from the depth profiles are two maxima for the boron and the B/Cl between 50 and 200m, and 300 and 500 m. Chlorinity also shows a maxima in these regions. The first maxima are at about the thermal

discontinuity layer and appear to be the result of accumulation of colloidal and particulate matter. This may also be influence by the Gulf water. The second maximum centered around 400m which lies in the region of maximum apparent oxygen utilisation (AOU) (Naqvi, 1978) and is possible due to combined effect of the higher boron in the oxygen minimum zone and the high boron level associated with the Red sea water flowing at its characteristic density level.

A fairly good correlation coefficient of -0.73 observed between the suspended solids and PAR (percentage addition or removal) in Purna estuary suggests the importance of suspended load in controlling the levels of dissolved boron. A more or less inverse relationship between PAR of boron and suspended solids is observed in Ambika estuary. Good correlation between PAR of boron and chlorinity in Purna and Auranga estuaries and similar pattern of boron removal at low chlorinities suggests that the ionic strength also play an important role in the behavior of boron in these estuaries. High and low B/CI ratios with the highest ratio of 0.269 and the lowest 0.202 observed in the coastal waters of eastern Arabian sea (Shirodkar, et al., 1992) indicate addition as well as removal of boron along the coast. The percentage addition and removal of boron calculated by taking in to consideration the world average B/CI ratio for sea water (Uppstrom, 1974) 0.232, gave an addition from 0.43 to 16% and removal from 0.43 to 11.6% in the coastal waters.

Boron content showed a high correlation with chlorinity in the Central West Coast of India (Shirodkar, 1982) at various depth stations, high average variations of B/CI was reported at oxygen minima. In general lower values of boron in the upper layers of shallow stations show that boron is utilised by phytoplankton and also it may be due to evaporation loss (Sevenko, 1977). In the south-western Indian Ocean region of the southern ocean (Sugandhini Naik et al., 1986). The average concentration of boron was 4.62 ± 0.44 mg/kg and the B/CI ratio is 0.242 ± 0.023 . The relation B (mg/kg)= 0.227 X CI ± 0.050 with the observed mean chlorinity in this case as 19.124 gives boron concentration as 4.39 mg/kg and the corresponding B/CI as 0.230. The ratio, as calculated, is somewhat as higher than that obtained from low chlorinity Baltic Sea water byDyrssen and Uppstrom (1974), which was 0.230. Culkin (1965) has reported that the average concentration of boron to the 4.5 mg/kg at a salinity of 35%.

It is well known that marine clay sediments are strongly enriched in boron (100 ppm), as they, together with altered oceanic crust, are the earth's most significant boron reservoirs (Goldschmidt, 1954; Thomson et al., 1970; Harder, 1974).

The water temperature, too, has an effect on the boron content (Harder, 1959b, 1961b). Sometimes the organic compounds influence the boron content of clays. Eager (1962), Curtis (1964) and Harder, (1970) have shown an inverse relation ship between boron content and organic carbon. It appears that the presence of organic matter and relatively low contents of total boron in these sediments (perhaps not in all) are related (Eager, 1962).

In an aquatic system, sediments store by removing it from the overlying water through the physical process of adsorption and chemical complexation. Boron from the overlying water is removed by the differential adsorption on to suspended solids during the estuarine mixing of sea and river waters (Levinson and Luddwick, 1976; Liss and Pointon, 1973). Biological removal of boron by phytoplankton have also been observed which adds boron back to the sediments on the death and decay of phytoplankton (Subba Rao, 1981; Shirodkar et al., 1982).

Acid treatment was carried out for two calcareous sediments, to examine the influence of biogenic carbonates on the bulk content and isotopic composition of boron in these sediments. The sediments contain 58% and 36% of HCl soluble component (carbonate), and the residues corresponding to pelagic clay and diatom ooze respectively. Boron content of pelagic clays determined in this study (Tsuyoshi Ishikawa & Nakamura., 1993.) vary from 96 to 132 ppm, which is essentially consistent with the range of 80-157 ppm given in Spivack et al., 1987 and 120 ppm for average argillaceous sediments (Harder, 1974).

5.5 Boron in CES - Results of the present study and administration of the present study at the pr

Dissolved boron in the surface water of Cochin estuary varied between ND level to 3.07 mg/l (Annexure.5a). Boron in the northern zone of the estuary varied from ND level to 2.36 mg/l and in the southern zone 0.16 to 3.07 mg/l, the maximum value was showed in the Barmouth. Boron in the bottom water has slightly higher concentration than surface water. In the bottom water dissolved boron varies from ND level to 5.91

mg/l in the north zone and the maximum value was showed in station.1. In the south zone, dissolved boron concentration varied from 0.31 to 3.58mg/l.

Dissolved boron generally showed an increasing trend in surface waters during the three seasons from upstream to downstream except in station 1 during premonsoon and monsoon (fig. 5.1 a & Table 5.1). Station 1 showed high values during premonsoon and monsoon season compared to station 2, this may be due to the input from anthropogenic sources.

Table 5.1. Seasonal average of dissolved boron in water (ppm).

Station	1	2	3	4	5	6	7
Postmonsoon. S	0.38	0.69	0.95	1.79	1.92	1.23	0.49
В	2.16	0.75	1.05	2.11	2.80	1.14	1.32
Premonsoon. S	0.29	0.15	0.38	1.09	1.81	0.87	0.67
В	0.39	0.48	0.94	1.61	2.22	1.14	0.79
Monsoon. S	0.70	0.27	0.29	0.69	0.38	0.98	0.52
B	0.59	0.15	0.32	0.33	2.27	0.88	0.79

S - surface; B -bottom

The bottom water also showed a similar trend as that of surface water monsoon (fig: 5.1 b) in all stations except station 1 and station 7 during postmonsoon. In postmonsoon and monsoon, station 1 have higher values than station 2 and also in the southern zone station 7 showed higher values than station 6 in postmonsoon season. Generally boron in the surface and bottom waters of Cochin estuary increases with salinity, clearly indicating the introduction of boron in to the CES through saline water.

Water-soluble boron in the estuarine sediments of Cochin varied from 4.5 to 61.18 ppm in the northern zone and in the southern zone, the value varied from 2.43 to 103.42 ppm (Annexure 5 b). Water-soluble boron in the sediment showed maximum values in station 2 during premonsoon monsoon (fig: 5.1 c & Table 5.2). There was no significant seasonal trend for water-soluble boron in the Cochin estuarine sediment.

Table 5.2. Seasonal average of water soluble boron in sediment (ppm).

Station	1	2	3	4	5	6	7
Postmonsoon	13.11	14.59	12.16	23.01	16.02	12.94	16.74
Premonsoon	13.94	56.91	17.26	22.05	20.18	22.97	15.25
Monsoon	26.45	11.56	19.28	16.75	14.08	53.26	30.46



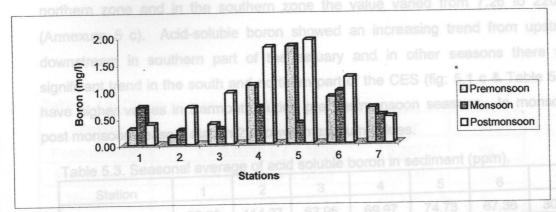


Fig: 5.1 a

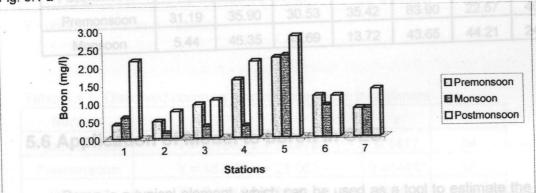


Fig: 5.1 b

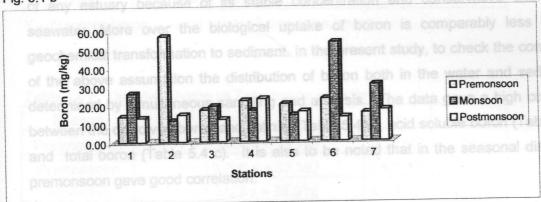


Fig: 5.1 c

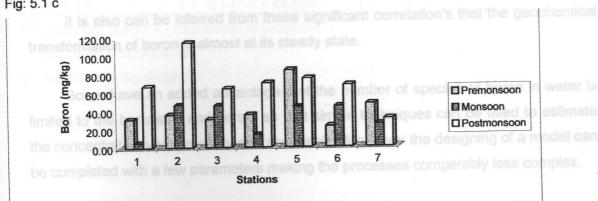


Fig: 5.1 d

Acid-soluble boron in the sediment varied from 5.42 to 191.54 ppm in the northern zone and in the southern zone the value varied from 7.26 to 220.5 ppm (Annexure 5 c). Acid-soluble boron showed an increasing trend from upstream to downstream in southern part of the estuary and in other seasons there was no significant trend in the south and northern part of the CES (fig: 5.1 c & Table 5.3), and have higher values in Barmouth during pre postmonsoon seasons. In monsoon and post monsoon season, station 2 showed maximum values.

Table 5.3. Seasonal average of acid soluble boron in sediment (ppm).

able 5.3. Seaso	1	2	3	4	5	6	7
Station	00.00	114.27	63.95	69.97	74.73	67.36	30.55
Postmonsoon	66.63	114.21	00.00	10.40	9444	00.57	46.72
Premonsoon	31.19	35.90	30.53	35.42	83.90	22.57	40.72
Premonsoon	E SUREY	10.05	45.00	13.72	43.65	44.21	24.53
Monsoon	5.44	46.35	45.69	13.12	40.00		

5.6 Application of Model to boron in CES.

Boron is a typical element, which can be used as a tool to estimate the reactivity of any estuary because of its stable concentration and conservative character in seawater. More over the biological uptake of boron is comparably less than its geochemical transformation to sediment. In the present study, to check the correctness of the above assumption the distribution of boron both in the water and sediment is determined by simultaneous sampling and analysis. The data gave a high correlation between the dissolved boron and salinity (Table 5.4 a), acid soluble boron (Table 5.4 b) and total boron (Table 5.4 c). It is also to be noted that in the seasonal distribution premonsoon gave good correlation.

It is also can be inferred from these significant correlation's that the geochemical transformation of boron is almost at its steady state.

Boron have an added advantage that the number of species of boron in water is limited to the boric acid and borate so that simple techniques can be used to estimate the concentration very precisely and accurately. Moreover the designing of a model can be completed with a few parameters making the processes comparably less complex.

Table 5 4 a Dissolved Boron vs Salinity

Season	and the management of the state	r	n
Postmonsoon	Y = 0.0339 x + 1.1038	0.28778*	68
Premonsoon	Y = 0.059 x + 0.0845	0.85407#	56
Monsoon	Y = 0.0617 x + 0.5395	0.53025#	40
Total	$Y = 0.0403 \times + 0.07$	0.43944#	164

the concentration levels generally observed makes boron the most suitable element for

southern arm of the CES shows considerable variations in almost all of the estuarine

Table 5.4 b. Dissolved boron vs Acid-soluble boron in sediment

Season	hip obtained and the correlation	on coefficient in	n
Postmonsoon	Y = 6.7063 x + 58.978	0.1417	34
Premonsoon	Y = 18.352 x + 21.063	0.41405*	28
Monsoon	Y = 7.8144 x + 29.928	0.20526	20
Total	Y = 12.906 x + 35. 984	0.284117#	82

Y - dissolved boron, x - acid soluble boron in sediment

Table 5.4 c. Dissolved boron vs Total botron in sediment

Season	or studied by theiry workers to	r	n
Postmonsoon	Y = 2.1454 x + 15.141	0.25242	34
Premonsoon	Y = 19.206 x + 37.392	0.40132*	28
Monsoon	Y= 7.9327 x + 54.919	0.13786	20
Total	Y = -5.147 x + 44.037	0.17042	79

Y - dissolved boron, x - total boron in sediment rved concentration of any species of any element will have

* - above 95% significance

- above 99% significance the discharge. The distribution studies of various chemical species done by earlier workers also confirm this (Lakshmanan et al., 1982; As the borate to boric acid ratio is part of an equilibrium coefficient, a definite relation between the total boron concentration and any one of the individual species will be present. In short, the reactivity, estimation methodology, the specious diversity and the concentration levels generally observed makes boron the most suitable element for modeling the reactivity of estuaries.

In this study, the observed concentration of boron is substituted to the flow reactor model. In the model C is replaced by the observed boron concentration and salinity (S) of that particular water mass. As the estuarine conditions during the three seasons, monsoon, premonsoon and postmonsoon are significantly different, the reactivity K is calculated independently for the three seasons. Also, the northern arm and the southern arm of the CES shows considerable variations in almost all of the estuarine properties, making it essential to apply the model separately to the two arms.

The relationship obtained and the correlation coefficient from regression analysis is given in table. 5. 5.

The analysis of the results obtained by substitution of data in the model indicate that

- 1) Except for monsoon season, the log C (Boron) exhibits highly significant correlation with salinity. This confirms the assumptions made during the development of the model. The dilution and flushing characteristics of the Barmouth of CES (coastal inlet) have been studied by many workers (Joseph and Kurup, 1989; Ajith Joseph, 1996). The major observations in these studies were that during the monsoon season, the Cochin inlet is subjected to considerable dilution. The monsoon season is characteristic for its (1) high flooding of the estuary with fresh water, (2) very low seawater intrusion and (3) very low flushing time. The geochemical reactivity as a result of the above reasons the estuary, during monsoon season will generally be very low and the observed concentration of any species of any element will have bearing only to the riverine discharge. The distribution studies of various chemical species done by earlier workers also confirm this (Lakshmanan et al., 1982; Anirudhan, 1988; Babukkutty, 1991; Shibu, 1992; Vasudevan Nayar, 1992).
- 2) The order of reactivity from the K values are Monsoon> Postmonsoon> Premonsoon. As the flushing time is minimum during the monsoon season and as the fresh water inflow is considerably large blocking the seawater intrusion, the monsoon season will show the maximum K value. The absence of a significant

Table 5.5. Model parameters and correlation coefficient. that the geochemical transformations are not the reason for

Lag Bye Salinity

North	es. This can only be attributed to	K	r chemical ci	n
Postmonsoon	Log C = 0.181025 S - 1.021343	0.18103	0.354821*	34
Premonsoon	Log C = 0.058261S - 0.883477	0.05826	0.75147#	29
Monsoon	Log C = -7.46826 S - 5.846901	-7.46826	0.100168	22
Total	Log C = 0.129865 S - 1.026544	0.12987	0.435539#	89
South	procept of geochemical reactivity w	as not att	ibuted to the	se
Postmonsoon	Log C = 0.052425 S - 0.710616	0.05243	0.699223#	30
Premonsoon	Log C = 0.054386 S - 1.134497	0.05439	0.820862#	23
Monsoon	Log C = -0.116861 S - 0.492112	-0.11686	0.044112	15
Total	Log C = 0.053914 S - 0.766983	0.05391	0.627226#	69

C - Boron, S - Salinity

Log MRAS vs Salinity

North		K	r	n
Postmonsoon	Log C = -7.733952 S + 46.22119	-7.73395	0.00263	38
Premonsoon	Log C = 0.079815 S + 0.356413	0.07982	0.449409*	31
Monsoon	Log C = -5.521811 S + 2.029834	-5.52188	0.329979	22
Total	Log C= 0.493925 S - 1.508051	0.49393	0.067384	93
South				
Postmonsoon	Log C = 0.044282 S + 0.632511	0.04428	0.463332*	30
Premonsoon	Log C = 0.177233 S - 2.363043	0.17723	0.15552	23
Monsoon	Log C = 37.45318 S -42.6779	37.4532	0.225078	15
Total	Log C = 0.136258 S - 0.56832	0.13626	0.139423	69

C - MBAS, S - Salinity

Log phenol vs Salinity

og phenol vs Sa North	a penod as a whole. The monsoo	K	r	n
Postmonsoon	Log C = -0.12006 S + 1.760073	-0.12201	0.328424#	40
Premonsoon	Log C = 0.268722 S - 1.905521	0.26872	0.241558	31
Monsoon	Log C = -10.2145 S + 2.720123	-10.2145	0.081104	22
Total	Log C = -0.326904 + 8.8666	-0.3269	0.191943	96
South	tion time because of the low flushin	dunng th	a premonso.	an, cea
Postmonsoon	Log C = -0.06367 S + 1.861709	-0.06367	0.47172#	25
Premonsoon	Log C = 0.137355 S - 1.870639	0.13736	0.233575	22
Monsoon	Log C = 0.008179 S + 11.811	0.00818	0.245472	17
Total	Log C = -0.79321 S + 2.057349	-0.79321	0.30748#	64

C- Phenol, S - Salinity and all the literature (Polisois set

^{* -} above 95% significance, # - above 99% significance

correlation but indicates that the geochemical transformations are not the reason for these high K values. This can only be attributed to the irregular chemical character of the floodwater of monsoon.

- 3) In the postmonsoon, though have a reasonably low flushing time; the estuary gets sufficient time to reach the steady state. The reaction between the reactivity and the flushing time is more evident when we compare the postmonsoon with premonsoon. Though the concept of geochemical reactivity was not attributed to the seasonal variations in the distribution of various chemical species, such a relation between the postmonsoon and premonsoon was observed earlier also (Lakshmanan et al., 1982; Anirudhan, 1988; Babukkutty, 1991; Shibu, 1992).
- 4) During the monsoon season, both the northern and southern regions exhibit more or less the same reactivity with respect to boron. The trend graphs give the same slope for both the regions. The prevailing of fresh water conditions, low salinity and fast flushing may be the reasons.
- 5) The reactivity coefficient K is greater in the northern region than the southern. The lower flow rate of river water and shallow nature of the water column may be the major contributors.

5.7 Application of the model to MBAS and Phenol.

The proposed model is applied to MBAS and phenol and the relation and correlation coefficients are given in Table. 5.5. In the case of phenol a significant correlation between log C(Phenol) and salinity was observed only for the postmonsoon season and the total period as a whole. The monsoon, as in the case of boron, showed the highest reactivity that can be attributed to the increased flushing (the decreased flushing time) and also to the insignificant addition of the seawater. The premonsoon, because of the higher flushing time, stagnant character of the water body and the high organic content shows only low geochemical reactivity as evidenced by a low value for K. The high retention time because of the low flushing during the premonsoon can lead to reactions other than geochemical such as photochemical reactions. The absence of any significant correlation between Log C and salinity during the premonsoon may be due to such side reactions. Also the insitu biochemical additions of phenols can Evidences for the photochemical contribute to the phenol concentration. transformations of phenols in aquatic systems are available in the literature (Polisois et -al., 1975; Stephen Opsal & Benner.,1998).

The absence of significant correlation between log C (MBAS) with salinity indicates that MBAS is totally out of the model. The reason can be (1) the major source of MBAS in the aquatic system is anthropogenic so that the concentration of MBAS will have bearing only to the fresh water rather than the seawater. The basic structure of the geochemical reactivity model is the modification of chemical species brought to the estuarine waters from the seawater. The flow reactor considers the fresh water contribution to the concentration of chemical species as negligible or at a steady state.

(2) MBAS belongs to a group of chemicals that are highly reactive, bioactive and biodegradable. The geochemical reactivity concept cannot be so applicable here; instead a model which can accommodate the above factors also to developed to explain the reactivity of MBAS. This requires more data on the reaction side of MBAS, which was beyond the scope of the present work. Work in this direction has already been initiated in this laboratory.

very difficult. Many pollutants adsorb on to suspended particulate material and get locked in the sediments on settling of these materials. Sedimentation of a pollutant may lead to serious long-term problems in an area, as resuspension of sediments and exchange with overlying water are regular process; in estuarine dynamics.

Surface active agents are substances which cause lowering of the surface tension of liquids, particularly water. They can influence the structure and physico chemical properties of natural interfaces and mediate the processes of mass and energy transfer between different phases. They are involved in the processes such as bubble flotation of particles, microlayer enrichment and other surface effects. They are able to control the transfer of gases between the air-water interface

Surfactants (surface active agents and detergents) are very widely used as industrial and household cleaning agents, dispersants and emulsifiers. The number of these compounds is very large and it is continually increasing, although only relatively few types are in extensive use. Synthetic detergents have caused much concern due to their tendency, even in small amounts, to cause foam in rivers and at sewage plants. Harmful effects of detergents in natural waters may result from their general types on the biogeochemical cycle of other pollutants and biogenic elements.

Natural surfactants from a major part of the (estimated about 50-70%) marine humic substances. The fulvic acid fraction, one of the most important soluble

SUMMARY And include entonic surfactants.

Estuaries are unique environments that play an important role in the transfer of products of continental weathering to the ocean. Because of the major physicochemical conditions existing in fresh and saline water, the distribution and chemical characteristics of constituents in dissolved and particulate forms get modified during the mixing processes in estuaries. Hence, assessment of the quality of water in an estuary is of great significance in the maintenance of a sustained ecosystem.

A wide spectrum of organic and inorganic compounds enters the estuary through different sources such as air, river and direct or indirect anthropogenic discharges etc. Since the subsequent behavior and fate of the substances depend on a variety of factors - physical, biological and chemical - the prediction of their behavior and fate is very difficult. Many pollutants adsorb on to suspended particulate material and get locked in the sediments on settling of these materials. Sedimentation of a pollutant may lead to serious long-term problems in an area, as resuspension of sediments and exchange with overlying water are regular process in estuarine dynamics.

Surface active agents are substances which cause lowering of the surface tension of liquids, particularly water. They can influence the structure and physico chemical properties of natural interfaces and mediate the processes of mass and energy transfer between different phases. They are involved in the processes such as bubble flotation of particles, microlayer enrichment and other surface effects. They are able to control the transfer of gases between the air-water interface

Surfactants (surface active agents and detergents) are very widely used as industrial and household cleaning agents, dispersants and emulsifiers. The number of these compounds is very large and it is continually increasing, although only relatively few types are in extensive use. Synthetic detergents have caused much concern due to their tendency, even in small amounts, to cause foam in rivers and at sewage plants. Harmful effects of detergents in natural waters may result from their general impact on the biogeochemical cycle of other pollutants and biogenic elements.

Natural surfactants from a major part of the (estimated about 50-70%) marine humic substances. The fulvic acid fraction, one of the most important soluble component involved in the marine aerosol process, show the characteristics of wet

surfactants. Coastal and estuarine waters in some densely populated areas receive significant quantities of various derived chemicals, which include anionic surfactants. These environments seem to be particularly sensitive to pollution with synthetic surfactants.

The CES is highly influenced by various anthropogenic activities like discharge of agricultural, industrial and urban wastes, operation of shipyard, oil and other transporting activities, fishing, dredging etc. A by-product of the dredging process is the resuspension of the bottom sediment, which contains stored toxic and other compounds. In the present study, the distribution and seasonal variation of anionic surfactants (MBAS), total phenols, chlorophenols and boron in the water and sediments of the CES has been studied

Seasonal values of MBAS showed high values in the surface water during monsoon compared to premonsoon and postmonsoon. In the surface water, MBAS showed a decreasing trend from upstream to downstream in the north zone during postmonsoon and in the bottom water an increasing trend towards downstream in premonsoon and postmonsoon in the southern part of the estuary. No significant trend was observed in the other seasons.

MBAS in the sediment showed an increasing trend towards downstream in the north zone. In the south zone a decreasing trend was observed during premonsoon.

MBAS in the sediments of CES showed significantly high correlation with sedimentary organic carbon.

The seasonal values of phenolic compounds in the surface waters of Cochin estuary showed higher values in the northern part during monsoon and in the southern part during postmonsoon. In all the seasons, the average value of phenolic compounds in the surface waters of Cochin estuary along the Periyar River showed an increasing trend towards the upstream. This can be attributed to the input of phenolic compounds in to the estuary from anthropogenic or natural sources. The southern part of the estuary also showed a similar trend as that of the north zone, where phenolic concentration showed high values in the upstream station (Station no.7). Highest value for phenolic compounds during the monsoon months in the northern part of the estuary was in June 96 and in the southern part was in August 96. In the bottom water also, phenolic concentration showed an increasing trend towards upstream.

Postmonsoon in the northern part and premonsoon in the southern part showed to be exceptions to this. In postmonsoon months, very high values of phenolic concentration were observed during October 96 in all the stations. In premonsoon, May 96 showed maximum values in stations except barmouth. In monsoon season, all stations showed higher values except barmouth in all months. Localised high values were observed for phenols because of anthropogenic activities such as retting of coir, industrial discharge etc.

Phenolic compounds in the sediment during monsoon showed an increasing trend towards the downstream and in other seasons there was no specific trend and in the southern area station 7 showed maximum value in all seasons compared to station 6. The concentration of phenols in the sediment was always very low in stataion 1, except in November 96.

Chlorophenolic compounds are toxic to aquatic species, but to varying degrees depending on the number and the position of the chlorine substituents on the benzene ring. The chlorinated phenols consist of a group of 19 different isomers which include mono, di, tri, tetra and one pentachlorophenol. All these compounds are toxic to aquatic species, but in varying degrees with PCP being the most toxic. They are brought in to the environment, intentionally or unintentionally, through anthropogenic activities. The release of industrially derived halogenated organic compounds in to the aquatic environment is of great concern, mainly because of their toxicity, resistance to degradation, and tendency to bioaccumulate. Chlorophenolic compounds have been found to be the major constituents produced from lignin residues. Some of them are toxic and can get accumulated in living organisms.

The identification of the different chlorophenols in CES could be done with o, p and m chlorophenols because of the non availability of pure standards. The rest of the species observed in the G.C. is represented as fraction 1, 2 etc. The trends given denote decrease or increase, as the case may, towards downstream.

In the Cochin estuary o-chlorophenol and p-chlorophenol showed low values in the surface water compared to bottom water in the northern part of the estuary and higher values in the surface water in the southern part. There was no significant trend for o-chlorophenol in the estuarine water. Seasonal average of o-chlorophenol showed high values during monsoon seasons in the surface and bottom water. In the

sediment samples o-chlorophenol showed a decreasing trend during monsoon season and an increasing trend in the postmonsoon and premonsoon season in the northern part. No significant trend was observed in the southern part. In the sediment, p-chlorophenol showed a decreasing trend during monsoon in the northern part and in other seasons there was no significant trend in the north and southern part.

m-chlorophenol showed only a decreasing trend in the surface water during premonsoon and in other seasons the water and sediment samples had no significant trend in the study area. In water samples m-chlorophenol showed higher values during monsoon season in the surface and bottom water, but in sediment samples it showed low values during monsoon except in station.1. Generally m-chlorophenol showed higher values in the bottom water than surface water.

Fraction 1 showed a decreasing trend in bottom water in the northern part of the estuary during monsoon. The surface water of the south zone also showed a similar trend during monsoon and in other seasons the surface and bottom water had no significant trend. The sediment showed an increasing trend during monsoon in the northern and southern part of the study area.

Fraction 2 showed an increasing trend in the southern part during the premonsoon and a decreasing trend in postmonsoon seasons in the bottom water. The surface water and sediment samples had no significant trend in the two zones was observed.

Fraction 3 showed an increasing trend in the surface water of northern part during monsoon and postmonsoon. The bottom water also showed a similar trend during premonsoon of southern part. The sediment concentration showed no significant station wise trend. High values were observed in station.1 during monsoon.

Except for the increase in concentration during premonsoon of the southern part and a decrease during postmonsoon in the surface water of the northern part, no general trend was shown by fraction 4. The surface water concentration generally showed high values during premonsoon compared to bottom water where as in monsoon season the bottom water showed higher values. The sediment also showed a decreasing trend during premonsoon and monsoon in the southern part and monsoon in the northern part of the estuary. But in postmonsoon season, the southern part showed an increasing trend.

premonsoon season in the northern part of the estuary except in station 1. Surface water of the northern part during premonsoon and postmonsoon showed an increasing trend. The southern part also showed an increasing trend during postmonsoon season. In the bottom water premonsoon showed an increasing trend in the north and in postmonsoon an increasing trend was showed in the southern part. The sediment in the north showed a decreasing trend during premonsoon and an increasing trend in postmonsoon. An increasing trend was observed in the south part during monsoon and a decreasing trend in premonsoon.

Fraction 6 showed generally higher values in the bottom water than surface in the northern part of the estuary where as in the southern part, surface water gave higher values except station 5. The surface water has showed an increasing trend in premonsoon and a decreasing trend in monsoon in the northern part. A decreasing trend was the observed in the bottom waters of the northern part in all seasons except station 4. In the northern part the sediment showed minimum during monsoon and maximum in postmonsoon season. In the sediment, the premonsoon values showed a decreasing trend in the northern and southern part of the CES and an increasing trend in southern part during postmonsoon.

Fraction 7 in the surface water showed low values during monsoon and in the sediment during postmonsoon. A decreasing trend was observed during postmonsoon and monsoon surface water in the southern part where as an increasing trend in the north during postmonsoon in the bottom water. Sediment samples showed an increasing trend in the southern part of the estuary during monsoon and in the north a decreasing trend during monsoon and postmonsoon.

A Kinetic flow reactor model is proposed to explain the overall reactivity of the estuary. The distribution data of boron was used to test the model. It was observed from the model that the order of reactivity was found to be Monsoon> Postmonsoon> Premonsoon. The absence of significant correlation between the model parameters in the monsoon season and the observed reactivity during monsoon was attributed to the irregular characteristics of the floodwater. The northern part was found to be more reactive than the southern part, may be due to the lower flow rate of river water and the shallow nature of the water column in the southern part. The model was found to explain the distribution pattern of phenol but MBAS was totally out of the model.

References

- Adams, T.D., Haynes, J.R. and Walker, C.T., 1964. Boron in Halocene illites of the dovey estuary, Wales and the its relationship to paleosalinity in cyclotherms. Sedimentology, 4, 189-195.
- Ajith Joseph, K., 1996. Straight dynamics of tropical tidal inlet. Ph.D. Thesis, Cochin University of Science and Technology, Cochin.
- Alagarsamy, R., 1991. Organic carbon in the sediments of Mandovi estuary, Goa. *Indian J. Mar. Sci.*, Vol. **20**, 221-222.
- Aldelman, I.R., Smith (Jr), L.L. and Siesenop, G.D., 1976. Accute toxicity of Sodium Chloride, Pentachlorophenol, gluthion and hexavalent chromium to fathead minnow and Gold Fish. *J.Fish.Res.Bd.Can.*, **33**, 203-208.
- Anirudhan, T.S., 1988. Studies on the nutrient chemistry of a tropical estuary. Ph.D. Thesis, Cochin University of Science and Technology, Cochin.
- APHA, 1995. Standard methods for the examination of water and wastewater. (Edited by Andrew D. Eaten, Lanore S. Clescori and Arnold E.Greenberg), 19 Ed. APHA, AWWA, WPCF, Washington, D.C.
- Argese, E., Bettiol, C., Todeschini, R. and Miana, P., 1995. Submitochondrial particles as toxicity biosensors of chlorophenols. *Environ. Toxicol. Chem.*, **14**, 363-368.
- Arsenaut, R.D., 1976. Pentachlorophenol and contaminated chlorinated dibenzodioxins in the environment: a study of environmental fate, stability, and significance when used in wood preservation. Presented at the *American Wood Preservers Association Annual Meeting*, Atlanta, Georgia, April 25-28, pp 122-148.
- Ayfer Yediler, Yong Zhang, Jun Peg Cai and Friedhelm Korte, 1989. Effect on the microbial population on the degradation of alkylbenzene sulfonate in Lake Waters. *Chemosphere*, **18** (7-8), 1589-1597.
- Babich, H. and Davis, D.L., 1981. Phenol: A review of environmental and health risks.

 *Reg. Toxicol. Pharmacol. 1, 90-109.
- Baeteman, M. and Vyncke, W., 1979. Phenolic compounds in the water and marine Organisms off the Belgian coast. *Medelingen van het Rijkstation voor Zeevisseri* (CL.O.Gent), pp 1-7. Pub. No. 157.
- Babukkutty, Y., 1991. Studies on the inter-compartmental exchange of trace metals in an estuarine system. Ph.D.Thesis, Cochin University of Science and Technology, Cochin.

- Barr, D.P. and Aust, S.D., 1994. Mechanisms white rot fungi use to degrade pollutants. *Envir. Sci. and Technol.*, **28**(2), 78A-87A.
- Beenamma Jacob, 1993. Studies on the Sulphur Chemistry of a tropical Estuarine system. Ph.D. Thesis, Cochin University of Science and Technology, Cochin.
- Berry, D.F. and Boyd, S.A., 1985a. Reaction rate of phenolic humus constituents and anilines during cross coupling. *Soil Biol. Biochem.* **17**(5), 631-636.
- Berry, D.F. and Boyd, S.A., 1985b. Decontamination of soil through enhanced formation of bound residues. *Envir. Sci. and Technol.*, **19**(11), 1132-1133.
- Bevenue, A., Ogata, J.N. and Hylin, J.W., 1972. Organochlorine pesticides in rain water, Oahu, Hawai, 1971-1972. Bulletin of Environmental Contamination and Toxicology, 8: 238-241.
- Blanchard, 1975. Bubble scavenging and the water to air transfer of organic material. In: *The Sea*. Advances in chemistry series **145**, 360-387.
- Bode, H., Ernst, R. and Arditti, J., 1978. Biological effects of surfactants. 111. Hydra as a highly sensitive animal. *Environmental Pollut.* 17, 175.
- Bollang, J.M., Liu, S.Y. and Minard, R.D., 1980. Cross coupling of phenolic humus constituents and 2,4- dichlorophenol. J. Soil Sci. Soc. Am., 44, 52-56.
- Bostrom, S.L. and Johansson, R.G., 1972. Effects of Pentachlorophenol on enzymes involved in energy metabolism in the liver of the eel. *Comp Biochem Physiol.*, **41**b: 359-369.
- Brown, V.M., Abram, F.S.H. and Collins, L.J., 1978. The acute lethal toxicity to rainbow trout of a LAS surfactant and of its residues and degraded products. *Tenside Dtergents*, **15**, 57-59.
- Bryant, S. E. and Schultz, T.W., 1994. Toxicological assessment of biotransformation products of pentachlorophenol; Tetrahymena population growth impairment. *Arch. Envirn. Contam. Toxicol.*, **26**, 299-303.

- Buikemia, A.L, Jr., Mc Ginniss, M.J. and Cairns, J.J. (Jr)., 1979. Phenolics in aquatic ecosystem: a selected review of recent literature. *Mar. Environ. Res*, **2**: 87-181.
- Burges, A., 1967. The decomposition of Organic matter in the soil. In: Soil Biology, edited by A. Burges and E. Raw, Academic Press, London, 15-47.
- Call, D.J., Brooke, L.T. and Lu.P., 1980. Uptake elimination and metabolism of three phenols by fathead minnows. *Archives of Environmental contamination and Technology.*, **9**, 699-714.
- Cammeron, W.N. and Pritchard, D.W., 1963. In: "The Sea" (M.N.Hill, ed.) Vol.2, pp 306-323. Interscience Publishers, London, New York.
- Carlberg, G.E. and Nashaug, O., 1986. Environmental impact of organochlorine compounds discharged from the pulp and paper industry. In: EUCEPA Symp. Environmental protection in 90's, Helsinki, Finland, pp 50-54.
- Cauwet, G., 1978. Organic chemistry of seawater particulate. Concepts and developments. *Oceanol Acta* 1 (1); 99-105.
- Christman, R.F. and Ghassemi, M., 1966. J.Amm.Water Works Assoc. 58, 723.
- Christoffer Rappe., 1984. Analysis of polychlorinated dioxins and furans. *Environ.* Sci. Technol., Vol.18, No.3, pp 78A-90A.
- Cini, R., Degli Innocenti, N., Loglio, G., Stortini, A.M. and Tesei, U., 1994 Spectrofluorometric evidence of the transport of marine organic matter in Antarctic snow via air-sea interaction. *International J. ournal of Environmental Anal. Chem.* **55**, 285-295.
- Connel, D.W., 1988. Bioaccumulation behavior of persistent organic chemicals with aquatic organisms. Reviews of Environmental Contamination and Toxicology. **101**, 117-154.
- Cookson, J.T (Jr.)., 1995. Bioremedation engineering: design and application.

 McGraw-Hill Book Co., Inc, New York, N.Y.
- Cosovic, B. and Vojodic, V., 1989. Adsorption behaviour of the hydrophobic fraction of organic matter in natural waters. *Mar. Chem.*, 28, 183-198.
- Cosovic, B and Vojovodic, V., 1982. The application of as polarography to the determination of surface active substances in sea water. *Limnol. Oceanogr*, **27** (92), 361-369.
- Cosovic, B., Zutic, V., Kozarac, Z., Vojvodic, V. and Novakovic, T., 1979. Investigation of surfactants in north Adriatic in the period of 1974 to 1978. Evaluations of natural variations and pollution effects. *Rapp. Comm. Int. mer. Medit.* **25/26** (9), 55-60.
- Crawford, R.L., and Mohn, W.W., 1985. Microbial removal of Pentachlorophenol from

- soil using Flavobacterium. Enzyme Microb. Technol., 7(12), 617-620.
- Crawthorne, B., Fielding, M., Steele, C. and Watts, C., 1984. Organic compounds in water. Analysis using coupled-column high-performance liquid chromatography and soft ionisation mass spectrometry. *Environ. Sci. Technol.* **18**, 797-802.
- Culkin, F., 1965. The major constituents of sea water, In: Chemical Oceanography, Vol.1, ed by J.P.Riley and G. Skirrow, pp121 161.

Chem., 53(5), 1051-1080.

- Cupuzzo, J.M., Burt, W.V., Duedall, I.W., Park, P.K. and Kester, D.R., 1985. Future strategies for nearshore waste disposal, In: *Wastes in the Ocean*, Vol. 6.

 Nearshore waste disposal, Ketchum, B.H., Capuzzo, J.M., Burt, W.V.
- Curtis, C.D., 1964. Studies on the use of boron as a paleoenvironmental indicator.

 Cosmochemita Acta, 28, 1125-1137.
- Dalla Vanezia, L., Fossato, V.U. and Scarfi, S., 1980. Combined effects of sodium dodecyl benzene sulphonate and low salinity on Tisbu bulbisetosa (Copepoda: Harpacticoida). *Progr. Wat. Technol.* 12, 109-117.
- Dec, J. and Bollag, J.M., 1990. Detoxification of substituted phenols by oxidoreductive enzymes through polymerisation reactions. *Archives Envir. Contamination and Toxicology.*, **19**, 543-550.
- DeGraeve, G.M., Geiger, D.L., Meyers, D.S. and Bergman, H.L., 1980. Acute and embryolarval toxicity of phenolic compounds to aquatic biota. *Arch. Environ. Contam. Toxicol.* **9**, 557-568
- Delfino, J.J. and Dumbbe, D.J., 1976. J. Environ. Sci. Health. A11, 345.
- Depledge, M.H. and Fossi, M.C., 1994. The role of biiomarkers in environmental assesment (2). *Invertibrates. Ecotoxicology.* **3**, 161-172.
- Desaiah, D., 1978. Effect of pentachloro phenol on the ATPase in rat tissues.

 Pentachloro Phenol, 12, 277-283.
- Doetsch, N.R. and Cook, T.M., 1973. Introduction to Bacteria and Their Ecobiology, University Park Press, Baltimore, pp 371.
- Deuel, H. and Neukom, H., 1949. Uber die Reaktion von Borsaure und Borax mit polysaccarides and andern hoc molekularen Polyoxy-Verbin-dugen. Makromol. Chem., 3, 13-30.
- Dyrssen, D.W.and Uppstrom, L.R., 1974. Boron Chlorinity ratio in Baltic seawater, *Ambio*, Vol 3, 33.
- Eager, R.M.C. and Spears, D.A., 1966. Boron content in relation to organic carbon and to paleosalinity in certain British carboniferous sediments. *Nature*, **209**, 177-181.

- Eager, R.M.C., 1962. Boron content in relation to organic carbon in certain sediments of British coal measures. *Nature* (London). **196**, 428-431.
- Eder, G. and Weber, K., 1980. Chlorinated phenols in sediments and suspended matter of the Weser estuary. Chemosphere, 9, 111.
- Eisenbud, M., 1988. Environmental Radioactivity, 4th ed., McGraw Hill, New York.
- El Wakeel, S.K. and Riley, J.P., 1957. The determination of organic croon in marine muds. *Journal du Conseil International Pour l'exploration de la Mer.* **22**, 180-183.
- Emery, K.O., 1956. Bull. Geol. Soc. Am., 72, 478.
- Environmental Protection Agency., 1973. *Priority Basin Accomplishment Report*,

 Division of water planing, Office of Air and Water Programes.
- Ericksson, K.E., Kolar, M.C., Ljungquist, P.O. and Kringstad, K.P., 1985. Studies on microbial and chemical conversions of chlorolignins. *Environmental Science and Technology*. **19**, 1219-1224.
- Ernst, 1962. Fortschr. Geol. Rheinland Westfalen, 3(3): 947-964.
- Exon, T. H., 1984. A review of chlorinated phenols. Vet. Hum. Toxicol. 26, 508-520.
- Fairbridge, R.W., 1980. The estuary; its definition and geodynamic cycle. In: Chemistry and Biogeochemistry of Estuaries. Olausson, E. and cato, I (eds.), John willey and sons, New York. pp 1-35.
- Fell, B (ed) 1989. Proceedings of the international status seminar "Alkylbenzene sulphonate (LAS)", Aachen, Nov 9th 10th, 1988. *Tenside Surfact Deterg.* **26**, 69-179.
- Fenical, W., 1975. Halogenation in the Rhodophyta: a review. *Journal of Phycology*, **11**, 245-259.
- Fetterolf, C.M., 1964. Proc. Ind. Waste Conf. Purde Univ. 115, 174.

Fisher, W.K., 1980. Entwicklung der tensidkonzentartionen in den deutschen gewassen. Tenside Deter. 17, 250-261.

- Fountaine, J.E., Joshipura, P.B. and Keliher, P.N., 1976. Some observation regarding pentachlorophenol levels in haverford township, Pennsylvania. *Water Research*, Vol.10, pp 185-185.
- Fujita, T. and Koga, S., 1976. The binding of a cationic dye detergent by yeast cells to its germicidal action. J. Gen. *Appl. Microbiol.* **12**, 229.
- Gage, J.D. and Tayler, P.A., 1991. A Natural History of organisms at the Deep Sea floor. In: Deep-Sea Biology;. Cambridge University Press, Cambridge.
- Geetha, T.S., 1982. Interactive effects of binery mixtures of metals on an estuarine calm-assainment and kinetics. Ph.D. Thesis, Cochin University of Science and Technology, Cochin.
- Gledhill, W, E., 1974. Linear Alkylbenzenesulphonate: biodegradaations and aquatic interactions. *Adv. Appl. Microbiol.* **17**, 265-293.
- Gledhill, W, E., 1975. Screening test for assessment of ultimate biodegradability.(LAS). Appl.Microbiol. **30**, 922-929.
- Goerlitz D. F., Troutman, D.E., Godsy, E.M. and Franks, B.J., 1985. Migration of wood preserving chemicals in contaminated ground water in a sand aquifer at Pensacola, Florida, *Enviro. Sci. Technol.* **19**, 955-961.
- Goldschmidt, V.M., 1954. Boron, In: *Geochemistry*, A Muir, ed., Oxford University Press, pp 280-291.
- Goldschmidt, V.M. and Peters, G., 1932. Zur geochemie des Bors. Naturwissench, Gottinge, *Mathphys. Kalasse* **111**, 25 1V, 402.
- Granmo, A., 1972. Development and growth of eggs and larvae of Mytilus edulis exposed to a linear dodecyl-benzene-sulphonate (LAS). Mar. Biol., 15, 356-358.
- Grant, T.M. and King, C.J., 1990. Mechanism of irreversable adsorption of phenolic compounds by activated carbon. *Industrial Eng. Chem. Res.*, **29**(2), 264-271.
- Gupta, S., 1985a. Effect of phenolic compounds on alkaline phosphatase activity in certain tissues of Notopterous notopterous (Pallas). Proc. Symb. Asses. Environ. Pollut., The accademy of environmental Biology, India, Muzaffarnagar, 151-157.
- Harder, H., 1970. Boron content of sediments as a tool in facies analysis. In:

 Sedimentary Geology, Elsevier Publishing Company, Amsterdam Printed in
 The Netherlands. 4, pp 153-175.
- Harder, H., 1974. Boron, In: Hand Book of Geochemistry, 1-11, K.H Wedepohl, ed., Springer.
- Harder., 1959b. Nachr. Akad. Wiss. Gottingen, 11. Math. Physik. Kl., 6:123-183.
- Harder.N., 1961b. Einbau, Von Bor in detritisch Tonminerale. Geochem. et. Cosmochim, Acta., 21. 284-294.

- Haslam, E., 1989. Plant polyphenols: Vegetable tannins. Revised, Cambridge.
- Healy, P.L., Ernst, R. and Arditti, J., 1971. Biological activity of surfactant 11. Influence on the ultrastructure of orchid seedling. *New Phytol.* **70**, 477.
- Hedges, J.I. and Ertel, J.R., 1982. Characterisation of lignin by gas capillary chromatography of cupric oxide oxidation products. *Anal. Chem.* **54**: 174-178.
- Hedges, J.I. and Man, D.C., 1979a. The charecterisation of plant tissue by their lignin oxidation products. *Geochim. Cosmochim. Acta.*, **43**, 1803-1807.
- Hedges, J.I. and Parher, P.L., 1976. Land derived organic matter in surface sediments from the Gulf of Mexico, Geochemica et. Cosmochimica Acta. 40, 1019-1029.
- Heimburger, S.A., Blevins, D.S., Bostwick, J.H. and Donnini, G.P., 1988a. Kraft mill bleach plant effluents: Recent developments aimed at decreasing their environmental impact, part 1. *TappiJ.*. Oct., 51-59.
- Hideshige Takada, Ryoshi Ishiwatari and Norio Ogura., 1992. Distribution of Linear alkyl benzenes (LABs) and Linear alkyl benzenes sulphonates (LAS) in Tokyo bay Sediments. Estuarine Coastal and Shelf Science, 35, 141-156.
- Hidu, H., 1965. Effects of synthetic surfactants on the larve of clams (M.mercineria) and oyester (C.virginica). *J.Water pollut Control Fed.* 37, 262-270.
- Hodson, P.V. and Blunt, B.R., 1981. Temperature induced changes in pentachlorophenc chronic toxicity to earlier life stages of rainbow trout. *Aquat Toxicol.* 1: 113-122.
- Hoffman, E.V., Milis, G.L., Latimer, J.G. and Quinn, J.W., 1984. Urban runoff as a source of PAH to coastal waters. *Environ. Sci. Technol.* **18**, 580-587.
- Holcobe, G.W., Philips, G.L. and Fiandt, J.T., 1982. Effects of phenol, 2, 4-dimethylophenol, 2, 4-dichlorophenol and pentachlorophenol on embryo, larvae and early juvenile fathead minnows. *Arch Environ Contam Toxicol.*, **11**: 73-78.
- Hon-Nami , H. and Hanya, T., 1980. Linear alkylbenzene sulphonate in river, estuary and bay water. *Wat. Res.* **14**, 1251-1256.
- Hunt.G., 1976. M S, Thesis, Rutgers- The State University, New Brunswick, N J.
- Hunter, K.A., 1980. Processes affecting particulate trace metals in sea surface micro layer. *Mar.Chem.* **9**, 49-70.
- Ishiwatari, R., Takada, H., Yun, S.-J and Matsumoto, E., 1983. Alkylbenzene pollution of Tokyo bay sediments. *Nature* (London)., **301**, 599-600.
- Janet A.Sweetman and Simmons, M.S.,1980. The production of bromophenols resulting from the chlorination of waters containing bromide ion and phenol. *Wat. Res.* 14, 287 290.
- Jin, K.R. and Raney, D.C., 1991. J. Waterways Port Coastal and Ocean Engg., Division Asce, 117, 451.

- Joanne G.Jennings., Rockey de Nys., Timothy S Charlton., Mark W Duncan and Peter D. Steinberg., 1996. Phenolic compounds in the nearshore waters of Sydney, Australia. *Mar. Freshwater Res.* 47, 951-959.
- John R. Ertel., John I. Hedges., Allan H. Devol., Jeffferry E. Richey. and Maria de Nazare Goes ribeiro., 1986. Dissolved Humic sudstances of the Amazon River system. Limnol. Oceanogr., 31 (4), 739-754.
- Jones, J.R.E., 1951. The relationship of the minnow, phoxinus phoxinus (L.) to solutions of phenol, o-phenol, p-cresol. *J. Exp. Biol.* **28**, 261-270.
- Jones, P.A., 1981. Chlorophenols and their impurities in the Canadian environment. Environmental Protection Service, Department of Environment, Ottawa, Canada, Catalogue No. En46-3/81-2, 434pp.
- Jorge A. Herrera Silveria and Javier Ramirez- Ramirez., 1996. Effect of natural phenolic material (tannin) on phytoplankton growth. *Limnol. Oceanogr.* **41** (5), 1018-1023.
- Joseph, J. and Kurup, P.G., 1989. Volume transport and estuarine features at Cochin inlet. *Mahasagar*, **22**, 165 172.
- Jungclaus, G.A., Lopez-Avila, V. and Hites, R.A., 1978. Organic compounds in an industrial waste water; a case study of their environmental impact. *Environmental Science and Technology.*, **12**, 88-96.
- Justin Hurst, C., Ronald C. Sims., Judith L.Sims., Darwin L.Sorensen., Joan E.McLean., and Scott Huling., 1997. Soil Gas Oxygen Tension and Pentachlorophenol Biodegradation. *Journal of Environmental engineering*, April 1997, 364-370.
- Kadam, A.N. and Bhangale, V.P., 1996. Rapid determination of total phenols in seawater by 4-aminoantipyrene colorimetry. *Indian J.: Mar. Sci.*, **25**, 46-49.
- Karanth, N.G., Lokbharathi, P.A. and Nair, S., 1975. Distribution of Phenolic Acids in soil from two mangrove areas in Goa. *Indian J. Mar. Sci.*, **4**. 215-217.
- Kaufman, D.D., 1978. Degradation of Pentachlorophenol in soil, and by soil micro organisms. Plenum Publishing corp, New York, NY.
- Kazuho Inaba, 1992. Quantitative assessment of natural purification in wetland for linear alkylbenzenesulphonates. *Wat. Res.*, **26** (7), pp 893-898.
- Kennish, M.J., 1990. Ecology of Estuaries. Vol.2, Biological Aspects, CRC Press, Boca Raton, FL.
- Khan, S.U., 1982. Studies on bound ¹⁴C-prometryn residues in soil and plants. *Chemosphere*, **11**, 771-795.

- Kikuchi, M., Tokai, A. and Yoshida, T., 1986. Determination of trace levels of linear alkylbenzene sulphonates in the marine environment by high-performance liquid chromatography, *Wat. Res.*, **20**, 643-650.
- Kimerle, R.A. and Swisher, R.D., 1977. Reduction of accute toxicity of Linear alkylbenzenesulphonate (LAS) by biodegradation. *Wat.Res.* 11, 31-37.

(PCP) on the oxygen consumption rate of the river puffer fish takifugu obscurus.

- Kinzel, L.H., McKenzie, R.M., Olson, B.A., Kirsch, D.G. and Shull, L.R., 1979. Priority pollutants. 1: a perspective view. *Envir. Sci. Technol.*, **13**(4), 416-423.
- Kirk, T.K., 1984. Degradation of lignin (Biomass production). *Microbiol. Ser.* 13. 399-437.
- Kishino, T. and Kobayshi, K., 1995. Relation between toxicity and accumulation of chlorophenols at various pH, and their absorption mechanism in fish. *Water Res.*, **29**, 431-441.
- Kitunen, V.H., Valo, R.J. and Salkinoja-Salonen, S., 1985. Analysis of chlorinated phenols, phenoxyphenols and dibenzofurans around wood preserving facilities. *Int.J.Envir.Analyt. Chem.* **20**, 13-18.
- Kitunen, V.H., Valo, R.j. and Salkinoja-Salonen, S., 1987. Contamination of soil around wood-preserving facilities by polychlorinated aromatic compounds. *Envir.Sci.Technol.* **21**, 96-101.
- Knoesel, D., 1959. Z. Pflanzernernuhr. Dung. Bodenk., 85, 225.
- Knut P. Kringstad., Filipe de Sousa and Lars M. strommberg., 1985. Studies on chlorination of chlorolignins and Humic Acid. *Environ. Sci. Technol.* 19, 427-431.
- Kozarac, Z., Cosovic, B., and Branica, M., 1975. Spectrophotometric determination of anionic surfactants in seawater. *Mar.Sci.communication*. 1, 147-763
- Krajnovic-Ozretic,M. and Ozretic B., 1988. Toxic effects of phenol on Gray Mullet, Mugil Auratus Risso. *Bull. Environ. Contam. Toxicol.*, **40**, 23-28.
- Krenger,H, Lui, S.D., Chapman, G. and Chang, J.T., 1966. Effects of pentachlorophenol on the fish Cichlasoma bimaculoatum. *Third Int Pharmacol Congr Abstr*, S Paul, Brazil.
- Krishna Kumar., P.K., Damodaran, R. and Nambisan, P.K., 1990. Acumulation, distribution and dipuration of mercury in the green mussel Perna viridis (Linnaeus). *Proc. Indian Acad. Sci.* (Anim. Sci.) 99, 345-352.

- Kristoffersson, R., Broberg, s. and Oikari, A., 1973. Physiological effects of a sublethal concentration of phenol in the pike (Esox lucius L.) in pure brakish water. *Ann Zool Fennici.*, **11**: 220-223.
- Krumbein, W.C. and Pettijohn, F.J.,1938. Manual of sedimentary Petrography. *Appletion Centuary Crafts Inc.*, New York, 549.
- Kuehl, D.W., 1981. Unusual polyhalogenated chemical residues identified in fish tissue from the environment. **10**: 231-242.
- Kuwatasuka, S. and Igarashi, M., 1975. Degradation of Pentachlorophenol in soils 11: The relationship between the degradation of PCP and the properties of soils, and the identification of degradation products of PCP. Soil. Sci. Plant Nutr., 21(4), 405-414.
- Lakhmanan, P.T. and Nambisan, P.N.K., 1977. Toxicity of copper on the bivalve Villorita cyprinoides var. cochinesis. *IndianJ. Mar. Sci.*. **6**, 83-85.
- Lakhmanan, P.T., and Nambisan, P.N.K., 1983. Seasonal variation of trace metals in bivalve molluscus Villorita cyprinoides var. cochinensis (Hanley), Meretrix casta (Chemnitz) and Perna viridis (Linnaeus). *IndianJ. Mar. Sci.*, **12**, 100-103.
- Lakhmanan, P.T., and Nambisan, P.N.K., 1985a. Tissue lactic acid and glycol level of molluscs exposed to Cu and Hg. *Curr. Sci.*, **54**, 478-479.
- Lakshmanan, P.T., and Nambisan, P.N.K., 1985b. Uptake and loss of mercury in three bivalve molluscs, Perna viridis (Linnaeus), Villorita cyprinoides var. cochinensis (Hanley) and Meretrix casta (Chemnitz). In: Harvest and Post Harvest Technonology of Fish. Soc. Fisher. Technologists (India), Matsyapuri, Cochin. Pp 419-423.
- Lakhmanan, P.T., and Nambisan, P.N.K., 1989. Bioaccumulation and depuration of some trace metals in the mussel Perna viridis (L.). *Bull. Environ. Contam. Toxicol.*, **43**: 131-138.
- Lakhmanan, P.T., and Nambisan, P.N.K., 1979. Accumulation of mercury by the mussel Perna viridis (L.). *Curr. Sci.*, **48**, 672-674.
- Lakhmanan, P.T., and Nambisan, P.N.K., 1980. Biochemical composition of the bivalve molluscus Villorita cyprinoides var. cochinensis (Hanley) and Meretrix casta (Chemnitz). *IndianJ. Mar. Sci.*, **9**, 65-67.
- Lakshmanan, P.T., Shynamma, C.S., Balchand, A.N., Kurup, P.G., and Nambisan, P.N.K., 1982. Distribution and seasonal variation of temperature and salinity in cochin backwaters. *IndianJ. Mar. Sci..*, **11**, 170-172.
- Lamar, R.T. and Dietrich, D.M., 1990. In situ deplation of Pentachlorophenol from contaminated soil by Phenerochaete spp. *Appl, Envir. Microbiology*, **56**(10), 3093-3100.

- Landergren, S., 1945. Distribution of boron in some Swedish sediments., rocks and iron ores. *Ark.Kemi.Min.Geol.*, **19**A, 1-31.
- Larson, R.A. and Rockwell, A.L., 1979. Chloroform and chlorophenol production by decarboxylation of natural acids during aqueous chlorination. *Envir. Sci. Technol.* **13**, 325-329.
- Larson, R. J., 1983. Comparison of biodegradation rates and laboratory screening studies with rates in natural waters. *Residue Rev.* **85**, 159-171.
- Lewis, M.A., 1990. Chronic toxicities of surfactants and detergent builders to algae.

 A review and risk assesment. *Ecotoxicology and Environmental Safety*, **20**, 123-140.
- Lewis, M.A., 1991. Chronic and sublethal toxicities of surfactants to aquatic animals. A review and risk assessment. Wat. Res. 25, 101-113.
- Liddicoat, M.T., Turner and Whitefield, D.R., 1983. Conservative behavior of Boron in the Tamar Estuary (England), Estuar. Coast. Shelf. Sci, 17, 467-472.472.
- Likens, G.E., Mackenzie, F.T., Richey, J.E., Sedell, J.R. and Hasss, J.R., 1981.

 Flux of organic carbon by rivers to the ocean. Conf. 8009140. DOE, Office Energy Res., Washington, D.C.
- Liss, P.L. and Pointon, M.J., 1973. Removal of dissolved boron and silicon during estuarine estuarine mixing of river water. *Geochemica Cosmochemita Acta*, **37**, 1493-1498.
- Liss, P.S. and Spencer, C.P., 1970. Abiological processes in the removal of silicate from seawater. *Geochimica et Cosmochimica Acta*. **34**, 1073-1088.
- Livingstone, D.A., 1963. Chemical composition of river and lakes, U.S Geol. Surv. Prof.Pap. 440, 64 pp.
- Loganathan, B.G. and Kannan, K., 1991. Time perspective of organochlorine contamination in the global environment. *Mar. Pollut. Bull.* **22**, 582 –584.
- Loglio, G., Dengli Innocenti, N., Tesei, U., Stortini, A.M. and Cini, R., 1989a.

 Surfactant and particulate matter exchange at the air-water interface in Antartic environment. *Ann. Chim.* (Rome) **79**, 571-587.
- Lombardini, P.P., Piazzese, F. and Cini, R., 1982. The Marangoni Wave in ripples on an air-water interface covered by a spreading film. 11 Nuovo Cimento. **C5**, 256-267.
- Loll, M.J. and Bollag, J.M., 1983. Protein transformation in soil. Adv. Argon, 36, 351-382.

- Lundhal, P. and Cabridenic, R., 1974. Molecular structure-biological properties relationship in anionic surfactant active agents. *Wat. Res.* **12**, 25.
- Maltulova, D., 1964. Influence on detergents on water algae, Sborn. Vys. Skoly Chem. -Techn. V Praze, techn. Vody, 8, 251-301.
- Mantoura, R.F. and Woodward, E.M., 1983. Conservative behaviour of riverine organic carbon in the Severn estuary: Chemical and geochemical implications. Geochim. Cosmochim. Acta. 47, 1293-1309.
- Margaritis, A. and Creese, E., 1979. Toxicity of surfactants in the aquatic environment:

 A review. In: Waste Treatment and Utilisation (M. Moo Yong and G.J.

 Farquhar, Eds.) pp 445-463, Oxford Pergamon Press.
- Martein, M.G., Bressan, M. and Brunetti, R., 1991. Effects of linear alkylbenzene sulphonate (LAS) on two marine benthic organisms. Submitted to *Aquatic. Toxic*.
- Martinez, J., Vives-rego, J. and Sanchez-Leal., 1989. The effect of chemical structure and molecular weight of commercial alkylbenzene on the toxic response of Daphnia and naturally occurring bacteria in fresh and sea water. *Wat. Res.* 23 (5), pp 569-572.
- Mathers, R.A., Brown, J.A. and Johansen, P.H., 1985. The growth and feeding behavior responses of large mouth bass (Micropterus salmoides) exposed to PCP. Aquat Toxicol., 6: 157-164.
- Matsumoto, G., Ishiwatari, R. and Hanya, T., 1977. Gas Chromatographic mass spectrophotometric identification of phenols and aromatic acids in river waters. Water Res. 11, 693 –698.
- Meybeck, M., 1981. River transport of organic carbon to the ocean, p 219-269. In: Flux of organic carbon by rivers to the ocean. Conf. 8009140.
- Meybeck, M., 1982. Carbon, nitrogen and phosphorous transport by world rivers. Am.J. Sci. 282, 401-450.
- Mc Creary, J.J. and Snoeyink, V.L., 1981. Environ. Sci. Technol. 15, 193.
- Mc Intyre, F., 1970. Geochemical fractionation during mass transfer from sea too the air breaking bubbles. *Tellus*, **22**, 451-461.
- McCalla, T.M. and Hasking, F.A., 1964. Bact. Rev., 28, 181.
- McConnachie, P.R. and Zahalsky, A.C., 1991. Immunological consequences of exposure of Pentachlorophenol. *Arc. Of Envir. Health*, **46**(4), 249-253.
- Menon, N.R., 1986. Combined toxicity of silver and copper on the sublethal response on Perna viridis. Proc. Natn. Sem. on Mussel Watch. University of Cochin. pp 13-14.

- Mikhaylov, V.I., 1978. Concentration of some Anthropogenic Substances in the surface microlayer (Exemplified by the Northeastern Part of the Atlantic).

 Oceanology, 18(5), 547-549.
- Mitrovic, V.V., 1982. Monohydric phenols: In: Water quality criteria for freshwater fish.

 Alabaster, J.S. and Lloyed, R. (eds). Butterworths, London, pp 103-125.
- Mitrovic, V.V., Brown, V.M., Shurben, D.G. and Berryman, M.H., 1968. Some pathological effects of sub-acute and acute poisoning of rainbow trout by phenol in hard water. *Wat. Res.* **2**, 249-254.
- Mitrovic, V.V., 1972. Sublethal effects of pollutants on fish. In: Marine Pollution and Sea life, Food and Agriculture Organisation, (fishing News books Ltd. M. Ruive.ed.). London, p 134.
- Moeller, J.R., 1979. Transport of dissolved organic carbon in streams of different physiographic charecteristics. *Organic Geochem.* **1**, 139-150.
- Mooper, K. and Dagem, E.T., 1979. Organic carbon in the ocean: nature and cycling. In: *The global carbon cycle* (Scope 13). Bolin B., Degens, E.T., Kempe, S. and Ketner, P. (eds). John Wiley & Sons, New York, p 2993-315.
- Mortland, N.M. and Halloran, L.J., 1986. Polymerisation on aromatic molecules on smectite. *J.Soil Sci. Soc. Am.*, **40**, 367-370.
- Nair, C.K., Balchand, A.N. and Nambisan, P.N.K., 1990. Metal concentration in recently deposited sediments of Cochin backwaters, India. Sci. Total Environ. 97/98, 507-524.
- Nair, C.K., Balchand, A.N. and Nambisan, P.N.K., 1991. Heavy metal speciation in the sediments of Cochin estuary determined using chemical extraction techniques. *Sci. Total Environment.*, **102**, 113-128.
- Nakhla, G., Qubaih, J., Abu-Zaid, N. and Abdulappa, M., 1990. Impact of nitrogen on the absorptive capacity of activated charcoal. *Envir. Technol.*, **11**, 731-738.
- Naqvi, S.W.A. and Reddy, C.V.G., 1978. On the variation of calcium content of the waters of Laccadives (Arabian Sea). *Mar.Chem.*, **8**, 1-7.
- Nedwell, D.B. and Lawson, P.A., 1990. Degradation of digested sewage sludge in marine sediment-water model systems and fate of metals. *Mar.Pollut.Bull.*, **21**, 87.
- Nihoul, J.C. and Ronday, R.T., 1987. Water Resources Res., 23, 143.
- Nyberg, H., 1976. The effects of some detergents on the growth of Nitzschia holsatica Hust. (Diatomeve). *Ann. Bot. Fenn.* **13**, 65-68.
- Odum, E.P., 1983. Basic Ecology. Saunders College publishing, Philadelphia, P.A, p 613.

- Olli-Pekka Penttinen., 1995. Chlorophenols in aquatic environments: structure activity correlations. *Ann. Zool. Fennici.* **32**: 287-294.
- Oppo, C., Bellandi, S., Dengli Innocenti, N., Storlini, A.M., Logio, G., Schiavuta, E. and Cini.R., 1999. Surfactant components of marine organic matter as agents for biogeochemical fraction and pollutant transport via marine aerosols. *Mar. Chem.*, 63, 235-253.
- Ouseph, P.P., 1987. Heavy metal pollution in the sediments of Cochin estuarine system. *Proc. Natn. Sem. Estuarine Management*, Trivandrum. Pp 123-127.
- Ouseph, P.P., 1990. Dissolved, particulate and sedimentary mercury in the Cochin estuary, southwest coast of India. In; *Estuarine Water Quality Management*, Michaelis, W (ed), Springler Verlag, Berlin. Pp 461-465.
- Paasivirta, J., Knuutinen, J., Maatela, P., Paukka, R., Soikkeli, J. and Sarrka, J., 1988. Organic chlorine compounds in Lake Sediments and the role of the chlorobleaching effluents. *Chemosphere*, **17**, 137-146.
- Painter, H.A. and Zabel, T.H., 1988. Review of the environmental safety of LAS. Water Research Centre Meedmenham, Maelow bucks, United Kingdom.
- Park, P.K. and O'Conner, T.P., 1981. Ocean dumping research; historical and international development, In: Ocean Dumping of Industrial Wastes. Ketchem, B.H., Kester, K.R. and Perk, P.K., eds., Plenum Press, New York, 3.
- Park, P.K., Kester, D.R. and Duedall, I.W., 1983. Radioactive Wastes and the Ocean, John Willey and Sons, New York.
- Paropkari, A.L., 1979. Distribution of Organic Carbon in sediments of the Northwestern continental shelf of India. *IndianJ. Mar. Sci.*, Vol 8, pp 127-129.
- Passow, U., Alldredge.A.L. and Logan, B.E., 1994. The role of particulate carbohydrate exudates in the flocculation of diatom blooms. *Deep Sea Res.*, 41, 335-357.
- Paul, A.C. and Pillai, K.C., 1983a. Trace metals in a tropical river environment: distribution. *Water Air Sioil Pollut.*, **19**, 63-73.
- Paul, A.C. and Pillai, K.C., 1983b. Trace metals in a tropical river environment: speciation and biological transfer. Water Air Sioil Pollut., 19, 75-86.
- Peit, G.J. and De Grunt, F., 1975. Organochlorocompounds in surface and drinking water of thr Netherlands. *European Colloquium*, Eur. 5196, Luxembourg 1974. Commission of the European Communities, Luxembourg, pp.81-92.
- Pillai, P. and Silas, E.G., 1975. Dynamics of zooplankton in a tropical estuary (Cochin Backwaters), with a review on the plankton fauna of the environment. *Bull. Dept. Mar. Sci. Uni. Cochin*, V11(2), 329-355

- Polisois, G., 1975. Degradation of phenolic compounds downstream from a petroleum refinery complex. *J.Fish. Res. Board. Canada.* **32**, 2125-2131.
- Prabhudeeva, K.N., 1987. Toxicity, accumulation and depuration of heavy metals in the brown mussel Pema Indica. Ph.D Thesis, Cochin University of Science and Technology, Cochin.
- Prandle, 1978. Proce. R. Soc. Lon., A 359, 189.
- Pritchard, D.W., 1952. Estuarine hydrography, Adv.Geophys, 1, 243-280.Robert H
 Byrne, Jr. and Dana R Kester., 1974. Inorganic Speciation of Boron in
 SeaWater. Journal of Marine Research, 28, 119-127
- Quimby, B.D., Delaney, M.F., Uden, P.C. and Barnes, R.M., 1980. Anal. Chem. 52, 259.
- Ragan, M.A. and Glombitza, K.W., 1986. Phorotannins, brown algal polyphenols, Progress in Phycological Research, 4, 129-241.
- Rajagopal, M.D., Rajendran, A. and Reddy, C.V.G., 1981. Distribution of dissolved boron in the waters of Zuary estuary (Goa). *IndianJ. Mar. Sci.*, **10**(1), 20.
- Ramaprasad, A., 1994. Limitation of biotransformation of Pentachlorophenol in soils.

 MSc Thesis, Utah State Univ., Logan, Utah.
- Rao, K.R., 1978. Pentachlorophenol: chemistry, pharmacology and environmental toxicology. Plenum publishing Corp., New York, N.Y.
- Ravanel, P., Taillandier, G. and Tissut, M., 1985. Effect of chlorophenol on isolated plant mitochondria activities: A QSAR study. *Ecotoxicol. Environ. Saf.*, **9**: 300-320.
- Ravanel, P., Taillandier, G. and Tissut, M., 1989. Uncoupling properties of a chlorophenol series on Acer cell suspensions. A QSAR study. *Ecotoxicol. Environ. Saf.*, **18**: 337-345.
- Reichenbach-Klinke, H.D., 1965. Arch. Fisch. Wiss., 16, 1. Composition of
- Reif, B., Lioyed, ., How, M.J., Brown, D. and Alabaster, J.S., 1979. The accute toxicity of eleven detergents to fish, result of an interlaboratory exercise.

 Wat.Res., 113, 207-210.
- Renato Cini and Guiseppe Loglio., 1997. Adsorption and pollution transport by Marine Aerosol. *Marine Pollution Bulletin*, **34**(7), 501-504.
- Rogers, I.H. and Keith, L.H., 1976. Identification and analysis of organic pollutants in water, Keith. L.H., Ed., Am. Arbor Science: Ann Arbor, MI, pp 625-639.
- Ruckdeschel, G., Renner, G. and Schwarz, K., 1987. Effect of Pentachlorophenol and some of its known and possible metabolites on different species of bacteria.
- Ryckman, D.W., Sawyer, C.N., 1975. Chemical structure and biological oxizability of surfactants. *Purdue Univ Eng Bull Etensice Series*. **12**, 270-284.

- Saarikoski, J. and Viluksela, M., 1982. Relation between physicochemical properties of phenols and their toxicity and accumulation in fish. *Ecotoxicol. Environ. Saf.* 11, 158-173.
- Saarikoski, J. and Viluksela, M., 1981. Influence of pH on the toxicity of substituted phenols to fish. *Arch.Environ. Toxico.* **10**, 747-753.
- Saarikoski, J., Lindstrom, R., Tyynela, M., Viludsela, M., 1986. Factors affecting the absorption of phenolic and carboxylic acids in the guppy (Poelicia reticulata). *Ecotoxicol. Environ. Saf*, **11**, 158.
- Sales, D., Quiroga, J.M. and gomez-Parra., 1987. Primary biodegradation kinetics off Anionic Surfactants in Marine Environment. Bull. Environ. Contam. Toxicol, 39, 385-392.
- Sankaranarayanan, V.N. and Rosamma Stephen., 1978. Particulate iron, manganese, copper and zinc in waters of the Cochin back waters. *Indian J. Mar.Sci..* **7**, 201-203.
- Sankaranarayanan, V.N., Udaya Varma, P, Balachandran, K.K., Pylee, A. and Joseph, T., 1986. Estuarine Characteristics of the lower Reaches of the River Periyar (Cochin Backwater). *Indian J. Mar. Sci.*. Vol.**15**, pp166-170.
- Sankaranarayanan, V.N., Udaya Varma, P., Balachandran, K.K., Pylee, A. and Joseph, T., .1986. Estuarine characteristics of the lower reaches of the river Periyar (Cochin Backwater). *Indian J. Mar. Sci.*, **20**, 49-54.
- Sarkanen, K.V. and Ludwick, C.H., 1971. Lignins, Wiley-Interscience.
- Saroj Gupta., 1987. Phenolic Intoxication in Fish: Symptom Complex and Physiological Activity. In: *Environment and Pesticide Toxicity*. Academy of Environmental Biology.
- Sasamal, S.K., Sahu, B.H. and Panigrahy, R., 1986. Texture & Composition of Sediments of Hooghly Estuary and Nearshore environment. *Indian J. Mar. Sci.*, Vol. 15, pp 201-202.
- Schubel, T.R. and Hirscheberg, D.J., 1978. The Chang Jing (Yantze) Estuary: Establishing its place in the Community of estuaries. In: *Estuarine comparisona*, Kennedy, V.S. (Ed)., Academic Press, New York, pp 649-666.
- Schultz, T.W., 1987. The use of the ionisation constant (pKa) in selecting models of toxicity of phenols. *Ecotoxicol. Environ. Saf.*, **14**, 178-183.
- Scott, J.C., 1986. The effect of organic films on water surface motions. In: Oceanic Whitecaps and Their role in Air-Sea exchange Processes, eds. E.C. Monahan and G. MacNiocall, pp 159-165. D. Riedel Publ., Co., Dordrecht Boston.

- Sengupta, R., Naik, S. and Singbal, S.Y.S., 1978. A study of fluoride, calcium, magnesium in the North Indian Ocean. *Mar.Chem.* **6**, 125-141.
- Seppala, J.J. and Kansanen, P.H., 1988. Fate of discharges of total organic chlorine and chlorophenol compounds in lake Etela-Saima, Finland. Water Science and Technology., 20(2), 199.
- Shanon, R.D., Boardman, G.D. and Dietrich, A.M., 1991. Mitochondrial response to chlorophenols as a short term toxicity assay. *Environ. Toxicol. Chem.*, **10**, 57-66.
- Sheldon, L.S. and Hites, R.A., 1978. Organic compounds in the Delaware River. Environmental Science and Technology, 12, 1188-1194.
- Shibu , M.P., Balchand, A.N, and Nambisan, P.N.K., 1990. Trace metal speciation in a tropical estuary: significance of environmental factors. Sci. Total Environ. 97/98; 267-287.
- Shibu, M.P., 1992. Trace Metal Speciation in the Cochin Estuary. Ph.D. Thesis, Cochin University of Science and Technology, Cochin.
- Shindo, H. and Huang, P.M., 1984. Catalytic effect of manganese (1V) and iron (111), aluminum and silicon oxides on the formation of phenolic polymers. *J.Soil Sci. Soc. Am.*, **48**, 927-934.
- Shirodkar, P.V and Kamat Dalal, V.N., 1988. Sediment boron and its relation to sediment properties in a tropical estuary. *Mahasagar*, **21**(1), 45-54.
- Shirodkar, P.V, Singbal, S.Y.S and Sen gupta, R., 1982. Studies of boron along the central West Coast of India, *Indian J. Mar. Sci.*, **11**(3), 176.
- Shirodkar, P.V. and Singbal, S.Y.S., 1992. Boron chemistry in relation to its variation in eastern Arabian Sea. *Indian J. Mar. Sci.*, **21**, 178-182.
- Shirokar, P.V.and Anand, S.P., 1985. Behaviour of boron in Mandovi estuary, Mahasagar-Bulletin of the National Institute of Oceanography, 18, 439-448.
- Sinha, P.C., Rao, Y. R., Dube, S. K. and Murthy, C.R., 1998. A numerical model for residual circulation and pollutant transport in a tidal estuary (Hoogly) of N E Coast of India. *Indian J. Mar. Sci.*, **27**, 129-137.
- Sinha, P.C., Dube, S. K., Rao, Y. R. and Chatterge, A. K., 1995. Nonlin World, 2, 257.
- Sivac, M., Goyer, M., Perwack, J. and Thayer, P., 1982. Environmental and Human aspects of commercially important surfactants. In; Solution behaviour of Surfactants (K. Mittal and E. Fendler Eds) Vol.1, Plenum, New York.
- Sivadasan, C.R., Nambisan, P.N.K. and Damodaran, R., 1986. Toxicity of mercury, copper and zic to the prawn Metapenaeus dobsoni (Meers) from Cochin backwaters. *Mar. Pollut. Bull.*, **19**, 579-580.

- Sloof, W. and Canton, J.H., 1983. Comparison of the susceptibility of 11 fresh water fishes. *Trans Amer Fish Soc* **108**, 415-419.
- Smith, L.H. and Chang, R.T., 1987. Water Resour. Res., 23, 143.
- Sontheimer, H., 1976. "The relationship Water Supply and River rhine", 11th Int. Water Supply Assoc. Congress, Amsterdam, The Netherlands.
- Sorial, G.A., Suidan, M.T., Vidic, R.D. and Maloney, S.W., 1993a. Competitive adsorption of phenols on GAC:1. Adsorption equilibrium. *J. Envir. Engrg., ASCE*, **119**(6), 1026-1043.
- Sorial, G.A., Suidan, M.T., Vidic, R.D. and Maloney, S.W., 1993b. Competitive adsorption of phenols on GAC:11. Adsorption dynamics under anoxic conditions. *J. Envir. Engrg., ASCE*, **119**(6), 1044-1058.
- Stephen, C., Spehar, D.L., Roush, T.H., Philips, G.L, and Pickering, Q.H., 1986. Effect of pollution on fresh water organisms. *J. Wat. Pollut. Control Fed.* **58**, 645-671.
- Stephen Opsahl. and Ronald Benner., 1998. Photochemical reactivity of dissolved lignin in river and ocean waters. *Limnol. Oceanogr.* **43** (6), 1297-1304.
- Stevenson, F.J., 1982. Humus chemistry: genesis, composition, reactions. John Wiley and Sons. Inc., New York, N.Y.
- Stout, D.J., Tate, R.K. and Molloy, F.L., 1976. Decomposition processes in New Zealand soils with particular respect to rates and pathways of plant degradation. In: The Role of Terrestrial and Aquatic Organisms in Decomposition Processes, 17th Symposium of the British Ecological Society, (edited by J.M. Anderson and A. Macfodyer), Blackwell scientific Publications, London, 97-141.
- Strickland, J.D.H. and Parson, T.R., 1972. A practical hand book of seawater analysis, Fisheries Research Board of Canada, Ottava, Bull.no. 125.
- Subba Rao, D.V., 1981. Effect of Boron on Primary Production of Nanoplankton. Can.J. Aquat. Sci., Vol.38, pp 52-58.
- Sugandhini Naik, R.J., Noronha, R.J. and Sen Gupta, R., 1986. Technical Publication No.1, FirstIndian Expedition to Antartica. Pages 87-94.
- Sujatha, C.H., 1992. Dynamics of some environmentally significant pesticides in a tropical waterway- A toxicological approach. Ph.D. Thesis, Cochin University of Science and Technology, Cochin.
- Sun, Y.B., Joyce, T.W. and Chang, H.M., 1989. Dechlorination and decoloration of high-molecular-weight chlorolignin from bleach plant effluents by an oxidation process. *Tappi. J.*. Sept., 209-213.
- Svedrup, H.U., Johnson, M.W. and Fleming, R.H., 1962. The Oceans, Prentice Hall, New York. pp 1087.

- Swisher, R.D., O'Rourke, J.T. and Tomlinson, H.D., 1964. Fish bioassay of Linear Alkylbenzenesulphonate (LAS) and intermediate biodegradation products.

 J.Amm. Oil Chem. Soc., 41, 746-752.
- Takada, H. and Ishiwatari, R., 1987. Linear Alkyl Benzene s in urban riverine environments in Tokyo: Distribution, Source and behaviour. *Environmental science Technology*, **21**, 875-883.
- Tee, K.T., 1976. J. Mar. Res. 34, 603.
- Tarazone, J.F. and Nunez, O., 1987. Accute toxicity of synthetic detergents to snails; Effects of sodium lauryl sulphate on Limnaea pregra shells. *Bull. Environ. Contam. Toxicol.* **39**, 1036-1040.
- Terada, H., 1990. Uncouplers of oxidative phosphorylation. *Environ. Health Perspect.* **87**: 213-218.
- Terzic, S., Hrsak, D. and Ahel, M., 1992. Enrichment and isolation of Linear Alkylbenzenesulphonate (LAS) dredging bacteria from estuarine and coastal waters. *Mar. Poll. Bull.*, **24**(4), 199-204.
- Thomas, N.A., 1973. Assessment of fish flesh tainting substances: Biological methods for the assessment of water quality. *ASTM.STP.52B*, 178-193.
- Thurman, E.M., 1985. Organic Geochemistry of natural waters. Kluer Academic.
- Tian-Min Xie, Katarina Abrahamsson, Elisabet Fogelquvist, and Bjorn Josefsson., 1986.

 Distribution of chlorophenols in marine environment. *Environ. Sci. Technol.*, **20**, 457-463.
- Tissut, M., Taillandier, G. and Ravanel, P., 1987. Effects of chlorophenol on isolated class A chloroplast and thylakoids: A QSAR study. *Ecotoxicol. Environ. Saf.*, **13**: 32-42.
- Tkalin, A.V., 1987. Background concentration of certain organic pollutants in Pacific Ocean Waters. *Oceanology*, Vol.28, No.6, 1. Pp735.
- Topping, G., 1976. Sewage and the sea, In: *Marine Pollution*, (Johnston, R., Ed.)

 Academic Press, London, 303.
- TramaF.B., 1955. The accute toxicology of phenolto the common blue gill Lepomis macrochirus Rafinesque. *Notulae Natute* (Philadelphia) **269**: 1-10.
- Tsuyoshi Ishikawa and Eizo Nakamura., 1993. Boron isotope systematics of marine sediments. Earth and Planetary Science Letters, 117, 567-580.
- U.S Environmental Protection Agency, 1972. "The pollution potential in pesticide manufacturing". EPA report TS-00-72-04, June.
- U.S. Environmental Protection Agency (EPA)., 1978. Reports of the adhoc study group of pentachlorophenol contaminants. *EPA/SAB/78/001*, Washington, D.C.

- U.S. Environmental Protection Agency (EPA)., 1980. Ambient water quality crieteria reports. Office of the Water Regulations and Standards, Washington, D.C.
- U.S. Environmental Protection Agency (EPA)., 1986. Ambiant aquatic water quality criteria for pentachlorophenol. *EPA/440/586/009*, Off., Of Res. and Devel., Washington, D.C.
- Uchiyama, M., 1979. Separation and determination of Fluorescent whitening agent and Alkyl Benzenesulphonate in Water. *Wat. Res.*, **13**, pp 847-853.
- Uppstrom, L.F., 1974. The boron/ chlorinity ratio of deep-sea water from the Pacific Ocean. Deep Sea Res. 21, 161-162.
- Valo, R., Kitunen, V.H., Valo, R.J., Salkinoja-Salonen, M. and Raisanen, S., 1984. Chlorinated phenols as contaminants as soil and water in the vicinity of two Finfish sawmills. *Chemosphere*, **13**, 835-844.
- Valo, R.J., Haggblom, M.M. and Salkinoja-Salonen, M.S., 1990. Bioremedation of chlorophenol containing simulated ground water by immobilized bacteria. Water Res., 24(2), 253-258.
- Van Leeuwen, J.A., Nicholson, B, C. and Hayes, K.P., 1993. Distribution of chlorophenolic compounds, from a pulp mill, in Lake Bonney, South Australia. Aust. J.Mar. Freshwater Res., 44, 825-834.
- Vasudevan NaiporT., 1992. Biogeoorganics in the sedimentary environments of Cochin Estuary. Ph.D. Thesis, Cochin University of Science and Technology, Cochin.
- Veith, G.D., Kuehel, D.W., Leonard, E.N., Puglisi., F.A. and Lemke, A.E., 1979.
 Polychlorinated bipheyls and other organic chemical residues in fish from major watersheds of the United States. 1976. Pesticide Monitoring J. ournal. 13, 1-11.
- Veningerova, M., Prachar, V., Kovaclcova, J. and Uhnak, J., 1998. Levels of chlorinated phenols in Danube river water. *Fresenius Environ. Bull.*, **7**(3-4), 224-231.
- Ventura, F., Caixach, J., Figueras, A., Espalder, I., Fraisse, D., and Rivera, J., 1989.
 Identification of surfactants in water by FAB mass spectrometry. Water Res.
 23, 1191-1203.
- Vidic, R.D. and Suidan, M.T., 1991. Role of dissolved oxygen on the addsorptive capacity of activated carbon for synthatic and natural organic matter. *Envir. Sci.and Technol.*, **25**(9), 1612-1618.
- Vives-Rego, J., Vanque, M.D., Sanchez-Leal, J. and Parra J., 1987. Surfactants biodegradation in seawater. Primary biodegradation and bacterial population response in seawater. *Tenside detergents*, **24**, 20-22.

- Wallace, D.T. and Duce, R.A., 1987. Open ocean transport of particulate trace metals by bubbles. *Deep Sea Res.*, **25**, 28-835.
- Walne, P.R., 1978. Toxicity experiments of Oyester larvae. *Ed.* Torrey Canyon Pollution and Amrine life, University Press, Cambridge.
- Waluga, D., 1966. Phenol effects on the anatomic histopathological changes in bream Abramis brama LO. *Acta Hydrobiol*, **8**: 55-78.
- Wang, T.S.C., Li, S.W. and Ferng., Y.L., 1978. Catalytic polymarisation of phenolic compounds by clay minerals. *J.Soil Sci.*, **126**(1), 15-21.
- Wang, T.S.C. and Chuang, T., 1967. Soil Science, 103, 239.
- Watanabe, I., 1977. Pentachlorophenol decomposing and PCP tolerant bacteria in field soil treatment with PCP. Soil Biol. and Biochemistry, 9(1), 99-103.
- Watanabe, I., 1978. Pentachlorophenol decomposing of field soils treated annually with PCP. Soil Biol. and Biochemistry, **10**(1), 71-75.
- Webb, P.W. and Brent, J.R., 1973. Effects of subleathal concentrations of sodium pentachlorophenate on growth rate, food conservation efficiency, and swimming performancein underyearing sockeye salmon (Oncorhychus neka), *Journal of the fisheries Research Board of Canada*, 30, 499-507.
- Wegman, R.C.C. and Hofstee, A.W.M., 1979. Chlorophenols in the surface waters of the Netherlands. *Water. Res.* **13**, 651-657.
- Wegman, R.C.C. and Van Den Broek, H.H., 1983. Chlorophenols in the river sediments in the Netherlands. *Water Res.*, **17**, 227-230.
- Weinbatch, E.C., 1957. Biochemical basis for the toxicity of pentachlorophenols. *Proc.*Natl. Acad. Sci. 43: 393-397
- Weinbatch, E.C., and Garbus, J., 1965. The interaction of uncoupling phenols with mitochondria and mitochondrial protein. *J. Biol Chem.*, **240**: 1811-1819.
- Werdelmann, B.W., 1984. Tenside in unserer Welte-heute und morgen. *Proc. Wld. Surfact. Congre.* **111**, 3-21.
- Weser, K., 1978. Levels and pattern of chlorophenols in water of the Weser estuary and the German Bright. *Chemosphere*, 873-874.
- Westall, J. and Stumm, W., 1980. The hydrosphere. In: *The HandBook of Environmental Chemistry*. Ed. O Hutzinger, **17-49**, Springer- Verlay, New York.
- Whitehead, D.C., 1964. Nature, Lon. 202, 417.
- Widdows, J. and Donkin, P., 1991. Role of physiological energetics in ecotoxicology. Comp. Biochem. Physiol., **100**C: 69-75.
- Wilfred W. Scott, 1958. Standard Methods of Chemical Analysis. Vol.1., D. Van Nostrad Company, Inc, Princeton, New Jercy. (Edited by N. Howell Furman).

- Wolfgang Ernst and Kurt Weber., 1978. The fate of Pentachlorophenol in the Weser Estuary and the German Bight. Veroff. Inst. *Meeresforsch. Bremerh*, **17**: 45-53.
- Xie, T.M., 1983. Determination of trace amounts of chlorophenols and chloroguaiacols in sediment. *Chemosphere*, **12**, 1183-119.
- Yaramaz, O. and Tuncer, S., 1988. Analyse spectrphotometrique des surfactants sur la cote Egeenne. Rapp. Comm. Int. Mer. Medit. 31, 153.
- Yassuda, H., 1981. Separation and determination of anionic active surfactants in river or seawater by solvent extraction method. *Nippon Kagaku Kaishi*, 456-461.
- Yoshinari Ambe, 1973. Determination of Alkylbenzenesulphonate (ABS) in bottom sediment. *Environmental Science and Technology*, **7**(6), 542-545.
- Zingde, M.D., Abidi, S.A.H., Sharma, P. and Rokade, M.A., 1987. Base water quality of Thal. Contributions in Marine sciences. Dr S.Z.Qasim Sastyabdapurti felicitation Volume.1, pp 307-318.
- Zingde, M.D., Narvekar, P.V., Sarama, R.V. and Desai, B.N., 1980. Water quality of river Damanganga (Gujarath). *Indian J. Mar. Sci.*, **9**, 94-99.
- Zingde, M.D., Sabnis, M.M., Mandalia, A.V. and Desai, B.N., 1980. Effect of industrial waste disposal on the water quality of the river Kolak. *Mahasagar*, **13**(2), 99-110.
- Zutic, V. and Legovic, T., 1987. A filim of organic matter at the fresh water/ seawater interface of an estuary. *Nature*, **328**, 612-614.

- Cini, R., Lombardini, P.P., Manifredic, C. and Cini, E., 1987. Ripples dumping due to monomolecular films. *Journal of Colloid Interface Science*. **119**, 74-80.
- Day, T.H., 1981. Estuarine Ecology: (ed: A. A. Balkema, Rotterdam) 58.
- Degobbis, D., Smodlaka, N., Pojed, I., Skrivanic, A and Precali, R.,1979.
 Increasing Eutrophication of the Northern Adric. *Marine Pollutution Bulletin.* **10**, 298-301.
- Faulkner, P.J.,1978. In: "Topics in Antibiotic Chemistry" (ed: PG Sames)., Vol.2, P.13, Ellis Horwood.
- Franz Bucher and Rudolf Hofer.,1993. Histopathological effects of sublethal exposure to phenol on two variously pre-stressed populations of Bullhead (cottus gobio). *Bull. Environ. Contam. Toxicaol.* 51, 309-316.
- Frelov, B.A., 1971. Problems of hydrobionts adaptation to a toxic factor. *Gidrobiol* Zn 7: 61-67.
- Galassi, S., Guzzellz, L., Mingazzini, L., Vigano, S., Capri and Sora, S., 1992.

 Toxicological and chemical characteristics of organic micro pollutants in River Po waters (Italy), *Wat. Res.* **26**(1), 19-27.
- Goldenberg, M.C, and Weiner, E.R., 1980. Anal. Chim. Acta., 115, 373.
- Grasshoff, K., Ehrhart, M and Kremliny, K., 1983. Methods of seawater Analysis. Verlag Chemie, Weinheim.
- Hega R. Gomas and Mavinkure, S., 1982. Studies on the Mangrove Swamps of Goa. Microorganisms degrading phenolic. *Mahasagar* 15(2), 111-115.
- Heimburger, S.A., Blevins, D.S., Bostwick, J.H. and Donnini, G.P.,1988 .

 Kraft mill breach plant effluents: recent developments aimed at decreasing their environmental impact, Part 2, *Tappi Journal*, November, 69-78.
- Isaccson, P.J. and Frink, C.R., 1961. Nonreversible sorption of phenolic compounds by sediment fractions: the role of sediment organic matter. *Envir. Sci. and Technol.*, 18(1), 43-48.
- James C McCarty and Richard A Wagner., Michal Macomber., Howard S
 Harris., Mervin Stephenson and Ermen A. Pearson., 1961. An
 investigation of water and sediment quality and pollution characteristics
 of three areas in San Fransisco Bay.

- Kadam, A.N. and Bhangale, V.P., 1992. Indian J. Environ Protect, 13, 15.
- Kishino, T and Koyabashi, K., 1994. Relation between the chemical structure of chlorophenols and their dissociation constants and partition coefficients in several solvent water systems. *Wat. Res.* 28, 1547-1552.
 - Kleinhempel, P., 1970. Albrect Thaer Arch., 14, 3. University of Science
- Kobayashi, K. and Akitake, H.,1975. Studies on the metabolism of chlorophenols in fish. Absorption and excretion of phenol by goldfish. *Bulletin of the Japanese Society of Scientific Fisheries.* **41**, 1271-1276.
 - Larson, R.A. and Hufnal, J.M. (Jr.), 1980. Oxidative polymerisation of dissolved phenols by soluble and insoluble inorganic species.

 Limnology and Oceanography, 25 (3), 505-512.
- Levinson, A.A. and Luddwick, J.C., 1976. *Geochim. Cosmochim. Acta.*, 30, 855.
- Makel, T.P., Petanan, T., Kukkonen, J. and Oikari, O.J., 1991. Accumulation and depuration of chlorinated phenolics in the fresh water mussle (*Anodonta anatine L*). *Ecotoxicology and Environmental Safety*. **22**., 153- 163.
- Mc Bridge, M.B., 1989. Oxidation of dihydroxybenzenes in hydrated aqueous suspension of birnessites. *Clays and clay Minerals*, **37**(4), 341-347.
- Narvekar, P.V., Zingde, M.D. and Sen Gupta. R., 1980. Behavior of Boron, Calcium and Magnesium in Mindola River Estuary (Gujarat). *Indian J. Mar. Sci..*, **10**(1), 90-92.
- Robert H Bryne (Jr) and Dana R. Kester.,1974. Inorganic speciation of Boron in Seawater. *Journal of Marine Research*. **32**(2), 119-127.
 - Sarkar., J.M., Malcom, R.L., and Bollang J.M., 1988. Enzymatic coupling of 2,4-dichlorophenol to stream fulvic acid in the presence of oxidoreductases. *J. Soil. Sci. Soc. Am.*, **52**, 688-694.
 - Sathyanathan, B., S.M. Nair, Jacob Chacko and Nambisan, P.N.K., 1988. Sublethal effects of copper and mercury on some biochemical constituents of estuarine clam villorita cyprinoids var Cochinensis (Hanley). *Environ.Contam. Toxicol*, **40**, 510-516.

- Sevenko, V.S., 1977. Oceanology, 17, pp 290.
- Subba Rao, D.V., 1979. Effect of boron on primary production of Nanoplankton. *Can. J. Fish Aquat. Sci.*, Vol.**38**. pp 52-58.
- Suresh, K., 1988. Haematology of some marine and estuarine molluscs of commercial importance. Ph.D Thesis, cochin University of Science and Technology, Cochin.
- Svedrup, H.U, John, M.W, and Fleming, R.H., 1942. The Oceans- their Physics, chemistry and General Biology, Prectice Hall, New York, 1087.
- Thomson, G and Melson., 1970. Boron contents of serpentines in the oceanic crust: implications for the boron cycle in the oceans, *Earth Planet. Sci. Lett.*, **8**, 61-65.
- Tichikawa, M., Sawmura, R., Okada, S and Hamada, A., 1991. Difference between fresh water and seawater killfish (Oryzias latipes) in the accumulation and elimination of pentachlorophenol. *Archives of Environmental Contamination and Toxicology,* **21**, 146-151.
- Uchiyama, M.,1979. Separation and determination of Flurescent whitening agent and Alky Benzene sulphonate in water. *Wat. Res.*, **13**, 847-853.
- Ulrich, H.J. and Stone, A.T., 1989. Oxidation of chlorophenols adsorbed to manganese oxide surfaces. *Envir. Sci. and Technol.*, **23**(4), 421-428.
- Wangersky, P.J.,1981. The fate of sediment organic carbon in estuaries.

 294-313. In: *Flux of organic carbon by rivers to the ocean.* Conf. 8009140. DOE. Office Energy Res., Washington, DC.
- Yasuhara, A.H., Shiraishi, M., Tsujl and Okuno, T., 1981. Analysis of organic Substances in highly polluted river water by Mass Spectrometry. *Environmental Science and Technology*. **15**, 570-573.

Annexure - 2.1. Distribution of Salinity in the surface and bottom waters of Cochin Estuarine System (ppt).

Jan-96	Feb-96	96-	Mar-96	Apr-96	May-96	96-unf	96-Inf	Aug-96	Oct-96	Nov-96
5.21		21	1.24	0.08	90.0	0.10	90.0	90.0	0.02	0.03
7.69	(())	6	10.42	0.11	0.15	0.1	0.07	90.0	0.02	0.04
6.47	44	7	1.27	0.31	0.24	0.14	0.07	0.07	0.03	0.04
14.05)5	12.84	0.93	0.4	0.1	0.07	0.07	0.02	0.04
5.79		6.	4.51	1.21	11.77	0.22	0.07	0.07	0.02	0.02
17.83	ω	33	17.69	1.52	13.09	0.22	0.7	0.07	0.02	0.02
22.59	5	6	19.30	8.12	23.75	2.65	0.10	0.13	0.24	17.63
30.39	3	0	29.71	15.06	25.05	2.82	0.1	0.12	0.024	27.6
30.50	5	0	33.44	25.00	24.65	2.42	*QN	3.08	2.10	31.75
32.72	1	2	33.75	34.24	33.57	2.94	*QN	32.51	19.65	32.15
13.87	w.	37	21.94	17.29	17.41	0.34	0.11	0.22	0.07	13.58
18.11	7.1	8	26.45	18.22	17.41	0.53	0.11	9.0	0.07	15
12.31		31	18.32	7.12	12.28	0.19	0.08	0.21	0.05	5.27
13.4	- 20	0.0	0000	200	12 02	010		0.34	0.05	6.76

S - Surface; B - Bottom; ND* - Not Determined.

Annexure - 2.2. Distribution of dissolved oxygen in the surface and bottom waters of Cochin Estuarine System (ml/l).

Nov-95	Dec-95	Jan-96	Feb-96	Mar-96	Apr-96	May-96	96-unf	36-Inf	Oct-96	Nov-96
*CN	E	3.43	3.3	3.06	3.67	4.17	4.79	5.46	4.05	4.33
*QN	3.8		3.43	2.45	3.92	4.49	5.42	5.06	4.03	4.3
3.68	3.51	3.43	3.3	3.67	3.67	5.14	4.45	5.09	4.91	3.65
4.66	3.73	3.23	3.55	3.06	1.71	4.75	4.88	4.9	4.59	4.02
4.08	3.9	3.13	2.94	3.75	2.82	5.32	4.66	4.55	3.68	2.98
281	3.04	2.79	2.57	3.43	3.3	4.37	4.58	4.63	3.95	3.22
3 86	2.38	2.94	3.06	3.67	2.82	3.76	3.71	4.17	3.26	2.98
2.74	3.04	2.45	2.82	3.79	2.94	2.99	3.83	4.19	3.68	2.98
2 91	2.95	3.33	3.92	4.29	2.45	2.9	4.44	*QN	3.28	3.31
3.68	4 09		3.55	3.79	3.18	2.54	3.46	*QN	2.54	3.46
2 66	4 18		3.67	2.94	2.69	3.53	5.54	4.54	2.6	2.89
3.41	4.18		2.45	2.94	2.45	3.25	4.92	3.57	2.53	3.01
3.95	0	2.64	3.55	2.57	2.69	1.99	3.22	2.75	2.15	2.06
271	d	2.45	3.06	2.45	1.96	3.13	3.1	3.83	2.98	2.79

S - Surface; B - Bottom; ND* - Not Determined.

Annexure - 2.3.a. Distribution of sand (%) in the sediments of Cochin Estuarine System.

Ototioto	Now 05	Dec. 05	Jan-96	Feh-96	Mar-96	Apr-96	Mav-96	Jun-96	Jul-96	Aug-96	Oct-96	Nov-96
1	*CN	99.10	98.85	96.99	94.72	98.99	97.82	97.85	97.83	97.29	99.75	99.49
0	98.59	97.23	97.28	95.72	98.10	97.77	91.39	96.76	97.77	96.97	97.87	98.96
1 6	37.77	84.11	76.54	37.40	70.34	86.15	18.99	17.31	20.23	76.14	27.57	58.65
4	0.63	4.00	38.37	1.02	1.65	4.37	4.30	7.92	10.71	8.56	21.80	16.80
. 2	19.01	14.42	21.09	10.84	5.92	24.43	38.22	42.21	*QN	48.47	42.57	57.34
9	79.76	40.72	67.82	72.71	35.63	39.82	72.56	4.02	58.85	63.16	86.38	27.42
7	46.92	83 64	86.17	85.94	73.92	73.83	6.41	75.55	89.51	54.73	52.52	90.15

Annexure - 2.3.b, Distribution of silt (%) in the sediments of Cochin Estuarine System.

Charles formation	Alzen, SA	Danida	Jan-36	Plants DR	Mar-96	Accr-96	May-96	SG-UND	OB-INF	A UG-80		NOV-VC
Stations	Nov-95	Dec-95	Jan-96	Feb-96	Mar-96	Apr-96	May-96	96-unf	96-Inf	Aug-96	Oct-96	Nov-96
-	*QN	0.52	0.11	2.91	4.79	0.56	0.89	06.0	0.19	0.26	0.15	0.01
2	1.01	0.71	0.44	3.78	0.71	1.93	6.98	0.84	0.53	0.70	0.43	0.04
6	40.55	14.45	17.11	47.71	24.12	6.41	54.12	49.01	33.43	7.52	36.36	22.79
4	45.26	53.59	36.57	52.54	59.94	40.33	51.21	30.95	27.35	28.85	22.77	32.54
5	43.20	56.58	44.28	65.83	52.85	54.89	39.39	16.82	*QN	11.38	22.35	18.17
9	16.75	36.69	21.76	19.54	38.97	42.15	22.26	28.82	3.51	1.50	0.42	29.72
7	30.89	16.27	13.09	12.03	17.29	20.52	55.04	10.36	2.33	6.97	16.91	8.16

ND* - Not Determined.

Annexure - 2,3.c. Distribution of clay (%) in the sediments of Cochin Estuarine System.

		0	90	Eoh 08	Mar.96	Apr-96	Mav-96	96-unf	Jul-96	Aug-96	Oct-96	Nov-96
Stations	CR-VON	CR-SO	Jall-90	08-00	ואומו	22 121	-			1, 0	0.40	0 4 0
~	*	0.38	1 04	0.10	0.50	0.45	1.29	1.25	1.98	2.45	01.0	0.30
-	GK.	2000			0,,	000	4 63	2 40	170	2.33	1.70	1.00
2	0.40	2.06	2.29	0.50	1.19	0.30	20.1	2.40	2	20.3		
	24.60	1 11	R 35	14 89	5.54	7.44	26.89	33.69	46.34	16.34	36.07	18.55
2	21.00	1.4.1	0.00									0001
,	54 11	42 40	25.06	46 43	38.41	55.30	44.49	61.13	61.94	62.59	55.43	90.00
4	04	74.70	20.03								00 = 0	0, 10
u	37 78	29.00	34.63	23.33	41.23	20.68	22.39	40.97	*QN	40.15	35.08	24.48
	0	20:04					9, 1	01.10	27.07	25 25	13.20	42 86
ď	3 49	22.58	10.43	7.75	25.39	18.02	5.18	67.16	37.07	35.35	13.20	12.00
1	22.40	000	0.74	2.04	8.79	5.66	38.56	14.08	8.16	38.29	30.57	1.69

ND* - Not Determined.

Annexure - 2,4. Distribution of organic carbon in the sediments of Cochin Estuarine System (in mg/g)

Stations Nov-95 1 ND* 2 3.63 3 15.3	Cec-an	90 00	Eah OR	Mar-96	Apr-96	Mav-96	Jun-96	Jul-96	Aug-96	Oct-96	Nov-96
1 ND* 2 3.63 3 15.3	Total and the second se	Jall-30	00-00-	20 20						000	0
3 15.3	1 03	161	0.53	3.12	1.41	1.41	0.82	0.27	0.68	0.36	0.48
3 15.3	30.							100	C	000	000
3 15.3	276	2 44	2.58	0.76	3.25	3.25	2.65	0.27	0.72	0.30	0.00
3 8 15.3	4.10			-	mandenocourante accessoration						1
0.01	7.04	8 54	8 22	4.86	5.75	5.75	0.82	13.49	4.78	8.59	7.48
	10.	0.0			-						0
1000	47.02	11.25	11 11	11.47	10.41	10.41	6.11	0.46	10.62	8.02 *	8.78
19.00	17.02	04:11									
123	14 72	8 79	6.98	15.02	10.62	10.62	8.58	*QN	5.33	6.56	5.41
0.71	7.12										000
0	10 13	90.0	4 66	8 04	1.18	1.18	6.87	4.38	4.75	28.6	77.7
10.0	13.13	0.00	20:-								1
12 53	577	5 95	48	8.73	7.79	7.79	0.69	8.33	6.51	3.5	2.77

Annexure - 3.1 a. Distribution of Methylene Blue Active Substances in the surface and bottom waters of Cochin Estuarine System (in ug/l).

Feb-96
5.20
9.84
4.88
17.07
8.13
13.25
12.93
20.00
62
23.99

S - Surface; B - Bottom; ND* - Not Determined.

Annexure - 3.1 b. Distribution of Methylene Blue Active Substances in the sediments of Cochin Estuarine System (in ug/g)

0 7	sira	. n 78	88.03	69.64	0.50	0.91	34,35	40.33	28.23	25 63	58.08	7.41
Stations	Nov-95	Dec-95	Jan-96	Feb-96	Mar-96	Apr-96	May-96	96-unf	36-Inc	Aug-96	Oct-96	Nov-96
0	*QN	1.09	0.72	0.91	0.91	1.29	0.88	0.57	1.14	0.07	0.24	0.11
2	0.25	1.66	0.97	2.73	0.85	1.99	1.08	0.95	1.60	1.13	0.36	0.53
က	0.39	2.42	1.75	1.44	2.56	3.10	4.77	3.64	3.45	1.51	2.88	1.21
4	06.0	5.17	3.33	3.10	3.31	3.55	4.03	3.68	2.53	2.90	2.06	4.53
2	0.32	3.05	1.96	2.19	2.48	4.44	1.26	2.29	*QN	1.08	1.85	0.84
ပ	0.29	5.57	2.99	2.65	4.54	3.55	1.41	4.35	2.20	3.40	3.00	4.30
7	0.28	2.95	2.20	2.27	3.02	2.87	6.87	2.73	2.53	2.20	1.37	1.27
(S)	(O)	13.40	30.7	0.10	180	00.03						
C												
ND* - Not [ND* - Not Determined											

Annexure - 3.2 a. Distribution of phenols in the surface and bottom waters of Cochin Estuarine System (in ug/I).

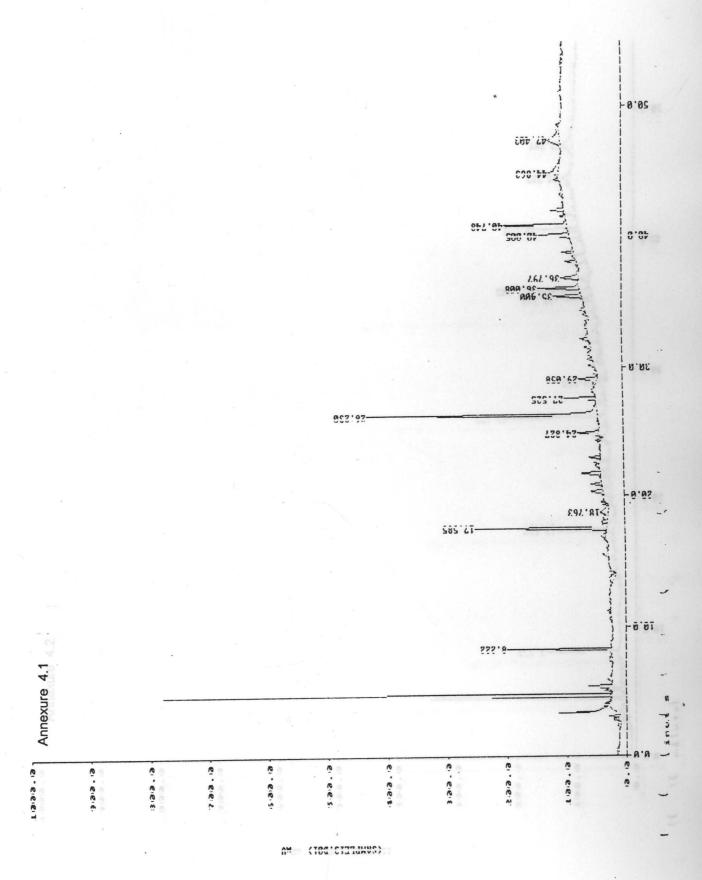
w	*QN	Dec-85	30 00	Eoh OR	Mar-96	Apr-96	Mav-96	Jun-96	Jul-96	Aug-96	Oct-96	Nov-90
m (0, m	ND	0 12	Jan-90	12 50	0.50	0.91	34.35	40.33	28.23	25.35	29.09	7.41
B S B		0.70	12.00	14.50	0.25	0.39	34.03	46.17	50.00	26.35	27.15	9.99
S	*ON	15.40	40.71	14.04	1.78	0.39	35.97	28.50	23.00	23.34	26.68	16.28
В	22.28	8.01	0.80	10.12	0.40	47.02	35.48	28.50	23.00	23.34	26.68	16.28
	35.31	6.26	9.12	10.51	8/.8	20.71	01.00	20.60	24 00	30.50	22.33	14.94
3.8	12.77	14.40	8.53	8.44	0.38	0.52	28.00	29.30	24:00	2000		44.00
a	17.53	11.89	15.17	6.36	1.63	0.52	28.06	29.50	24.00	30.50	22.34	14.39
	200	n 7	10.78	10.25	10.42	13.77	15.32	15.12	15.50	24.77	16.55	9.88
0 1	00.	0.0	1 1 1 2	4.67	5 65	7.53	18.39	14.76	22.50	21.19	21.69	11.12
9	10.02	10.7	2 3	200	20.0	20.30	15.00	25.73	*QN	21.05	30.53	8.87
5. S	6.51	13.40	78./	27.5	0.0	20.00	16 13	10.85	*GN	8.16	34.39	6.51
В	3.13	14.90	3.32	10.01	0.0	14.23	7 2	47.40	000	24 06	21 69	9.10
8.9	27.79	11.64	8.65	9.60	9.80	2.07	17.20	61.71	00.0	20.72		27.0
	3.76	12.02	13.03	5.45	9.67	12.73	20.81	19.51	16.17	20.76	39.85	9.43
	20.0	15.27	10 19	1.69	9.17	3.38	25.16	15.24	11.33	17.76	33.26	10.44
0 0	0.40	18 48	14 22	4 80	7.66	11.17	17.42	20.00	12.33	23.48	66.53	12.73

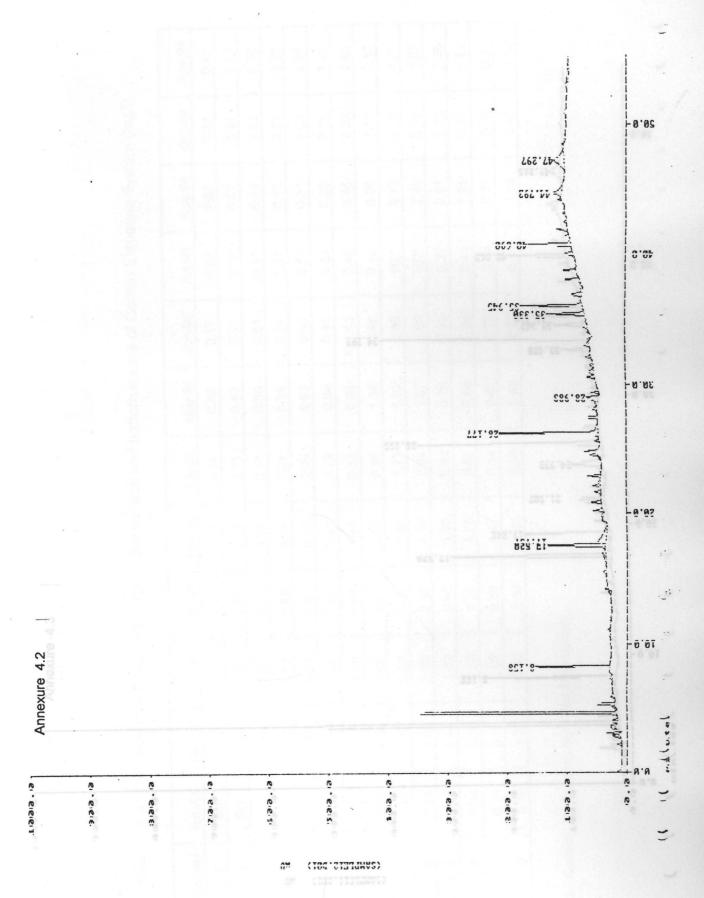
S - Surface; B - Bottom; ND* - Not Determined.

Annexure - 3.2 b. Distibution of phenols in the sediments of Cochin Estuarine System (in ug/g)

	Pr Colored	-									-	
040	NOV OF	Dec-95	Jan-96	Feb-96	Mar-96	Apr-96	May-96	Jun-96	Jul-96	Aug-96	Oct-96	Nov-96
Stations	*010	0.67	5.28	0.39	3.97	1.04	1.70	00.00	0.03	0.27	0.38	46.17
- 0	7 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7	0.54	000	3.56	14.41	3.80	0.42	00.00	1.17	00.00	8.27	0.57
7 0	7.58	20.0	000	0.94	0.94	1.10	8.89	1.63	0.82	4.34	0.38	21.06
2	00.7	0.05			28.27	4 08	9.87	61.57	7.07	1353	0.00	4 53
4	9.41	2.32	0.00	5.55	20.5	20:-	5	5	14: -	0.0		
Ľ	0.22	0.58	8.10	5.15	5.09	5.37	00.00	13.36	*QN	2.42	35.18	36.77
2 0	1 80	1.45	19.92	1.72	4.28	1.10	00.00	00.0	0.60	00.0	1.12	3.36
1 0	000	26.0	7.78	1 98	1.61	14.54	7.16	9.65	0.31	4.64	35.04	12.13
,	0.84	16.3	2	20:1			0.7					

ND* - Not Determined.





Distribution of dissolved boron in the surface and bottom waters of Cochin Estuarine System (mg/l). Annexure - 5 a.

Nov-96	0.49	2.23	0.36	0.36	2.36	1.87	2.90	3.12	3.21	3.97	1.96	1.92	0.27	112
Oct-96	0.78	5.91	2.33	2.33	0.23	0.16	2.02	1.17	0.70	3.58	0.23	0.31	0.78	2.10
Aug-96	1.87	0.67	0.13	0.13	0.31	0.22	0.45	0.09	0.18	2.45	2.01	1.29	0.71	143
Jul-96	0.07	0.79	0.17	0.17	0.17	0.34	0.48	0.45	*QN	*QN	0.27	0.41	0.14	0.34
Jun-96	0.17	0.31	0.51	0.14	0.38	0.41	1.13	0.45	0.58	2.09	0.65	0.93	0.72	080
May-96	0.46	0.43	0.39	0.30	0.63	0.69	0.89	1.54	1.02	1.67	0.76	0.98	0.67	200
Apr-96	0.14	0.71	0.14	0.24	0.09	0.19	0.52	0.66	1.23	2.04	06.0	0.57	0.14	000
Mar-96	0.57	0.14	0.05	0.38	0.57	1.47	1.47	1.56	1.94	2.18	06.0	1.75	1.47	4 00
Feb-96	0.00	0.28	00.00	0.99	0.22	1.40	1.47	2.67	3.06	2.98	06.0	1.27	0.38	000
Jan-96	0.15	0.00	0.00	0.12	0.53	1.87	1.78	3.05	3.07	3.30	1.53	2.18	1.07	
Dec-95	0.10	0.49	0.14	0.38	1.03	0.69	2.22	1.45	1.45	1.89	1.44	1.04	0.18	
Nov-95	*QN	*QN	0.61	0.56	0.62	0.66	0.04	1.78	1.17	1.26	0.98	0.24	0.16	
Stations	s.t	В	2.5	В	3.8	В	4 S		S	8	6.8	8	7.8	

S - Surface; B - Bottom; ND* - Not Determined.

Annexure - 5 b. Distribution of water soluble boron in the sediments of Cochin Estuarine System (in mg/kg).

Nov-96	11.09	15.52	15.65	19.34	22.99	18.27	22.74
Oct-96	12.86	16.61	17.14	22.26	12.37	18.56	18.29
Aug-96	21.76	11.33	3.56	5.09	14.06	6.07	21.06
96-Inf	49.26	10.94	44.42	37.10	*QN	50.29	63.37
96-unf	8.33	12.41	9.86	8.07	14.09	103.42	6.95
May-96	12.18	10.50	20.08	40.82	22.34	22.29	11.40
Apr-96	26.17	11.35	10.88	13.21	18.52	44.12	21.83
Mar-96	11.19	25.67	7.63	21.79	28.05	16.46	17.30
Feb-96	6.23	6.39	13.19	12.37	11.81	9.00	10.47
Jan-96	22.01	31.80	12.94	61.18	28.58	14.00	29.30
Dec-95	6.46	4.50	12.60	6.05	7.31	6.27	10.92
Nov-95	*QN	4.54	2.47	6.24	8.83	7.59	2.43
Stations	1	2	3	4	5	9	7

Anexure -5 c. Distribution of water insoluble (acid soluble) boron in the sediments of Cochin Estuarine System (in mg/kg).

Nov-96	182.60	181.16	31.77	51.05	136.43	220.50	69.30
Oct-96	67.25	191.54	137.99	132.27	83.28	45.01	35.84
Aug-96	27.90	46.18	12.68	14.12	67.07	60.12	11.67
36-Inf	50.59	72.53	80.58	5.16	*QN	7.26	6.61
Jun-96	16.31	20.35	43.82	21.87	20.23	65.25	55.32
May-96	24.94	20.66	37.54	23.20	23.47	24.26	21.43
Apr-96	54.49	58.46	15.24	26.76	30.68	37.47	99.95
Mar-96	31.72	49.85	48.89	38.07	122.94	14.37	49.86
Feb-96	13.60	14.64	20.44	53.64	158.52	14.16	15.65
Jan-96	5.42	134.08	20.14	42.33	73.68	38.03	30.29
Dec-95	11.26	13.53	10.88	13.52	9.77	20.95	9.60
Nov-95	*QN	51.06	118.98	110.68	70.50	12.31	7.74
Stations	-	2	3	4	5	9	7

ND* - Not Determined.