

ZOOPLANKTON STUDIES IN THE
COCHIN ENVIRONS

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DOCTOR OF PHILOSOPHY

By

P. HARIDAS M. Sc.



REGIONAL CENTRE
NATIONAL INSTITUTE OF OCEANOGRAPHY
COCHIN - 682 018

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C E R T I F I C A T E

This is to certify that this thesis is an authentic record of the work carried out by Mr. P. Haridas, M.Sc. under my supervision at the Regional Centre of National Institute of Oceanography, Cochin and that no part thereof has been presented before for any other degree in any University.

T S S R

(T.S.S. RAO)

HEAD

BIOLOGICAL OCEANOGRAPHY DIVISION
NATIONAL INSTITUTE OF OCEANOGRAPHY
GOA.

SUPERVISING TEACHER.

Dona Paula.

30.12-'82

C E R T I F I C A T E

This is to certify that this thesis is an authentic record of the work carried out by Mr. P. Haridas, M.Sc. under my joint supervision with Dr. T.S.S. Rao at the Regional Centre of National Institute of Oceanography, Cochin and that no part thereof has been presented before for any other degree in any University.

Mk7888

**(M. KRISHNANKUTTY)
SCIENTIST-IN-CHARGE,
NATIONAL INSTITUTE OF OCEANOGRAPHY,
REGIONAL CENTRE,
COCHIN - 682 018.**

Cochin-18.

30.12.82



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1. INTRODUCTION.

Estuaries are important areas of human use for fisheries, transportation, food production and recreational pursuits. They form the nursery grounds for the larvae and young ones of numerous marine species, many of them commercially important. However, often they are turned into receptacles of human wastes and subject to pollution from industrial effluents.

Estuaries are highly productive areas and contribute substantially to the productivity of the coastal waters. With the depleting marine food resources and increasing population, the focus has now shifted to aquaculture. Estuaries because of their accessibility and semi enclosed nature, have emerged as ideal sites for these activities.

Various kinds of estuaries from coastal plain estuaries to steep sided fjords exist, all with the common feature of being areas where fresh water meet with sea water, thus forming buffer zones between the marine and limnetic counterparts. The most glaring environmental feature of the estuaries is the wide nature of fluctuations that occur in them. Understandably, estuaries have developed its own flora

and fauna which are capable of withstanding or adapting to these changes. A cursory look at any estuarine biota will show predominantly estuarine forms, mixed with some stragglers from the adjoining marine and fresh water environments.

A proper understanding of the environmental parameters and their effects on the biota is a must in the management of any ecosystem. Zooplankton form a vital link in the trophic chain of any aquatic ecosystem. Their production and abundance can be directly linked with the potential of an area both for capture and culture fisheries. Apart from this, their composition and distribution provides interesting insights into many ecological concepts like community structure, species diversity, species coexistence and evolution. In the ensuing account it will be shown that the estuarine zooplankton have its own distinctive assemblages - different from the adjoining marine or fresh water environments - able to withstand the peculiarities nature has imposed on them.

India has several major riverine/estuarine systems (Fig. 1). Along the east coast are the Hooghly, Mahanadi, Godavari, Krishna and Cauvery. On the west coast we

have the Narmada and Tapi of the Gujarat and Mandovi-Suari estuaries of Goa. The Pulicat and Chilka lakes of the east coast and the Vembanad lake of the west coast form large bodies of brackish water. Apart from these, many medium and minor rivers (about 100) also contribute to the estuarine wealth of India. The rivers together have a catchment area of more than 3 million sq. km and a runoff of 1,600 million cu. m (Khosla, 1981; Rao, 1975). The total runoff of the major riverine systems of the east and west coasts of India is given in Table 1. Of this, the discharge from the rivers of the Kerala coast is estimated at 2.56×10^4 million cu. ft/year (Karunakaran, 1982).

The State of Kerala has many estuarine systems, the hydrobiology of which are greatly influenced by the monsoons. Of the tropical monsoonal estuaries of Kerala, the Cochin backwaters have received most of the attention of the scientific community. The present work on zooplankton of brackish water environs of Cochin was made more broad-based as data from seven other estuaries located along a 500 km stretch of the coast also came on hand. Thus this is the first attempt of its kind to study and compare the estuarine zooplankton of a coast receiving the full impact of both the monsoons. However, in carrying out sampling programs covering such a wide area several logistic problems arose and the observations had to be limited to one station each near

the mouth in the estuaries, except Cochin backwaters. In the following account, the distribution and other aspects of zooplankton from Cochin backwaters will be used as the spring board to evaluate the similarities and variations in the ecology of estuarine zooplankton in the other areas of study.

1.1. Definitions and classifications of estuaries.

Various definitions have been put forward by different authors for estuaries. Ketchum (1951) defined an estuary "as a body of water in which the river water mixes with and measurably dilutes sea water". Eury and Stevenson (1957) described it as the mouth of a river or an arm of the sea where the tides meet the river currents. They differentiated two types based on salinity and tidal characteristics.

1. 'Normal' type where due to river discharge salinities are reduced as one goes upstream.
2. 'Hypersaline' or negative estuary where exchange is poor and salinities are much higher than neighbouring sea. Various classifications are also put forward by Day (1951) and Redford (1951).

Fritchard (1952a) defined estuary as a semi-enclosed coastal body of water having a free connection with the open sea and containing a measurable quantity of sea salt. He classified the estuaries in terms of fresh water inflow and evaporation into (1) 'Positive' estuaries, where there is a measurable dilution of sea water by land drainage, (2) 'Inverse' estuaries where evaporation exceeds precipitation and (3) 'Neutral' estuaries where neither fresh water inflow nor evaporation dominates. But Fritchard later (1967) modified his original definition for estuaries as "a semienclosed coastal body of water which has a free connection with the open sea and within which sea water is measurably diluted with fresh water derived from land drainage". He prefers to reserve the term 'estuary' without any qualifying adjective to those bodies which he previously called 'positive estuaries'. This is probably the most recent and commonly recognized definition for estuaries.

A classification based on topography has been presented by Fritchard (1952b). He divides the estuaries into four groups: Drowned river valleys or coastal plain estuaries, fjords, bar built estuaries and estuaries produced by tectonic processes.

1. **Drowned river valleys or coastal plain estuaries are those which have been formed by drowning of former river valleys either from a subsidence of land or from a rise in sea level. They are usually an elongated indentation of the coastline with the river flowing into the upper end.**
2. **Fjords are generally 'U' shaped in cross-section most of them having a river entering at the head and exhibiting estuarine features in the upper layers.**
3. **Bar built estuaries result from the development of an offshore bar on the shoreline and have a relatively small channel connecting the estuary with the ocean. They are shallow within, and run parallel to the coastline with frequently more than one river entering the estuary.**
4. **Coastal indentures formed by tectonic processes like faulting or local subsidence having an excess supply of fresh water form another group.**

The original definition of Fritchard is further refined by Caspers (1967) who feels that it would include both estuaries and lagoons. He differentiates lagoons

as those having a stable body of brackish water whereas in estuaries the mixing of fresh and marine waters is not stable but shows periodic changes.

1.2. Review of the earlier work.

Considerable amount of work on the physical, chemical, biological and other related aspects of various estuaries have come out in the last 50 years. These studies have contributed to a better understanding of the physical and biological processes taking place in the estuarine systems.

Earlier reports on estuaries include studies on the South African estuaries by Day (1951, 1957); Day *et al.* (1952, 1954, 1956); Australian estuaries by Rockford (1951) and Chesapeake Bay by Hritchard (1952a, b, 1954, 1956). Contributions on estuarine hydrography, circulation, fauna and their ecology have come from Gordon (1960, 1963); Hury and Stevenson (1957), Hodgkoth (1957), Jeffrey (1962a, b, c, d), Netchum (1951, 1954) and Odum (1971). A treatise on various aspects of estuarine research and management by outstanding authors is presented in "Estuaries" (Lund, 1967).

Pioneering studies on the estuaries in India date back to the beginning of this century. The fauna of the Ganges delta was described by Annandale (1907), Alcock (1911) and Kemp (1917). Excellent studies on systematics and taxonomy have come from the Chilka Lake (Annandale and Kemp, 1918; Sewall, 1934). Some interesting work was carried out on the brackish water fauna of Madras area by Panikkar and Aiyar (1937) and Panikkar (1951) reviewed the physiological adaptations of animals. Godavari estuary has been studied in detail in 1964 (ICAR Report) by Chandra Mohan (1963) and Chandra Mohan and Rao (1972). Vellar estuary of Porto Novo has been studied by Sachaya (1959); Rangarajan (1959); Krishna Moorthy (1961), Ramamoorthy *et al.* (1965), Subbaraja and Krishna Moorthy (1972), Krishna Moorthy and Sunder Raj (1973), Devendran *et al.* (1974). Various aspects like hydrography, circulation, chemistry, phytoplankton, benthos etc. from Mandovi-Nezari estuarine system of Goa have been studied by Rao *et al.* (1972), Singhal (1973), Parulakar *et al.* (1973, 1980), Cheriyan *et al.* (1974, 1975), Bhargava and Divedi (1974), Gowami and Singhal (1974), Gowami and Selvakumar (1977), Rao (1976), Varma *et al.* (1975). Compiled information on the more recent works on the various aspects of estuarine biology has come from Natarajan (1973) and Murien (1977).

The backwaters of Cochin is one of the better studied estuaries in India. General hydrography of the estuary was studied by Ramamirthan and Jayaraman (1963), Durbishire (1967), Wallershous (1972), Haridas *et al.* (1973) and Srinivasa and Balakrishnan (1973). The tidal fluctuations were reported by George and Krishna Kartha (1963) and Gopin and Gopinathan (1969); solar radiation by Gopin *et al.* (1968); nutrient distribution by Sankaranarayanan and Gopin (1969), Joseph (1974) and Manikoth and Salih (1974); siltation by Gopinathan and Gopin (1971); sediments by Murty and Veerayya (1972 a, b) and Veerayya and Murty (1974); phosphate regeneration by Reddy and Sankaranarayanan (1972) and nanoplankton by Gopin *et al.* (1974). The organic production and phytoplankton ecology and related aspects have been studied by Gopin and Reddy (1967), Gopin *et al.* (1969), Gopin (1970) and Euvassy and Bhattachari (1974). Some work on the pollution problem in the estuary has been reported by Unnithan *et al.* (1973), Sargadevi *et al.* (1979), Resmi *et al.* (1980 a, b; 1981) and Venugopal *et al.* (1980). The changes in the ecology of the system brought about by man-made changes were reviewed by Gopin and Madhupratap (1979).

The general composition of the zooplankton of Cochin backwaters was published by George (1958). Some aspects of seasonal changes in zooplankton have been studied by Nair and Trenter (1971), Wallerstein (1974) and Nair *et al.* (1971). Distribution and ecology of some of the groups of zooplankton has been studied by various authors, such as hydromedusae by Senthilumari and Varma (1971); chaetognaths by Vijayaalakshmi Nair (1971, 1973) and Sreenivasan (1971); copepods by Pillai (1971), Pillai and Pillai (1973), Pillai *et al.* (1973) and species of the family Acartiidae by Trenter and Abraham (1971). An account of the taxonomy of copepods in the estuary is given by Wallerstein (1968, 1970), species composition and their seasonal fluctuations in the estuary by Madhupratap and Haridas (1978) and Rao *et al.* (1978). Studies on the tidal influence on the estuarine zooplankton, community structure and ecology of some species of copepods have come from Madhupratap (1978, 1979, 1980). Magnitude of secondary production in the Cochin backwaters has been studied by Madhupratap *et al.* (1977).

Studies on the other estuarine systems along the Kerala coast are scanty and discontinuous. Some preliminary hydrobiological and planktonological investi-

gations in the river mouth at Korapusta estuary were made by ^{Suryanarayana Rao} Rao and George (1959), George (1953 a, b) and Varkey John (1971); at Bhyppore by Varkey John and Alexander (1968). Recently Murugesan et al. (1980) have studied the benthic fauna of Veli Lake. Mathew and Balakrishnan Nair (1980) have studied the phytoplankton of Ashtamudi (Neendakara) estuary. The numerous other estuaries of Kerala coast have received little attention.

2. MATERIALS AND METHODS.

2.1. AREA OF STUDY.

Kerala with a coastline of about 600 km is endowed with numerous and extensive brackish water bodies, mostly running parallel to the coastline. Eight major estuarine systems along this coast, covering a distance of 200 km were investigated over a period of one year during 1978, namely Veli at Trivandrum, Neendakare at Quilon, Thattappally near Alleppy, Cochin backwaters at Cochin, Kozhuppuha, Kallai and Neyyore at Calicut and Maho near Thalicherry (Fig. 2).

Veli Lake:

The Veli lake is primarily a fresh water body and is one of the smallest of the lakes confined to the southern part of the Kerala State, situated 5 km north-west of Trivandrum. It is 1 km long and 0.3 km broad. A narrow strip of sand bar across the mouth of this lake during the dry season obstructs free exchange of marine and fresh water. The lake is very shallow, depth being 1 to 2.5 m. The bottom is sandy. Observations were made near the sand bar on the lake side.

Mandakara:

The estuary at Mandakara is situated about 9.6 km north of Cochin at 9°28' - 8°45'N lat. and 76°28' - 77°17'E long. This estuary is one of the foremost centres of marine fish landing along the southern Kerala coast. It is also known as Achamudi estuary. The point of observation was near the mouth and in this area, the water is always churned up by the propellers of fishing boats and tidal action. The depth at the area of observation is 4 m. The bottom is muddy sand.

Thattamully Lake:

This lake is situated about 25 km south of Alleppy. There is a narrow strip of sand bar across the mouth of this lake. A barrage is constructed on the eastern side in the lake about 0.5 km away from the sand bar to prevent incursion of saline water into the hinterlands so as to enhance paddy cultivation. The spill ways are opened during the monsoon period. The samplings were made still further on the eastern side of the barrage.

Cochin backwaters:

The Cochin backwaters include a system of interconnected lagoons penetrating the main land and enclosing many islands in between, whose tidal area amounts to approximately 500 sq. km. The backwaters around Cochin is located along $9^{\circ}58'N$, $76^{\circ}15'E$. It has a permanent connection with the Arabian Sea on the western side by a channel about 450 m wide which forms the entrance to the Cochin harbour. At the northern extremity it opens into the Arabian Sea at Ashikode and at the southern end it terminates into a large body of fresh water namely Vembad lake.

The coastline is of an emergent type formed of a number of narrow sand bars running parallel to the coastline often in several rows (Dashidisa, 1967). The sand bar and the harbour channel at Cochin are periodically dredged to accommodate the traffic of the port. The channel area around the mouth where observations were made is about 15 m deep. The depth of the estuary gradually reduces further upstream to about 2 m at Alleppy. The depth of the area north of the bar mouth is uniformly shallow, being 2 to 4 m. The bottom of the estuary is generally muddy.

The major source of fresh water in the estuary are the two large rivers namely Pamba on the south and Periyar on the north. In addition to these, Mannathil and Muvattupuzha rivers and several small tributaries and irrigation channels join the backwaters.

The tidal effects of the estuary is maximum near the mouth, the amplitude of the tides being about 1 m at the harbour area decreasing towards the upper reaches. Ingression of sea water to the upper reaches oscillates depending on the fresh water efflux (Narayan *et al.*, 1973).

Karapuzha estuary:

The Karapuzha estuary is shallow and 32 km long. Two rivers feed the estuary. The Klothur river joins the Karapuzha backwater system about 1 km away from the mouth. Another stream running from the foot of the high mountain range surrounding the Nedimada Malai empties into it about 16 km from the river mouth. The depth of the estuary is 4 to 5 m and the bottom is sandy. During monsoon the silt laden fresh water run down the river suppressing the ebb tide almost completely.

Kallai estuary:

Kallai estuarine system has its origin as a small stream from Kunnemangalam and flows down through Kunnethupalam and Marthava to Kallai town and then joins the Arabian Sea. It has a length of 30 km. All along the river banks there are large numbers of coir cottage industry units and considerable amount of coconut husks are dumped into the river for retting. These husks get sedimented at the bottom and due to organic decay of these, a black layer is formed on the substratum mixed with sand. Further, large number of logs are kept in this river for conditioning all through the year. These logs also foul the water. During the rainy season water level rises up by 2 to 4 m in this river. In fact torrential flow of fresh water from uplands cleans the river seasonally. Because of the small size of the river the tidal currents enter upstream into the river by about 18 to 20 km.

Beypore estuary:

Beypore estuary has its source principally in the Nilgiri Hills and flows down as a major river through Nilambar, Mambad, Arikkod, Mavur, Kanniyanparanku, Furoku and Beypore and empties into the Arabian

Sea. The rivers Cherupuzha and Iriinjipuzha open into this estuary at Kannimparumba and Muvur respectively.

The river Chaliyar which receives the effluents from the Rayon factory at Muvur also joins this estuary. During monsoon the water level rises up by 4 to 5 m and the water becomes turbid. The bottom is sandy.

Maha_naduvayal

River Mayyazhi (Maha) has its origin in the Western Ghats. The drainage basin has similar characteristics as those of other three rivers of north Kerala described earlier. It enters the Arabian Sea at Maha, a small Union Territory and part of Pundicherry State. During the monsoon seasons the water level at the point of sampling rises by about 1.5 to 2 m. The bottom is muddy sand and has an average depth of 5 m. State net fishery operates here almost round the year. It has a small fishing harbour and is a landing centre for the green mussel Mytilus yamadai collected by local divers. This river also has setting yards in it.

2.2. Methods.

All observations were conducted monthly in the year 1978. Observations were made at one station near the mouth at all the estuaries except Cochin backwaters. In the Cochin backwaters apart from collecting monthly information at a station fixed at the mouth, the distribution of zooplankton to the interior of the estuary was also studied. For this purpose zooplankton and environmental characters were observed from 6 more stations in four months, representing various seasons, viz. January (early monsoon), April (peak saline period), July (monsoon) and November (postmonsoon). The stations were fixed to cover the entire backwaters from its mouth (at Cochin) to its head (Alleppy) (Fig. 3).

2.2.1. Sampling procedure:

Water samples were collected from the surface with a clean plastic bucket and from mid depth and bottom using a water sampler for the estimations of hydro-graphical parameters. The temperature was recorded immediately after sampling using a thermometer. Salinity

was estimated using Mohr-Nelson titration method and oxygen by Winkler's technique. Zooplankton was collected by oblique hauls from the bottom to surface using an HF net (Hagen-Grantner net, mouth area 0.25 m^2 , length 3 m, mesh size 300 microns).

A flow meter (Rigoch Model No. 850) was attached to the mouth of the net to estimate the volume of the water filtered. The net was gradually drawn from the bottom to the surface, the duration of haul being 5 minutes. Zooplankton was preserved in 5% formaline.

The plankton samples were filtered, drained of excess water using absorbent paper and added to known volume of water to find out the displacement of volume. Subsamples were often taken for the analysis using a Folsom Plankton Splitter. Large organisms like hydrozoans, sponges and chaetognaths were removed and counts taken for the whole sample. The subsample was then spread on a counting tray and counted to species level as far as possible. The whole sample was used for counting whenever the sample was small. The counts were then transformed into numbers per unit volume of water filtered using the flowmeter data.

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3. HYDROGRAPHY.

Assessment of environmental features and their changes is essential for understanding the ecology and inter-relationships of the organisms inhabiting an area. Several studies mentioned earlier have contributed to the understanding of hydrobiological aspects of Indian estuaries. The State of Kerala gets the rainfall from both Southwest and Northeast monsoons and they have profound influence on the hydrography and biology of the estuaries. The seasons can broadly be divided (although this is somewhat arbitrary, since the onset of rainfall varies from year to year) into pre-monsoon (January to April), monsoon (May to October) and post-monsoon (November-December) periods. However, at the mouth regions of most of the estuaries saline conditions prevail at the beginning of the monsoon period also (May-June) although the salinity towards the interiors is brought down.

3.1. Gulfia_hackberry.

3.1.1. Salinity:

The most important factor controlling the biological processes in tropical estuaries is the monsoon and the associated changes in salinity. Wide variations were observed in the salinity structure of the water column in this estuary during the different seasons (Fig. 4 A).

Pre-monsoon:

A more or less vertically homogeneous pattern in salinity distribution was observed by the beginning of pre-monsoon. Salinity values were high ($> 30‰$) both at surface and bottom. It steadily increased through the season and registered the maximum value (34.8‰) in April. The influence of the saline water could be traced upto the head of the estuary (Fig. 4 B). The recovery after the monsoon was gradual and the values towards the head increased from 3.0‰ in January to 13.0‰ by April. Some amount of stratification occurred at the middle reaches during the early pre-monsoon. But the water column became well mixed

during the peak monsoon period.

Monsoon:

The onset of monsoon in May brought about a total change in the physical characteristics of the environment. Large quantities of fresh water were discharged into the backwaters through the rivers and land runoff during the monsoon. During the year of observation (1978) rains started from May and lasted upto October with intermittent breaks.

The surface salinity fell from 34.8‰ in April to 9.8‰ in May. Bottom salinity varied from 36‰ to 33.8‰ at mouth of the estuary. The water column became stratified showing a two-layered flow. In June an increase in salinity was observed both at the surface and bottom due to a short break in the monsoon. Bottom salinity reached 38.8‰, which was the highest value recorded. The presence of this high saline water in the bottom layers was probably due to the intrusion of upwelled Arabian Sea water into the channel (see 3.1.2) during this period. Surface salinity reached near zero values in July when the monsoon was at its peak. It showed an increasing trend in August and September owing

to an abatement of the force of the monsoon but decreased again in October when the rainfall increased. Towards the upper reaches the estuary remained fresh water dominated throughout the monsoon (Fig. 4 B).

Postmonsoon:

By November surface salinity started to show an increase. The water column continued to be stratified with high saline water (32.5‰) at the bottom. Sea water started to dominate by December, stratification being less apparent, the conditions leading into homogeneous situation of the premonsoon.

Salinity recovery was rapid at the mouth and gradual at the upper reaches. Even after the rain fall was decreased, the fresh water flow was still strong enough to restrict the incursion of marine water towards the head of the estuary during the postmonsoon period.

3.1.2. Temperature:

Fluctuations in temperature were not much and as pronounced as that of salinity but the seasonal changes were reflected in the temperature structure also.

Pre-monsoon:

Temperature was higher during the pre-monsoon season. In January the surface temperature was around 28°C. There was a gradual increase as the season progressed and by April the surface temperature reached 31.6°C. The water column was well mixed and homogeneity prevailed with little difference in temperature between the upper and bottom layers (Fig. 4 A).

Monsoon:

The effect of monsoon was reflected in the temperature distribution also. Temperature suddenly dropped from 31.6°C in April to around 26.0°C in May at both surface and bottom. It increased to 29.8°C in June corresponding to a temporary break in monsoon and fluctuated between 28°C and 30°C during the monsoon season. Steep vertical gradient in temperature existed during this season. Lowest temperature (24.4°C) was recorded at the bottom in July. The difference between the surface and bottom temperature fell within the range of 1.6°C to 2.1°C from June to October.

Thus, during the monsoon period, especially in July-August, thermal stratification was very steep. The cold high saline water at the bottom layers is probably the upwelled Arabian Sea water entering the channel. Presence of cold dense waters upwelled from the sub-surface levels of Arabian Sea in the continental shelf and its incursion into the north area of Cochin backwaters has been reported during these months earlier (Ramesh, 1959; Ramakrishnan and Jayaraman, 1960, 1963; Venugopal et al., 1979).

Discussion:

During this season, temperature differences between the surface and bottom became less sharp and by December the water column became more or less homogeneous.

3.1.3. OXYGEN:

The pattern of oxygen distribution was also similar to that of salinity and temperature. During pre-monsoon, the surface and bottom oxygen content did not show any significant variations and the values ranged between 2.0 ml/l to 3.0 ml/l.

Surface oxygen content showed a general increase during the monsoon period. It increased to 4.0 to 5.0 ml/l in July and August. But the bottom values decreased during this period, the minimum observed being 0.5 ml/l in August (Fig. 4 D). The very low oxygen content at the bottom layers during July-August was further evidence to the presence of upwelled water. The low oxygen content generally found in the bottom during monsoon period is probably owing to the high turbidity and low light penetration (Gosia *et al.*, 1960; Venugopal *et al.*, 1979) limiting primary production in these layers.

During the postmonsoon period, the surface and bottom oxygen values were more or less same and the water column fairly homogeneous. However the surface values were still less, compared to the monsoonal period.

3.2. Comparison with other estuaries.

The hydrographical characteristics of Cochin backwaters is typical of other estuarine systems of Kerala coast. In all these estuaries monsoon is the key factor bringing about the annual cyclic changes in the hydrobiology of the environment. As mentioned

earlier, the most important parameter controlling the biological processes in tropical estuaries is salinity. The large quantities of fresh water which inundate the estuaries during the monsoon often abruptly alters the salinity structure and causes total transformation in the faunal composition.

The basic difference in the physical characteristics of Cochin backwaters with other estuaries studied is that the Cochin backwaters form a large basin of brackish water into which several rivers empty. The other estuaries apart from Veli and Thottappally (which are primarily fresh water lakes having limited periodic connection with the sea) are river mouths where estuarine conditions develop. More over the channel area of Cochin backwater system is constantly dredged and deepened to accommodate port traffic while the mouth of other estuaries are comparatively shallower (some dredging operations are conducted periodically at Kaypara and Koodakara also). However, the general hydrography at the mouth regions of Koodakara, Kallai, Kaypara, and Mahé estuaries is more or less similar to Cochin Backwater system and hence only the salient features are discussed.

3.2.1. Comparison of Mandakara, Kallai, Baypara, Koraputa and Maba estuaries.

Salinity:

Salinity distribution in these estuaries was similar during the premonsoon period. At Mandakara the surface salinity ranged between 31.6‰ to 34.2‰, while at the bottom it varied between 32.1‰ to 34.6‰ (Fig. 5). At Kallai maximum salinity of 35.7‰ was recorded in April (Fig. 6). At Baypara, the salinity after registering a maximum (33.3‰) in February fell during the subsequent months of the season and at the peak of premonsoon, in April, it was only 29.2‰ at bottom (Fig. 7). The conditions were similar at Koraputa and Maba estuaries also. The water column was well mixed and the values ranged between 30‰ to 32‰ during the premonsoons (Figs. 8 and 9).

Salinity fell with the onset of monsoon in May. At Mandakara it was reduced from 34.2‰ in April to 16.9‰ at the surface and 18.6‰ at the bottom (Fig. 5). Similar decrease was noticed at Koraputa, Kallai, and Maba but the salinity remained high at Baypara. A slight reduction in the force of monsoon on the southern

parts of Kerala in June resulted in a temporary rise in salinity in the estuaries at Cochin, Mandakara, Veli and Thottappally. But again it fell to near zero values both at surface and bottom in all the estuaries with the heavy rains in July. But at Mandakara the surface and bottom salinities were reduced only to 6.6 and 14.6‰, respectively in July probably because of the lesser rainfall in this area.

Vertical stratification during monsoon was less pronounced in these estuaries as compared to that in Cochin backwaters. However, a lesser gradient was observed at Kallai, Neyyore and Karapuzha during the later part of the monsoon period (Figs. 6, 7 & 8). Virtually no stratification occurred at Mandakara and Mabe estuaries (Figs. 5 & 9). The probable reason for the less marked stratification compared to Cochin backwaters is that the sand bar at the mouth of the latter is deepened periodically allowing free incursion of sea water along the bottom as a typical salt wedge.

Because of the same reason, the salinity recovery in some of these estuaries was slower in the early post-monsoon period (November) compared to Cochin backwaters. The regional variations in the rainfall also influenced this to some extent. By December the salinity started increasing in all the estuaries.

Temperature:

Temperature also fell in tune with the pattern of salinity distribution (Figs. 2-9). In general, it fell by 7.0 to 8.0°C in monsoon compared to the pre-monsoon values. But a sharp gradient in the vertical thermal structure was not apparent in these estuaries unlike Cochin backwaters except at Kallai (Fig. 6). The differences between surface and bottom in general were only about 1.0 to 1.5°C. At Kallai the highest gradient observed was 3.5°C in August.

Oxygen:

Oxygen content at the surface layer in these estuaries was generally higher compared to Cochin backwaters. At Mandakara it fluctuated between 3.3 ml/l to 3.8 ml/l during the pre-monsoon. It further increased to 4.9 and 5.3 ml/l in August and November (Fig. 5). The bottom values did not vary much from the surface values during early monsoon period. But from August a difference of about 1.5 ml/l was observed between them. The distribution of surface oxygen at Kallai was different. From 3.3 ml/l in April it increased to 4.7 ml/l in June. Values then decreased steadily during the monsoon season

and registered the minimum in October (Fig. 6). The bottom values showed similar trend of distribution as that of surface without much difference except in April and May. A steady increase in the surface oxygen content was noted at Sappore estuary from the pre-monsoon to monsoon period (Fig. 7). The maximum value 8.5 ml/l was observed in August. Here also the bottom oxygen did not show any significant variation from that of the surface. The oxygen distribution did not exhibit any definite trend which could be correlated to seasons at Marayunta and Lake where high and low values occurred in all seasons (Figs. 8 & 9).

One notable aspect emerging in the comparison of the hydrographic parameters is the absence of upwelled Arabian Sea water in any of these estuaries during the July-August period unlike the Cochin backwaters probably owing to the shallower nature of their mouths.

3.2.2. Comparison of Veli and Thattamvilly lakes.

These two lakes on the southern part of the Kerala coast differ from the other estuarine systems in their hydrobiological aspects. The Veli lake has a sand bar across its mouth which permits exchange of fresh and

marine waters only for a short span of time. At Thottappilly a man-made barrage obstructs free inflow of water from the sea (see 2.1).

Salinity:

At Veli the surface salinity was low throughout the year. Maximum value recorded was 4.5‰ in November. During the premonsoon the values ranged from 0.7‰ to 3.2‰. Bottom salinity was higher during this period, registering the maximum in March (23.8‰) (Fig. 10). It fell to near zero values during monsoon and the water column became practically fresh. During the post-monsoon the bottom salinity increased to 23.4‰, while the surface values remained low. Narayan *et al.* (1980) have also observed low salinities in this lake but maximum value they recorded was only 4.8‰.

The Thottappilly lake was dominated by fresh water throughout the year. The salinity was very low, always less than 2‰, during all the seasons except in January when the bottom salinity was 7.8‰ (Fig. 11).

Temperature:

High temperature prevailed during the pre-monsoon at Veli. From 31.2°C in January it increased to the maximum of 34°C in March (Fig. 10). During the monsoon the values fell and fluctuated between 25°C and 30°C. The minimum was observed in October. During post-monsoon the surface temperature was around 30°C. The bottom values did not show any variation from those of the surface during all seasons since it is a shallow area (2.5 m).

The temperature distribution was almost similar at Thottappally also with maximum temperature (33.0°C) being recorded in March and April at the surface and minimum (26.5°C) in July at the bottom (Fig. 11). The bottom temperature was only slightly less compared to the surface except in July and October.

Oxygen:

The surface water at Veli had high oxygen concentrations throughout the year ranging between 4.0 ml/l to 5.9 ml/l except during early pre-monsoon period (January and February). Oxygen in the bottom layer was lower in most of the months, maximum difference being recorded in August (Fig. 10).

Oxygen distribution at surface at Thottappilly was more or less similar to the Veli lake (Fig. 11).

The major difference of Thottappilly and Veli lakes from the other estuaries is the presence of low saline water at the surface throughout the year, while at Thottappilly the bottom salinity also remains low more or less throughout, salinity goes quite high at Veli lake in part and pronounced seasons. The origin of this high saline bottom water at Veli although there is no free connection with the sea is subject to some speculation.

It is perhaps best explained in terms of the dynamics of a coastal aquifer (Glezer, 1959; Cooper, 1959). It involves ocean tides, the rise and fall of the ground water table and the bed of the sand bar acting as a permeable medium. A zone of diffusion is formed in the bed and sea water and fresh water become intimately mixed in this zone. To quote Cooper "it appears to be reasonably certain that wherever a zone of diffusion exists in a coastal aquifer a flow of sea water from the floor of the sea into the zone of diffusion will occur. The flow may be interrupted or reversed during low stages of tide or high stages of the water table but on the average it will persist in a land ward direction".

Thus the salinity increase in the bottom layer of Veli lake may be through this diffusion, occurring along the bed. (However, the presence of some typically coastal species in the Veli lake in some months - see Chapter 4 - makes it impossible to arrive at a firm conclusion. There could also be a possibility of some water spilling over during intense wave action and sinking to the bottom of the lake).

At Thottappally the configuration of the terrain where collections were made is different. The barrage is built about 0.5 km away from the sand bar and the collections were made from inside the spillway. The distance from the sea to the place of collection may account for the lower salinity at the bottom encountered at this station.

4. ZOOPLANKTON.

Estuaries are transition areas between the more stable conditions of neighbouring sea and fresh waters and exhibit increased gradients and fluctuations of abiotic and biotic factors (Nixon, 1967). The physico-chemical conditions and their fluctuations in an estuary are determined by the tide, the quality and quantity of the river water discharged and the morphology of the area. The unpredictability of these factors render estuaries physically controlled rather than biologically accommodated habitats (Sanders, 1968). The zooplankton occupying this biotope have to be tremendously accommodative to put up with the stress. Thus, true estuarine organisms form a class by themselves apart from the more common euryhaline marine forms, and to a lesser extent stenohaline forms and fresh water organisms which frequent these waters.

The composition, distribution and abundance of various groups and species of zooplankton in the eight estuaries studied are presented in this chapter. Three way analysis of variance was performed to study the significant differences between groups and species,

area and season (Drozdica, 1960; Fisher and Yates, 1957).

The model used for the analysis was

$$Y_{ijkl} = \mu + \alpha_i + \beta_j + \gamma_k + \delta_{ij} + \delta_{ik} + \delta_{jk} + \epsilon_{ijkl}$$

where

- μ = Grand mean
- α_i = i^{th} species effect
- β_j = j^{th} station effect
- γ_k = k^{th} month effect
- δ_{ij} = interaction between i^{th} species and j^{th} station
- δ_{ik} = interaction between i^{th} species and k^{th} month
- δ_{jk} = interaction between j^{th} station and k^{th} month

ϵ_{ijkl} = The deviation of the cell mean from the values expected on the assumption that they would be the grand mean plus the species, station and months effect plus the three first order interactions and ϵ_{ijkl} = random effects which are normally distributed with mean = zero and variance = σ^2 .

4.1. Zooplankton biomass and abundance.

Monthly biomass and total number of zooplankton in the eight estuarine systems are listed in Table 2 (A) and Table 2 (B). Analysis of variance (Table 3 A and Table 3 B) showed that significant variations between seasons as well as between estuaries existed for biomass and total zooplankton counts. Maximum biomass occurred in April and May followed by February and March. Minimum was observed in August-September. Maximum counts were also in May and April and July, August and September showed the minimum. Cochin backwaters topped the list in both biomass and total counts. Kappuzha and Kallai ranked next and minimum abundance was observed at Thottappilly. Higher abundance of zooplankton observed in May apart from premonsoon months is because the observations were made at the north area of the estuaries where salinity was fairly high in this month also. Also, many medium saline species shift to this area when salinity becomes too low in the interiors of the estuaries.

Maximum abundance in Cochin backwaters was noticed in April (8.2 ml/10 m³ and 10,4107 nos/10 m³). Kallai which ranked next in the abundance showed only 3.5 ml/10 m³ (total number - 30297 nos/10 m³) during this month.

In the other estuaries maximum population was usually in April, however the month when peak biomass was recorded varied. This also showed that the total numbers and biomass are not ~~strongly~~ correlated since if larger organisms like mysids and sergestids or other decapod larvae are present, the picture of the biomass can be very much altered. The average values for pre-monsoon for biomass varied 0.005 and 4.9 mg/10³ and zooplankton counts from 200 to 40000/10 m³ at Thottappally and Cochin backwaters respectively.

Biomass values fell sharply during the monsoon period. Averages for biomass fell by 67% at Cochin, 37.5% at Kallai, 19.2% at Kuzhupata, 72.5% at Bappara, 90.5% at Naha, 74.5% at Veli and 40% at Thottappally when compared to the premonsoon. Similar decrease could be observed in the total numbers of zooplankton also.

It may be mentioned that the zooplankton counts and biomass at the mouth of an estuary during monsoon period could be sometimes a little deceptive. The trend of the values in Table 2 A and Table 2 B do not strictly conform to the general idea that zooplankton become scarce in the estuaries during the monsoon. This

is because at the mouth area even a slight let up in rain or river discharge can result in moderate salinity recovery (especially during high tide) bringing in the higher saline estuarine zooplankton elements. Also, often many medium saline species are able to thrive in this area, atleast for short durations during this season, as would be shown in the forthcoming sections. Regional differences in rainfall, depth at the mouth, and general configuration of the estuary could also lead to these variations. A true picture would emerge when the abundance of zooplankton inside the estuary also is taken (Table 2 C) which shows sparse population during this season.

During the postmonsoon months (November-December) most of these estuaries with a free connection to the sea showed an increase in salinity as well as in zooplankton population. But the recovery was still slow towards the upper reaches.

Results of the three way analysis of variance (Table 4 A) showed that among the major groups of zooplankton Copepoda was significantly higher in numerical abundance followed by naup larvae and sargastids and the minimum was observed for Ctenophora and Ctenocera.

All the first order interactions between seasons, areas and groups were significant at 2% level. Comparatively maximum abundance of zooplankton was in Cochin backwaters followed by Kallai, Karapuzha, Beypore, Veli and Maho. Least abundance was noticed in Thottappilly and Neendakara estuaries. In all the estuaries, Copepoda was the dominant group, maximum density being obtained usually in April except at Beypore, Neendakara and Thottappilly where they occurred more in November, December and October respectively.

4.2. Composition and distribution of zooplankton.

The zooplankton of these estuaries comprised of various groups belonging to almost all phyla. Altogether sixteen major groups were identified namely hydromedusae, siphonophora, Ctenophora, Chaetognatha, Copepoda, Ostracoda, Cladocera, Cumacea, Isopoda, Amphipoda, Mysidacea, Sargostidae, invertebrate eggs and larvae, fish eggs and larvae, Copepoda and Thalassia. The distribution of total zooplankton, common groups and species in the estuaries are given in Figs. 12-27. Seasonal distribution of major groups in the estuaries is presented in Table 5. (The distribution in May and June is treated separately from other months of

monsoon period, since saline conditions prevailed at north areas of most estuaries in these months and often a mixed assemblage of high and medium saline species was encountered).

Seventy three species from various groups were identified from Cochin backwaters. Of these, 47 belonged to Copepoda. The biomass and total number of zooplankton were lower in Mandakara estuary when compared to others. However, 44 species belonging to various groups occurred in this estuary and was next to Cochin backwaters in the number of species. This is followed by Valli (39 species), Marayur (36), Koyyare (34), Malini (29), Mhu (28), and Thottappally (24). The systematic list of species and the presence and absence of these in various estuaries are given in Table 6. The general distribution of groups and species which are not common and not represented in the figures are given in Table 7.

4.2.1. Hydromedusae:

These carnivorous organisms are all high saline forms and occur in estuaries principally during inter-monsoon period. Santhakumari and Vannucci (1971) recorded nineteen species of hydromedusae from the Cochin backwater system. They have reported some species of

hydromedusae in the Cochin backwaters to be endemic to this area. Only six common species namely *Blackfordia virginica*, *Eutima gummucalis*, *E. punctulidensis*, *Eutima gummucalis*, *E. gummucalis* and *Eutima gummucalis* were identified in the present study. Among these *Eutima gummucalis*, *Blackfordia virginica* and *Eutima gummucalis* were found to be most abundant in the estuary and agrees with earlier findings of Vanucci *et al.* (1970) and Madhupratap and Haridas (1975). These three species together accounted for 92% of the numerical abundance of hydromedusae in the Cochin backwater system. All these species showed higher abundance in May and June at the north area (340 and 430 /10 m³ respectively) (Fig. 18). They were absent during monsoon and postmonsoon period except for the sole appearance of *Eutima gummucalis* in December.

The common hydromedusae species occurred in large numbers in the middle and upper reaches of the estuary during premonsoon period (Fig. 17). By April they were present right upto the head (Station 7) and peak density of 1000/10 m³ was observed in the middle reaches. These species exhibited considerable salinity tolerance and while *Eutima gummucalis* occurred only in salinity above 18‰, *Blackfordia virginica* and *Eutima gummucalis* occurred in salinity as low as 8‰. (Fig. 20).

Hydromedusae were completely flushed out of the backwaters by July. The three common species made their first appearance in the middle reaches of the estuary in the early premonsoon (January) period (Fig. 17), when the salinity recovered. Their population shifted to the mouth only later by March-April. Their presence in the middle of the estuary during the early premonsoon supports the view of Venkatesi *et al.* (1970) that the hydroids of the hydromedusae undergo a quiescent stage to tide over the unfavourable low saline environment and become active when the salinity conditions become optimal.

Hydromedusae had a less diverse population in the other estuaries. Only three species viz. *Eutima communalis*, *Eutima arvensis* and *Blackfordia virginica* were usually observed in these waters. *Eutima communalis* was the most abundant species among them. All these species were encountered in high saline months. Higher densities were usually observed in April and May. At Houndkera the distribution of hydromedusae was somewhat different compared to other estuaries. Here, the species *E. communalis* was observed by September and the other two species appeared a little later (*E. arvensis* in October and *E. virginica* in November - Fig. 20).

They occurred throughout the postmonsoon season in fairly good numbers. This is probably because the conditions were favourable for them since salinity recovery was faster at Mandakara both at the surface and bottom due to lesser rainfall after July. Even by September salinity was around 32‰ throughout the water column. It fell in November due to a spate of heavy rains but it did not affect the density of hydro-medusae at Mandakara.

These three species were observed at Maropasha and Bayyare during the premonsoon period and also in December. *Eutima gummucalis* and *Eutima sylvianensis* were the two species which were common at Mahe and Kallai estuaries. Maximum population of *Eutima gummucalis* was observed in April at Bayyare (3440/10 m³), *Eutima sylvianensis* in the same month at Kallai and Maropasha (210/10 m³) and *Blackfordia virginica* in March at Bayyare (100/10 m³) (Figs. 19 & 20). *Eutima gummucalis* was observed at Mahe in fairly good numbers in the premonsoon period (112/10 m³) (in April).

These three common species occurred sparsely in April at Thottappilly (12/10 m³). At Veli lake they were observed in April and May (maximum density - 300/10 m³, Table 7).

The three common hydromedusae species which occurred in these estuaries are essentially brackish water forms. *Alaskiella virginica* is a euryhaline species usually found in temperate and tropical estuaries and swamps and even in Caspian Sea and has a scattered geographical distribution (Vannote et al., 1970). *Limnopsis exilis* also attain peak densities in the estuaries though it has been recorded from coastal waters of Travancur (Nair, 1951) and Bombay (Lala and Das, 1935). *Limnopsis* is confined to the estuarine waters only. The other species which occurred at the mouth of estuaries are neretic.

The average annual density of hydromedusae varied from $442/10 \text{ m}^3$ at Neyyare to $2/10 \text{ m}^3$ at Thattappilly. The numerical abundance of hydromedusae is much less compared to groups like copepods (they formed only 1.6% of the annual total counts from all estuaries). Nevertheless, the ecological dominance exerted by the highly prolific groups like hydromedusae, Ctenophora, and Chaetognaths cannot be overlooked. These groups flourish only when their food (herbivorous zooplankton like copepods) are abundant. Higher numerical abundance of these groups often drastically reduce the copepod population in the particular area. At Neyyare, in April,

when hydromedusae and ctenophores together constituted 63.4% of the total zooplankton population ($6814/10\text{ m}^3$), the density of copepods was only $1033/10\text{ m}^3$ (9.6% of the total counts). A similar situation was observed in May also. But in March when density of hydromedusae was only $390/10\text{ m}^3$, the copepods constituted 61% of the zooplankton population. Earlier observations (unpublished data, Estuarine Survey Project, Regional Centre of National Institute of Oceanography, Cochin, 1975) also showed that copepod densities were drastically reduced when there was a towering abundance of hydromedusae and ctenophores. Copepod densities were 11 and $381/10\text{ m}^3$ at two stations where that of hydromedusae were 1930 and $2050/10\text{ m}^3$ respectively. However, at a nearby station copepod density was $7392/10\text{ m}^3$ where the density of hydromedusae was $37/10\text{ m}^3$.

4.2.3. Siphonophora:

Two species, *Rhizoma shaniannaia* and *Lanina subuloides* occurred in these estuaries except Veli and Thottappilly. At Cochin, Mandakara and Bypore they were observed during the postmonsoon months (Table 7). Both the species were present at Cochin during this period. Only *Rhizoma shaniannaia* occurred at Bypore ($22/10\text{ m}^3$ in November)

while *Leania subuloides* was observed at Mandakara (26/10 m³ in November). At Maho L. *subuloides* was observed in May. Maximum abundance of this group was noted at Kallai (226/10 m³ in April) comprising both the species.

These two species are common in the inshore waters of India (Daniel and Daniel, 1963; Ranganathan, 1973). These species do not propagate in the estuaries and their occurrences at these river mouths is purely accidental. Obviously they are stragglers into the estuaries.

4.2.3. *Stomatopoda*.

Stomatopoda *gibbosa* and *gibbosa* sp. were the two species that occurred in these estuaries. They did not occur at Valli and Thottappilly. *S. gibbosa* was the common species and their distribution was similar to that of hydromedusae (Figs. 13-15) being abundant during high saline months. Peak abundance was noted at Bayyare (3226/10 m³) and Karapuzha (1916/10 m³) in April. This species also occurred in large numbers in the middle and upper reaches of Cochin backwater system during premonsoon period. (Maximum density - 2180/10 m³ in April at Station 4).

Limnaea sp. was observed in small numbers at Cochin and Mandakara during high saline months.

4.3.4. Chaetognaths.

This group was common in the estuaries during the saline period. In the Cochin backwater system maximum abundance of Chaetognaths was noted in May ($200/10m^3$). Four species, *Limnoria gillata*, *L. helveta*, *L. gossia* and *L. robusta* were identified from this system. Two more species - *L. pulchra* and *Limnoria pacifica* have been recorded earlier from this estuary (Vijayalakshmi, 1971; ^{Nair} Sreenivasan, 1971). Of the four species encountered *L. helveta* was the most common with peak abundance in May ($190/10m^3$). *L. gillata* was more common during post-monsoon months (maximum density $60/10m^3$ in December - Fig. 18). *L. gossia* appeared during the peak saline months of March and April and dominated the chaetognath population in this period. *L. robusta* was observed in June and December in small numbers.

During the monsoon the chaetognaths were washed out of this estuary. With salinity recovery they appeared at the mouth in low numbers by November and gradually spread further towards the head along with the salinity

incurion (Fig. 17). By April they were present at the head, though in low numbers.

Chaetognaths were less abundant in other estuaries. They were totally absent at Thottappilly and were poorly represented at Kallai. *Squilla onflata* and *S. hutchini* were the two species commonly observed in other estuaries. In Kumbalangi estuary *S. hutchini* was the common species and occurred throughout the year except January, June and December while *S. onflata* was observed in low numbers during the postmonsoon season. *S. gnamptis* was recorded from this estuary in April. *S. hutchini* occurred in all the months except during the low saline period from June to October at Kayyore (peak density - $124/10m^3$ in January - Fig. 18) and in premonsoon months and June at Mabe (peak density - $372/10m^3$ in April - Fig. 18). At Kozhupattu *S. onflata* was the more common species during the premonsoon while *S. hutchini* dominated during postmonsoon. *S. onflata* occurred only in a couple of months at Kayyore and Mabe. The two species were present in April and May at Valli.

The three carnivorous groups viz. hydromedusae, ctenophora and Chaetognaths together constituted only 4.3% of the total zooplankton counts. Nevertheless, as mentioned earlier they have profound influence on the

population of other zooplankton groups, especially Copepoda.

Chaetognaths are exclusively marine forms and all the species recorded in the present study from the estuaries are common in the inshore waters of India. However, the species *Paragastrea indica* has been observed to breed in the Cochin backwater system during high saline period (Nair, 1973). This species is by far the commonest chaetognath occurring in the estuaries. In the Cochin backwaters, *P. guilfordi* was more abundant in postmonsoon season whereas *P. indica* dominated during early premonsoon. In the peak saline period of March-April *P. guilfordi* was the dominant form. No such clear pattern in distribution of chaetognaths was discernible in the other estuaries. The Chaetognaths occurring in the estuaries are probably only extensions of their population from adjoining inshore waters and the three common species recorded in the estuaries are obviously able to withstand some salinity variations. *P. guilfordi*, *P. indica*, and *P. guilfordi* occurred, albeit scantily, in salinities as low as 17.0, 8.5 and 13.0‰ respectively.

4.2.3. Cladocera.

Three species of Cladocera, *Bosmina longirostris*, *Diacyclops thomasi* and *Bosmina longirostris* were recorded from the estuaries of Kerala coast during this study. *B. longirostris* was observed only at Kallai, Neyyare and Veli in small numbers. The distribution of Cladocera in the estuarine systems have been discussed earlier by Madhupratap (1961). This group was usually observed in the monsoon or post-monsoon months. Maximum density recorded was 3420/10m³ in November. In other estuaries also they had a similar distribution and were found during the low salinity period (Table 8), except at Karapuzha where *B. longirostris* was observed in April in low numbers (2/10m³).

Cladocera form a dominant component of the limnetic zooplankton. In the oceans they often bloom into large swarms because of their ability to reproduce parthenogenetically. The three species observed in the estuaries are essentially neretic and are common in the coastal and open waters of Indian Ocean (Dalla Croce and Venugopal, 1972). But they were usually not observed in the estuaries during the high saline premonsoon months. Their presence in the estuaries in very low salinities is therefore surprising. Cladocera were present in the

interior parts of the Cochin backwaters during the low saline monsoon period, albeit their distribution being discontinuous both in space and time (Madhupratap and Naridas, 1975). Similar observations have been made by Hair and Trenter (1971) in the Cochin backwaters and Gowami and Selvakumar (1977) in Mandovi-Suari estuaries of Goa.

Peak populations of cladocerans have been recorded along south west coast of India during Southwest monsoon season (Naridas *et al.*, 1980). Selvakumar (1970) observed cladoceran swarm off Goa in October associated with a diatom peak,

Their abundance in the coastal waters along the west coast is however not restricted to the monsoon period. Devassy *et al.* (1979) have recorded cladoceran swarms off Goa in April. Parvatham *et al.* (1974) observed cladocerans to be abundant along south west coast of India in February-April. A swarm of *Penilia stylirostris* associated with blue green alga *Trichodesmium* and *Pteropod *Gemma* *sigula** has been reported off Cochin in March (Sathivel and Naridas, 1974).

Curiously, the cladoceran population in the inshore waters during this period did not penetrate into the estuary even when the salinity was as high as 3‰.

during peak premonsoon period. Wickstead (1963) had suggested a relation between diatom concentration oxygen level and cladoceran abundance. Association of Cladocera with phytoplankton especially diatoms is fairly well known. Along the west coast of India there is a general outburst of phytoplankton with the out break of monsoon and associated decrease of salinity (Dasin *et al.*, 1972). Competition from other zooplankton organisms which thrive in abundance during the saline period may be restricting the occurrence of cladocerans in the estuaries during this season. But during the monsoon period oxygen in the surface layers increases and there is a decrease in the zooplankton abundance in the estuaries (Maridas *et al.*, 1973; Rao, 1977). This and higher primary production (Dasin, 1970; Devassy and Bhattachari, 1974) in the estuaries during this period may facilitate the viability of cladocerans during low saline period.

4.2.6. *Schizosoma*.

Schizosoma asiatica was observed in very low numbers at Neendakara and Cochin during peak premonsoon month (April) when the salinity was very high. This species is a common neritic form in the south west coast

of India (Jacob George *et al.*, 1978) which prefers salinities higher than 10‰. Their presence in these estuaries may be accidental. Other fresh water crustaceans were also present in the estuaries during monsoon period in small numbers.

4.2.7. Copepoda.

In the marine and estuarine zooplankton copepods almost invariably dominate the counts. These small crustaceans play a vital role in the food chain of the aquatic environment. Mostly herbivores, they form the bridge between the primary and tertiary levels. However, many copepods are omnivorous and some are known to be carnivorous. Copepoda consists of thousands of species, but in estuaries like other fauna only those adapted to this fluctuating environment thrive.

In the present study Copepoda constituted 67.7% of the annual counts. Calanoid copepods constituted the majority of the counts as well as species composition. The herbivores of Cochin sustained a higher standing stock of this group compared to other estuaries.

Peak abundance of Copepoda was in most cases during the premonsoon especially in April. Wide variations existed in their densities during different seasons (Fig. 21). At Cochin maximum abundance was

observed in April ($90,900/10m^3$) and minimum ($30/10m^3$) in August. In other estuaries the number of species and their densities were much less compared to Cochin. Annual averages (Table 5) showed that Kallai and Karapuzha estuaries reached next (maximum density - $21,929/10m^3$ at Kallai in April and $8,485/10m^3$ at Karapuzha in May). Distribution of copepods was also similar in these two estuaries. During the peak of the monsoon period copepods were totally absent at Kallai (July and August) and Karapuzha (August and also in November). At Baypare this group was absent during the low saline months from July to September. The highest density observed here was in November ($10,365/10m^3$). Copepods were generally poor at Mabe estuary also (maximum density - $6,729/10m^3$ in April) and were absent during monsoon period. The distribution of total Copepoda at Neendakara estuary had a different pattern (Fig. 21). During premonsoon period they occurred in low densities. During the monsoon there was a general increase (but lowest was recorded in September) and registered a maximum ($2,342/10m^3$) in December. At Veli higher abundance of copepods was noted during April/May (maximum density - $6,370/10m^3$ in April). They were absent during August and September. Copepods were poorly represented and generally comprised of low saline species at Thottappilly except in April (maximum density in October - $963/10m^3$).

Maximum number of copepod species was observed in Cochin (46) followed by Neendakara (29), Veli (27), Marayunka (23), Kallai (20), Mabe (19), Thottappally (19) and Rappare (17).

Three way analysis of variance (Table 4 B) was performed for the 13 common copepod species to find the significant variations in their abundance over months and areas of collections. *Acartia cartusina* was significantly most abundant species followed by *A. spinicarpa*, *Acartia similis*, *Acartia hirsuta* and *Paracalanus parvicornis*. Minimum abundance was shown by the medium saline species like *Acartia sinuata*, *Acartia hirsuta*, the maritic copepod, *Acartia cartusina* and the low saline estuarine copepod *Acartia granulata*. A true picture of the abundance of the medium and low saline forms do not emerge since the collections were made at the mouth of the estuaries. The seasonal collections made in the middle and upper reaches of the Cochin backwaters show that they dominate these areas for a long period in the annual cycle. However, at one time or another along with the rains and consequent salinity variations all these species shift to the north area of all estuaries giving a fairly good idea of their preferred habitats. The peak saline presentation month April showed maximum

abundance of copepod followed by December and May and minimum was observed in August, September and July. Cochin backwaters again had highest copepod population and Thottappilly ranked lowest.

The distribution of common species of copepods in different estuaries is given in Figs. 23-27. Occurrence of other species and their abundance are listed in Table 6.

Family Acartiidæ had the maximum diversity in these estuaries. Ten species belonging to this family were recorded. Family Pseudodiaptomidæ ranked next by having eight species. Other families represented had lesser number of species and were more common during high salinity regime.

A total of 51 species of Copepoda belonging to 24 genera were observed in the estuaries. Calanoid copepods comprising of 43 species belonging to 13 families constituted the majority. Six species of cyclopoids belonging to 3 genera and 2 genera of harpacticoids represented by a species each constituted the remaining. About 20% of the species occurred sporadically or in small numbers.

Maximum species assemblage was observed in Cochin backwaters. Among Acartiidæ, *Acartia* ~~sp.~~
A. hutchinsii and *A. guineensis* were the most abundant

species during premonsoon and postmonsoon seasons and had more or less similar distribution (Fig. 22 A). They occurred in the middle reaches of the estuary in postmonsoon and early premonsoon period. By April these species penetrated into the interior along with salinity recovery and could be traced upto the head (Fig. 22 B and C).

Acartia pacifica and *A. australis* were also predominantly high saline forms occurring in these seasons. But they were numerically not as abundant at the earlier group. Also, these two species showed a more or less restricted distribution and did not cover beyond the middle reaches. *A. griffithsiana* and *A. pacifica* were recorded in few numbers during the peak salinity period. Their occurrences were restricted to the mouth area. Both are typically marine forms and while *A. griffithsiana* is neritic, *A. pacifica* is an oceanic species.

The distribution of these high saline species of Acartidae in other estuaries is more or less similar as that in Cochin backwaters. *A. griffithsiana*, *A. mini-gutia* and *A. hutchinsoni* were the common species occurring during the high saline months. In general they were numerically less abundant than in Cochin backwaters. At Mandakara *A. griffithsiana* was present throughout the year except in August. Higher numerical abundance

was observed during the postmonsoon than premonsoon in this estuary. *A. guineensis* occurred in comparatively lesser numbers and was absent during early premonsoon and postmonsoon (Fig. 23). *A. hutchinsii* did not show any consistent pattern in its distribution. These three species were abundant at Kallai and Marayathu during the premonsoon and postmonsoon periods (Figs. 24 & 26). Higher abundance of these species was noted only in April at Mals and in November at Neyyare. Their population was poor especially at Mals during other months.

Other high saline species like *Acartia nasutifera* and *A. guineensis* occurred in low numbers in these estuaries in different months. *A. guineensis* occurred only at Mals estuary in November.

Acartia sinuata and *Acartiella hutchinsii* were species that occurred in medium saline conditions and were absent at the mouth of the Cochin backwaters during the premonsoon period. They showed a similar distribution. During postmonsoon and early premonsoon months they were the dominant copepods in the middle reaches where salinity values were roughly between 10 and 10‰. As the premonsoon season progressed and salinity values in this region increased (Fig. 4 B - salinity distribution April) their population maximum shifted to the head region

where the salinity was optimum. By July when salinity was near zero inside the estuary they were completely absent. However, during the monsoon period they were present at the mouth area where stratified waters with medium saline conditions occurred. It may be mentioned ^{that} during postmonsoon and early premonsoon seasons these two species formed the dominant copepods in the interior region of the backwater system.

Acartia ginnana and *Acartia huxleyana* were observed at Marupattu during June and October. Both were more abundant in June (density $1791/10m^3$ and $3581/10m^3$ respectively). At Kallai *A. ginnana* was observed only in November ($19/10m^3$) and *A. huxleyana* in June ($351/10m^3$). They were absent at Sappara and Naha. At Mandakara these two species were encountered only during postmonsoon period. However, the observations at the mouth of these rivers do not necessarily reflect the distribution of the medium and low saline species towards the interiors.

Acartia ginnana is a low saline species and thrived in the estuary during the monsoon period. At Cochin this species thrived throughout the estuary during the monsoon period when the estuary became fresh water dominated (Fig. 22 C). They outnumbered other low saline species of families Diaptomidae (*Diaptomus*

ginseng and *Allodiaptomus nishikii* and *Pseudo-*
diaptomus in this season. However copepod abundance
is far less during low salinity regime compared to
saline period.

A. ginseng occurred at the mouth of Koraput
and Kallai estuaries during the monsoon period. Maximum
density recorded at Koraput was $1754/10m^3$ in October
and $745/10m^3$ in June at Kallai. This species was not
observed at Neendakara, Neyyare and Naha.

Family Pseudodiaptomidae had eight species in the
estuarine waters. *P. parvicornis* and *P. jama* were
the common high saline forms. The two species occurred
in higher abundance at Cochin and showed less tolerance
to lower salinities. *P. parvicornis* occurred in
other estuaries also during the premonsoon and post-
monsoon seasons. At Neendakara estuary they were less
abundant during premonsoon and occurred in higher den-
sities ($391/10m^3$ in December) during postmonsoon. At
Koraput, Kallai and Neyyare also they were observed
during premonsoon and postmonsoon. Other high saline
species of this genus that were found in these waters
were *P. nishikii* and *P. surirella*. *P. nishikii* was
recorded only once from Cochin backwaters. They occurred
more frequently at Neendakara and Koraput while

E. auxillii frequented Kallai and Buppore estuaries (maximum density - $1220/10m^3$ in April at Kallai).

E. annandalei exhibited a wide range of salinity tolerance (0 - 32‰) but preferred stratified waters of the early premonsoon, monsoon and postmonsoon. At Cochin they occurred throughout the year at the mouth region with maximum abundance in July, when the monsoon was at its peak and the surface salinity fell to near zero values. But this species was absent in April when the water column was vertically homogeneous. In spite of the wide range of salinity tolerance exhibited, it did not occur towards the upper reaches where near fresh water conditions prevailed. At Karapuzha and Buppore this species was present only in June. At Kule they were observed in March in high densities ($345/10m^3$) and also in postmonsoon. At Kallai they appeared in March and May and occurred in small numbers at Mandakara during early premonsoon, monsoon and postmonsoon months.

E. hincheyi malayana, *E. pallinosa* and *Ambidiantoma senkung* were the low saline species belonging to family Pseudodiaptomidae. *E. hincheyi malayana* was observed at Cochin and Kallai estuaries during the monsoon period. Maximum density $212/10m^3$ was noted at Kallai in June. *E. pallinosa* and *Ambidiantoma senkung*

were observed during September and October respectively in small numbers. These two species were not encountered in other estuaries except *A. gmelini* which was recorded once from Thottappilly lake.

Four species belonging to Paracalanidae namely *Paracalanus similis*, *A. gibber*, *Paracalanus grandirostris* and *P. pulchellus* were encountered in the estuaries. All but *A. gibber* were observed in high abundance in Cochin backwaters and thrived in the lower and middle reaches during the high saline months.

A. similis was common during high saline months at Neendakara, Kallai, Beypore and Mals estuaries. At Neendakara and Beypore this species occurred more abundantly during the postmonsoon. *P. pulchellus* occurred more frequently at Beypore and Kallai estuaries and sparsely at Neendakara and Mals. Both the species occurred only in December at Korupuzha estuary. *A. gibber* was observed in small numbers at Neendakara, Korupuzha and Mals estuaries in December. Unlike families Acartidae and Pseudodiaptomidae, family Paracalanidae has no low saline species occurring in the estuaries.

The other calanoid copepods which occurred in the estuaries were mostly euryhaline marine forms belonging to various families like Calanidae, Eucalanidae, Centropagidae, Puntellidae, Leucostictidae and Temoridae.

While some of them occurred up to the middle reaches of the Cochin backwaters during high saline period, most occurred only at the mouth areas. Among them *Lebidosoma hastinata* and *Centropomus alcocki* were the species which were common during the premonsoon in all the estuaries. They were less tolerant to lower salinities and hence were not observed beyond the middle reaches of Cochin backwater system. *Lebidosoma hastina* was recorded in small numbers at Cochin and Neyyore estuaries during May and January respectively. Other species belonging to the genus *Centropomus* namely *C. suvattus*, *C. janninensis* and *C. irianensis* were also recorded from Cochin and Neendakara during high saline months. *C. janninensis* occurred in higher density ($562/10m^3$) at the mouth of the Cochin backwaters in March.

Species like *Basilichthys monostus*, *Centropomus nanus*, *Infusilia yiloxia*, *Chromis allicina* which are common in coastal waters were observed at Cochin barmouth during high saline months. *B. monostus* and *B. alternatus* were recorded at Neendakara during April and August in low numbers. *Infusilia flaviventris*, was recorded from Neendakara in August. This is an oceanic bathypelagic species and is probably carried to the coastal waters during the upwelling period. *Thynnus thynnus* and

E. striatella were encountered at Cochin and Mandapam during January and November. *E. striatella* occurred in higher densities ($115/10m^3$) in the Cochin backwaters in January. *E. turbinata* was present at Mandapam also in March. *E. turbinata* is known to form large swarms at times in the coastal waters (Haridas et al., 1960).

The cyclopooid copepods in these waters included five species of the genus *Dithona* namely *D. magna*, *D. lutea*, *D. viridis*, *D. burmannia* and *D. sinuifera* and species of *Sarconema*, *Sassa* and *Saichiria*. Among the genus *Dithona*, *D. magna* and *D. lutea* were more common and exhibited tolerance to lower salinities. *D. sinuifera* and *Sassa* sp. were observed at Mandapam during March. *Sarconema* and *Saichiria* spp. occurred at Cochin in small numbers during the high saline months. *Sarconema* sp. was observed at Mandapam estuary also during pre-monsoon and post-monsoon period.

The harpacticoid copepods namely *Heteromella sinuifera* and *Heteromella sinuifera* were observed at Cochin and Mandapam estuaries. *H. sinuifera* is a coastal species. The other is estuarine and exhibit wide tolerance to salinity variations. *Heteromella sinuifera*, another neritic harpacticoid (Haridas and Rao, 1961) copepod of the coastal waters was once observed at Mandapam estuary during April.

Copepoda of Veli and Thattamally lakes:

Copepoda which constituted 69.2% of the total counts of zooplankton at the Veli lake was the dominant group throughout the year except in July. Twenty seven species belonging to 15 genera were observed, of them, 23 belonged to Calanoida while Cyclopoida and Harpacticoida shared two each.

Maximum density of copepods in the Veli lake was noticed during April ($4370/10m^3$) which formed 81% of the total zooplankton counts. During the monsoon months the number of copepods dwindled to 12.7% (in July) of the total numbers. Copepods were absent in August and September.

During January and February when the surface salinity was around 2‰, and bottom salinity was 18‰, copepod fauna was a mixed assemblage. It comprised mainly of medium saline species like *Acartia salina* and *A. harrisi* and low saline species like *Acartiella maritima*, *Diaptomus minutus*, *Diaptomus minutus*, *Diaptomus minutus* and *Diaptomus minutus*. High saline species like *Acartia salina* and *A. harrisi* also occurred only in small numbers. *A. maritima* showed maximum abundance during this period ($355/10m^3$)

in January). *Pseudodiaptomus grandis* and harpacticoid *Heterosira glacialis* were also observed.

The bottom salinity in this estuary was around 2‰. From March to May (the surface salinity was still low, between 0.7 to 3.2‰) and it fell to 2.5‰ in June. However, during this period high saline species like *Acartia costata*, *A. glacialis*, *A. griffithsae*, *A. pacifica*, *A. hutchinsoni*, *Acartiopsis similis*, *Paracalanus* ~~*E.*~~ *laticornis*, *Centropages alcocki*, *Pseudodiaptomus garrisoni*, *E. jonesi*, *E. gairdneri*, *Emerita stylifera*, *Leptocera hastinata*, *Coronula* sp., *Githona* sp. and *Estheria aestivans* dominated the copepod fauna. Along with these a few low saline forms like *Pseudodiaptomus hutchinsoni* *glacialis* and *E. grandis* were also observed. Of these, *A. costata*, *A. glacialis*, *A. pacifica* and *Acartiopsis similis* were the common species. These species occurred in high densities during April and May. In June and July the salinity of the water column was very low. But still high saline forms like *A. costata*, *C. alcocki*, *E. gairdneri*, *E. garrisoni*, *E. grandis*, *L. hastinata* survived albeit scantily. The low saline species *E. hutchinsoni* *glacialis* was also present in July in small numbers.

In October and November only medium and low saline species like *Acartiella karalensis*, *A. gurnaxi* and *Acartia sinuosa* were present (Fig. 27). *A. karalensis* occurred in high density in October ($734/10m^3$). *Pseudodiaptomus grandis* was also observed in October (salinity of the water column was 0.8‰). In December, the bottom salinity went up to 23.4‰, and the surface values remained at 2.8‰. But the copepod fauna included, in addition to all the high saline species present during premonsoon, some coastal species like, *Acartia pacifica*, *Taraxacum gracilis*, *Acartia sinuosa* in few numbers. *Pseudodiaptomus birchani malayana* was also present and showed the maximum density in this month ($25/10m^3$).

Salinity was very low at Thottappilly lake throughout the year (maximum value observed was only 1.8‰ at surface in June and 7.8‰ at the bottom in January). The copepod fauna was comprised of mainly low saline species belonging to Families Acartiidae, Pseudodiaptomidae and Diaptomidae during most of the months except in April (Table 7). Maximum density of copepods was observed in October ($963/10m^3$). During January to March a mixed assemblage of species like *Acartia sinuosa*, *A. birchani*, *A. sinuosa*, *Acartiella karalensis*, *A. gurnaxi* and *Pseudodiaptomus birchani malayana* was observed.

Low saline species were more abundant among them during this period. *E. hirsutum* maintained showed maximum density ($214/10m^3$) in February. In April higher saline species like *A. mulleri*, *A. hirsutum*, *A. guineensis*, *Amphileptus similis*, *Pseudodiaptomus serricaudatus*, *E. surivilli*, *Centropages alcocki* and *Sithona gona* formed the bulk of the copepod fraction. *A. mulleri* was comparatively more abundant in this month ($34/10m^3$).

During the other months, except in June, July and November when the copepods were absent in this lake, only the low saline species of the family Acartiidae and Pseudodiaptomidae and species like *Halodiaptomus ginsburgi* and *Alodiaptomus mixtilingus* of Diaptomidae and some fresh water *Cyclops* sp. were observed. *H. ginsburgi* and *A. mixtilingus* were the common species having maximum densities in October ($342/10m^3$ and $382/10m^3$ respectively). *Amphidiaptomus senegalensis*, a low saline Pseudodiaptomid was recorded from this lake in December. This species was present in the Cochin backwaters also during October.

The copepod fauna of the eight estuaries studied comprised of 18 families. Of these species belonging to families Acartiidae, Pseudodiaptomidae and Furcaceae were the most common forms and formed the

bulk of the copepod component of the zooplankton. Among these Acartiidae always dominated the counts. In the Cochin backwaters, the three families constituted 95.3% of the total copepods (Acartiidae, 63.3%; Pseudodiaptomidae 12% and Paracalanidae 20%). At Marapuzha Acartiidae formed 80.4% followed by Pseudodiaptomidae (9%) and Paracalanidae (2.3%) together forming 91.7% of the total copepods. At Malai these families contributed to 88.6% of the total copepod counts, Acartiidae being 72.3%, Pseudodiaptomidae 7.3% and Paracalanidae 9.0%. At Neyyare and Mabe Acartiidae constituted only 53.6% and 57.6% respectively. Pseudodiaptomidae showed a higher concentration at Mabe being 25.3% of the total copepods. At Neyyare it formed 9%. Family Paracalanidae constituted 20.3% and 8.9% in these two estuaries respectively. 89.3% of the total copepods was constituted by these three families at Neendakara, Acartiidae contributing 67.6%, Pseudodiaptomidae 14.3% and Paracalanidae 7.7%. At Veli they formed 79.3%, 6.7% and 9.7% respectively, together constituting 95.7% of the total. At Thottappilly they constituted only 51% (Acartiidae 33%, Pseudodiaptomidae 15.3% and Paracalanidae 2.7%). The rest 49% was formed by species of the family Diaptomidae and some cyclopoidea.

4.2.3. Amphipoda.

This group included mostly species belonging to Family Gammaridae while a few hyperid amphipods occurred in small numbers in some months. Being mostly benthic, only part of the population which periodically migrate to the water column is normally represented in the plankton samples. They contribute significantly to the benthos of the estuary and have been found to occur in large numbers in muddy areas.

Three species, *Gammarus trisacculus*, *Malina varians* and *Exochus digitata* were commonly found in these estuaries. *G. trisacculus* was the most abundant species and the other two species were met with only occasionally. In the Cochin backwaters, this group was present throughout the year and maximum density was recorded in July ($960/10m^3$). Of the eleven species in this backwater system, occurring *G. trisacculus* is the commonest, exhibiting a wide range of salinity tolerance (from 0.1 to 27.7‰) with peak abundance attained in medium salinities (Nair, personal communication).

At Karapuzha, Maho and Kaypara they were found in all seasons. Highest density was observed at Maho ($1000/10m^3$) in November. They occurred during pre-monsoon and monsoon periods only at Neendakara and at

Nauplii during premonsoon and postmonsoon. They were observed only in small numbers at Thottappilly and Veli lakes. Murugesan *et al.* (1980) have recorded 6 species from Veli where *Exochia digitata* was the abundant species.

4.2.9. Sargastidae.

Leptocryptus bengali was the common sargastid encountered in the estuaries. Higher abundance was always noted during the high saline months. At Cochin and Neendakara it occurred throughout the year except in one or two months during monsoon and postmonsoon period. Maximum density of this species at Cochin was $260/10m^3$ in May and $777/10m^3$ at Neendakara. Another species *L. kuma* was also present in the Cochin backwaters. It was present only from May to June and maximum density was only $80/10m^3$ in April. This species was not recorded from other estuaries. During peak premonsoon period (April) *L. bengali* was present upto the head of the Cochin backwaters. Another sargastid, *Leptocryptus* sp. was also present during the premonsoon and monsoon months in small numbers. *L. bengali* had a more or less similar distribution in the other estuaries with higher abundance during the high saline period. It was not

observed in these waters during the peak monsoon period. Maximum abundance was observed at Bayyare (2000/10m³ in May) when they constituted 39.2% of the total zooplankton. At Kallai they occurred in higher abundance (1230/10m³) in April. *Amphioxus* sp. was observed at Veli in small numbers in September. *L. hassani* was observed at Thottappally in various months.

4.2.10. *Nysidacea*.

Rhynchostylax indicus was present in very small numbers during the high saline months in the Cochin backwaters and once at Veli lake. They were not observed in the other estuaries. Like amphipods, the distribution pattern and abundance of *Nysidacea* cannot be gauged from their numbers in the present collections since they are actively migrating forms and usually come to the surface only during night.

4.2.11. *Gammarus*.

Another benthic group, occasionally found in plankton collections, gammarans were observed in all the estuaries in small numbers during different seasons. At Kallai, Njrapuzha and Kala they were observed once

during the postmonsoon period. At Beypore they occurred during April, May and June with maximum density in May ($144/10m^3$). They were present at Velli and Cochin estuaries only during the monsoon season. Maximum density noticed at Cochin was $157/10m^3$ in July. They occurred during postmonsoon and monsoon periods at Neendakara - maximum density observed was $226/10m^3$ in August.

4.2.12. *Zanclus*.

These organisms occurred in very small numbers in the plankton samples collected from Cochin and Marupuzha estuaries. While they were observed more frequently at Cochin backwaters during monsoon and postmonsoon period (maximum density $12/10m^3$ in May) they were present only in March at Marupuzha ($9/10m^3$).

4.2.13. *Branchiostoma laevis*.

Escaped larvae constituted the majority of invertebrate larvae in these estuarine waters. They ranked next to copepods in overall abundance of zooplankton in the estuaries (9.6%). They formed 17.8% of the total annual counts at Mabe estuary followed by Velli (15.6%), Beypore (14.7%), Neendakara (11.6%),

Kallai (9.3%), Thottappilly (8.6%), Cochin (7.8%) and Marupuzha (7.3%).

Zoea larvae were very common and occurred in all the seasons with peaks usually during the pre-monsoon months, except at Beypore and Mandabara estuary where maximum density was observed in December ($3214/10m^3$ and $269/10m^3$ respectively). At Kallai and Velli maximum density was in May ($1471/10m^3$ and $850/10m^3$ respectively) while it was in April at Marupuzha and Naha. At Cochin they were observed throughout the year except in May and July with maximum density in February ($720/10m^3$). Zoea larvae were quite abundant in the middle and upper reaches of the Cochin backwaters in pre and post monsoon seasons. At Thottappilly zoea occurred in small numbers during the pre-monsoon months and also in August and December.

Larvae of penaeid and caridean decapods at various stages of development were present in these waters almost throughout the year. These included the larvae of the commercially important species like *Penaeus indicus*, *Metapenaeus schomburgkii*, *P. monodon*, *P. affinis*, *Macrobrachium manoharanii* and *M. idella*. Higher abundance of these were noted during the pre-monsoon and post-monsoon periods. They were more

abundant in the Cochin backwaters, and maximum density was recorded in December ($4450/10m^3$). At Baysore also highest density was in this month ($1004/10m^3$) (Figs. 13 & 15). Higher densities were observed during the premonsoon period at Mandakara, Korapuzha and Mabe. These larvae were observed in small numbers at Thottappilly also throughout the year except November).

Alima larvae of squilla were present in small numbers at Cochin, Mandakara and Mabe estuaries, during the premonsoon period. Megalopa larvae were observed in these waters during this period and also in slightly higher numbers in August at Mandakara ($41/10m^3$). Nylisema larvae occurred at Mandakara estuary in April ($32/10m^3$ - Table 7).

Cirripeds larvae were present in these estuaries except at Thottappilly. They were present throughout the year in Cochin backwaters except in July (maximum density - $260/10m^3$ in February). In the other estuaries these were observed only during the premonsoon months and June. Highest density was in Kallai estuary in April ($1404/10m^3$). At Baysore, Mandakara and Velli they were poorly represented.

Polychaeta larvae occurred in all the estuaries in various periods, in small numbers.

Cyprinae larvae of *Mytilus* and *Actinostrea* larvae of *Strenidae* occurred in low numbers at Cochin backwaters during April. Pluteus larvae of *Setina-dornata* were observed at Kallai and Mabe estuaries during March. Lingula larvae of *Brachyopoda* were present at Mandabara and Mabe in May.

4.2.14. Fish eggs and larvae.

Fish eggs commonly occurred in the estuaries except at Thottappilly. They were usually sparse at the peak of the monsoon. In the Cochin backwaters fish eggs were observed throughout the year except in July and August. Maximum density was $420/10m^3$ in November. At Kallai, Bypass and Mabe they were present in all seasons. Maximum density was observed at Kallai ($300/10m^3$) in March.

Larvae of fishes mainly belonging to the families Amblycidae, Mugilidae and Cobitidae were common. They were observed throughout the year in the Cochin backwaters (maximum density $200/10m^3$ in July). They were present at the middle reaches by November and at the head during the peak monsoon months. At Koraganha estuary also they occurred round the year with maximum

density ($207/10m^3$) in December. The larvae were present during all seasons at Mandakara, Kallai, Buppura, Maba, and Veli estuaries although period of peak density varied. They occurred in small numbers at Thottappilly lake.

4.2.15. Appendicularia.

Appendicularia showed higher abundance in the premonsoon and postmonsoon seasons. This group was absent at Veli and Thottappilly lake. At Cochin, their highest abundance was noticed in February ($2610/10m^3$). They were observed mostly during the premonsoon period at Kallai, Buppura, Maba and Maruputha. While they were more common in the Kallai estuary (maximum density $702/10m^3$ in April), they occurred only in small numbers in others. They were also observed in low numbers at Mandakara during early premonsoon (January), monsoon and postmonsoon period (November-December).

4.2.16. Thaliacea.

Thaliacea which is common in marine ecosystems are usually sparse in the estuaries. The salp Thalia gambusia was noticed at the mouth of the Cochin backwaters in April. Thaliacea did not occur in other estuaries.

4.3. General Discussion.

The zooplankton of the estuaries comprises of truly estuarine species, euryhaline marine forms and a few stenohaline marine and freshwater species. The latter two are only stragglers into the estuaries, carried by waves or currents, and never occur in large numbers. On the other hand many of the euryhaline marine species are able to thrive in appreciably large numbers near the lower reaches of the estuaries during the saline period. Nevertheless, there is no such evidence to suggest that they could breed in this area since their juvenile population is very low. The entire recruitment of this class is probably from the adjoining neritic waters through tidal currents.

Among the species which could be classified as true estuarine forms, three classes - high saline, medium saline and low saline can be recognized. This classification, especially regarding the high saline forms is somewhat arbitrary since most of them exhibit a wide range of salinity tolerance. However, this grouping is based on their observed abundance, and these species occur in large numbers in higher salinities.

Some of these high saline forms which are classified as truly estuarine occur in the inshore waters, but in low numbers. The probable reason is competition. Grindley and Woodbridge (1974) found the salinity tolerance of Pseudoisotomid species of Richards Bay, South Africa, ranged from near freshwater to 6‰, but peak survival was at around 3‰ salinity. They contend that it is not salinity but competition from marine organisms that prevent them from surviving in the sea. The adaptation of the estuarine fauna to tolerate extreme fluctuations in environment allow them to flourish in the estuary. Similarly, the euryhaline marine organisms which frequent these waters must be facing competition from the estuarine organisms restricting their abundance during the saline period.

One striking feature in the zooplankton abundance of the estuaries is its high standing stock, counts and thus the high turn over at secondary level in general, compared to the adjoining sea. The peak zooplankton standing crop and counts observed at Cochin backwaters in this study was 9.8 mg/m^3 and $10110/\text{m}^3$ respectively. In other estuaries also zooplankton standing stock and counts were high during the saline period. Subbaraju and Krishnamurthy (1972) observed an average standing

stock of 2 mg/m^3 (maximum 4 mg/m^3) and counts of $92,000/\text{m}^3$ (maximum $204,000/\text{m}^3$) from Vellar estuary during summer months. Grindley and Wooldridge (1974) recorded the density of a single estuarine copepod *Paracalanus parvus* as high as $42,700/\text{m}^3$. In contrast, the coastal waters, let alone the open ocean, are far less productive. The annual range of zooplankton biomass was from 0.07 to 0.3 mg/m^3 and counts were between 90 and $1001/\text{m}^3$ in a nearshore environment in the southwest coast of India (Varidas *et al.*, 1980). Representative figures show that the average biomass ranged between 0.07 and 0.08 mg/m^3 (Wair *et al.*, 1977) and 0.09 and 0.3 mg/m^3 (Wair *et al.*, 1981) in the Bay of Bengal and 0.09 and 0.1 mg/m^3 (Wair *et al.*, 1978) in the Arabian Sea. In the Andaman Sea the range was from 0.02 to 0.1 mg/m^3 and with counts of 7 to $14/\text{m}^3$ (Madhupratap *et al.*, 1981).

The real reason for this enormous differences is not probably due to large scale variations in primary production between the two environments (the differences in primary productivity of coastal and estuarine waters is only marginal, and not consistent - see, Qasim *et al.*, 1969; Radhakrishna *et al.*, 1970 a,b; Shettethiri *et al.*, 1980), but because many of the estuarine organisms are

omnivores and feed on detritus and bacteria as well (see Chapter 3). Large quantities of detritus and associated bacterial flora are carried/produced into the estuaries and food is probably never a limiting factor for the estuarine zooplankton.

Clinal changes associated with seasons are much more apparent in the estuaries compared to neritic or oceanic environments. Zooplankton standing crop is high and high saline species are diverse during the saline period. In copepods, about 10 true estuarine species belonging to families Acartiidae and Pseudodiaptomidae abound in the estuaries during this season. As the salinity increases from mouth to head of the estuaries, these species are able to invade and propagate throughout the estuaries. A few species of the family Paracalanidae are also able to successfully compete with these species but are essentially neritic forms. The other species which occur during this period are euryhaline marine forms with limited distribution as mentioned earlier.

On the otherhand, species which preferred medium saline conditions were represented by only two form viz. *Acartia hirsuta* and *Acartia hirsuta*. Nevertheless, they occurred in large numbers during the saline period, but towards the middle and upper reaches, where the salinity was optimum.

When the monsoons reduce the salinity to near freshwater conditions, zooplankton standing crop and counts are very poor inside the estuaries. All the high saline and medium saline species are totally flushed out of them. About 6 low saline copepods represented by the families Diaptomidae, Pseudodiaptomidae and Acartiidae occur in the estuaries during the monsoon season. However, *Acartiella granulata* is the only species which show some abundance in this period. But its peak densities are nowhere near the abundance showed by the high or medium saline species. Other low saline species such as *Halodiaptomus gibbus*, *Allochydona kirchilina*, *Pseudodiaptomus kirchilina*, *P. hallanensis* and *Ambidaptomus senegalensis* occur in very low numbers.

Thus monsoonal inundation, tidal inversion and associated changes in salinity are the main factors controlling the zooplankton of the estuaries. Other factors such as temperature, oxygen content and availability of food are apparently of secondary importance as far as tropical estuaries are concerned. The inability of the low saline zooplankton fauna to thrive in larger numbers during the low saline period is surprising. It is probably because they are not able to withstand the strong currents and these estuaries are left virtually unexploited at secondary level during this period.

Variations of wide nature, both seasonal and spatial, in zooplankton counts have been reported from other estuaries also. Secondary production is low in the low saline season compared to the pronounced period in the Mandovi-Suari estuarine system of Goa (Salvanesar *et al.*, 1980). Peak population occur in summer months in Vellar estuary of Forto Novo (Subbaraja and Krishnamurthy, 1972) and they conclude that salinity and rainfall control the zooplankton abundance of these waters. Similar observations supporting lesser zooplankton abundance during low salinity have been made from mangrove environments of Forto Novo (Subbaraja and Krishnamurthy, 1981), Hooghly estuary of west Bengal (Sarkar and Choudhury, 1981) and Kall estuary of Kerala (Kumar *et al.*, 1981).

The zooplankton counts of the Kall estuary were reported to vary from $1700/m^3$ in July to $21,000/m^3$ in October. Zooplankton counts ranged from $100/m^3$ to $300,000/m^3$ in the estuaries in Victoria, Australia (Moale and Dayly, 1974). In South Africa, Pynna estuary, ^{where} rainfall is more or less evenly distributed throughout the year, has an abundant fauna compared to St. Louis estuary where rains flood the system during part of the year (Day, 1967).

Copepoda dominated the average zooplankton counts in all the estuaries presently studied. While this is so from data available from other estuaries of India, the dominant component has been reported to vary in different waters. *Cirrropoda nauplii* dominate the Southampton water (Raymont and Currie, 1938) and York river, U.S.A. (Jeffries, 1964). Polychaete larvae form the major component in Raritan Bay and both polychaete larvae and lamellibranch larvae dominate in Narragansett Bay. (The absence of veliger larvae in the present collections is probably due to the larger mesh size used). Egg larvae of *Brachura* have been reported to dominate the zooplankton of Cochin backwaters (Madhupratap, 1978) in certain months. In the present collections copepod larvae dominated the counts in a few months at Mandakara and Bhyppore estuaries.

While the distribution of the zooplankton species in the estuaries with a perennial connection to the sea follows a confirmed pattern, their distribution in the two lakes studied (Veli and Thottappilly) is perhaps a little tangential to the usual assumptions. There is no handy alibi to the occurrence of marine forms like *Thanaos strifera*, *Leptocera gracilis*, *Acartia pacifica* and several others at Veli in some months (March, April, December) when the lake has no free connection to the sea.

It has to be assumed that they might have come through the spill over of wave action and were able to survive although in low densities because of the higher saline bottom water.

A mixed assemblage of estuarine copepod species (consisting of low, medium and high saline forms) was often encountered in these estuaries. At Valli during January-February months when salinity was between 2‰ (surface) and 12‰ (bottom) medium and low saline species dominated with high saline forms occurring in small numbers. But from March to May high saline species were dominant forms (salinity 2 to 23‰). But in June when salinity of water column was as low as 2‰, a few high saline forms were observed. Again in July when near fresh water conditions existed low saline species were found along with a few high saline forms (*Paracalanus parvus* and *P. parvus* dominated, ^{the latter} *P. parvus* dominated copepods at Kallai in April when the salinity of the water column was around 3‰). In October when salinity was 0.6‰, medium saline species like *Paracalanus parvus* and *Paracalanus parvus* occurred although one would have expected low saline species like *P. parvus* to dominate. At Thottappilly lake, low saline forms were more abundant in most months as could be expected but a

5. COMMUNITY STRUCTURE.

Estuarine plankton, as mentioned earlier, form a class by themselves chiefly because they are adapted to the vagaries of this environment. The zooplankton element consists of both holoplankton and meroplankton. It is evident from the distribution and abundance (Chapter 4) that the holoplankton is dominated by a single group - Copepoda. The meroplankton may consist of larvae of many benthic invertebrates and fish. In the estuaries investigated in the present study, naupliar larvae of *Brachyura* was the dominant form. Other larvae of polychaetes, cirripedes, ceridarians and other decapods were also frequent.

The stenohaline marine forms and some fresh water organisms which enter accidentally, do not have any role in the ecosystem of the estuaries. Records of these species from estuarine mouths or heads are of academic interest only. It could be seen that many groups of zooplankton like euphausiids, ostracods, appendicularians, salps and doliolids which play an important role in the oceanic ecosystem are usually excluded in the

estuarine plankton. Curiously, almost all of them are filter feeders and the absence of these groups in the estuarine habitat is interesting. Cladocera, another filter feeding group, also had a peculiar distribution in the estuaries (Chapter 4.2.5). Many of the groups which are diverse in the oceanic environment like hydromedusae, ctenophora and chaetognaths (all carnivores) have only a few representatives (mostly allochthonous forms entering the estuaries from the adjoining neritic waters during the saline period) in the estuary.

The zooplankton is thus dominated by true estuarine forms which have evolved adaptations to the fluctuations mainly salinity. The salinity ranges for the common species that occur in the estuary (Fig. 25 A & B) are, however, not their salinity tolerances, but only the recorded ranges in the present observations. Experimental studies have shown that many low saline species can be acclimatized slowly to readapt to sea water indicating that their low salinity adaptation was physiological than a fixed genetic change (Grindley, 1960).

As discussed earlier (4.3), estuaries sustain enormous standing stock of zooplankton compared to the sea. But the role of the zooplankton in the food chain

is difficult to be pinpointed in the estuarine ecosystem as there is no shoaling fishery in the shelf waters. Even larger estuarine fishes like Mugil and Chanos are mainly herbivorous or detritivorous (Hiett, 1944). The coefficient of energy transfer from primary to secondary level was only 7.6% for Cochin backwaters and 6.6% for Mandovi-Zuari estuaries of Goa (Salvalkar et al., 1980) indicating excess phytoplankton production is also available for alternate pathways. Perhaps a large portion of phytoplankton and zooplankton production in estuaries contribute to the productivity of the coastal waters or forms a major source to the organic matter in bottom deposits. This could help the sustenance of a rich benthic life and in fact, high benthic biomass has been reported from several estuaries (Kurian, 1972; Ansari, 1974; Parulkar et al., 1980). The population of penaeid prawns which abound in the estuaries during the saline period may also be a major direct or indirect consumer.

Calanoid copepods play a pivotal role in the ecosystem of any aquatic environment. In the estuaries all over the world they are dominated by only a few genera or families. In the estuaries of India species of the families Acartiidae and Pseudodiaptomidae are dominant. A few species of the family Paracalanidae also occur in appreciable numbers. Species of the genera Eurytemora,

Acartia, Pseudodiaptomus and Tropidocyclops characterise the estuaries of South Africa (Grindley, 1980). Australian estuaries include a few other genera also like Boeckella, Glabidocyclops and Salinacyclops (Tew and Ritz, 1978).

Heinrich (1962) recognises three 'types' of life cycles in the zooplankton in relation to feeding. Type 1 depends on the availability of food and breed only when food is plenty. Breeding in type 2 is independent of food supply and they usually store fat. These two types mainly occur in the higher latitudes and bathypelagic systems. Type 3 occurs mostly in the tropics where the species are more or less continuous feeders and breeders. The estuarine copepods probably fall under this category since the copepodites of true estuarine forms are present in the collection although the saline premonsoon season. Biochemical studies on zooplankton from Cochin backwaters (Madhupratap *et al.*, 1979) indicate that most species have a very low lipid storage indicating that they feed continuously. In such cases protein may function as an important food reserve (Conover and Corner, 1968; Raymond *et al.*, 1969) which may be mobilised to meet the metabolic requirements.

The common carnivorous copepods of the pelagic realm belonging to the families Euchaetidae, Pentalidae and Candacidae are not represented in these estuaries (but for the genus Lehidocopa). The few species of the family

Paracalanidae are probably herbivores whereas most of the species belonging to families Acartiidae and Pseudodiaptomidae are omnivores or detritivores. Only a few species like *Acartiella skanavi* and *A. bengalensis* appear to be carnivores (Tranter and Abraham, 1971).

The importance of detritus and associated bacterial load as a direct nutritional source to zooplankton especially copepods is now widely recognised (Heinle and Flower, 1975; Heinle *et al.*, 1977; Conover, 1979). Obviously there is no dearth of detrital material in the estuaries. Studies from Cochin backwaters reveal that phytoplankton production is in excess when compared to zooplankton grazing pressure (Qasim, 1970; Madhupratap *et al.*, 1977). Hence availability of food is not the limiting factor for zooplankton survival in these estuaries.

In the higher latitudes there is usually a pulse of phytoplankton production in the spring. This is followed, after a lag, by an increase in zooplankton standing stock. In the tropics however, there is usually, no such sudden pulse and phyto- zoo- plankton production rates show a more or less an even curve. In contrast, in the coastal regions upwelling may cause sudden blooms of phytoplankton associated with large swarms of filter feeders like tunicates (Madhupratap *et al.*, 1980).

The production trends of phytoplankton and zooplankton in the Cochin backwaters are not much varied, but the peak abundance attained by zooplankton in mid-summer (Fig. 29 A) is probably due to a relatively more stable environment (see Chapter 7) than the availability of food supply. Similarly salinity and currents must be affecting their survival in the monsoon period since phytoplankton production was fairly high during this period.

The carnivores of the zooplankton component (hydro-medusae, ctenophore and chaetognaths) also show a close relation to increase in abundance of other zooplankton (Fig. 29 B). Naturally, their numbers are much less compared to secondary producers as in any ecosystem (say, terrestrial). All of them are high saline forms and apart from a low availability of food, salinity must be a major factor causing the decline of their population during the monsoon.

These estuaries are thus left unexploited at secondary level during the monsoons. The pathways of energy (derived from primary production and detritus etc.) transfer during this period is yet to be worked out.

The food chain in the estuaries are apparently simple compared to the more complex oceanic environment. The shallowness restricts vertical compartmentalization, unlike the sea where vertical migration and depth range of zooplankton species are critically associated with distribution of food. The excess phytoplankton production and bacteria along with zooplankton and their fecal pellets contribute to the richness of the bottom deposits. The benthic community flourish and many of these estuaries are rich in clam beds, polychaete and amphipod communities. The benthic community is in turn exploited by an abundant population of prawns and other predators which feed on them.

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6. SPECIES DOMINANCE AND SUCCESSION.

In tropical estuaries, salinity is the key factor that controls the distribution and abundance of various zooplankton species. During the monsoon, because of heavy discharge of fresh water, the salinity is reduced to near fresh water conditions in the estuarine systems. Almost all the zooplankton organisms are wiped out with the exception of a few low saline species. Repopulation of these waters start during the postmonsoon period. Their intrusion and propagation towards the interiors of the estuaries depend largely on the salinity incursion. The successional pattern of various species could be deduced from the numerical abundance.

Compared to the low saline season the zooplankton numbers increase many fold during the favourable saline period. The estuarine species are r selected (Highly unpredictable or seasonal environments favour opportunistic species with high rate of increase - r selected - while the more constant environments do not - K selected, MacArthur, 1972). The

succession shows that although many species appear in the sequence a few species among them tend to dominate numerically. The percentage of dominance may vary, but often a few species together constitute the major portion of the population.

Comparisons of species rich communities (like the open ocean) to species poor communities (like estuaries) have led to the generalisation that there are fewer numerically dominant species in the former (Mac Arthur, 1969). While there is an increasing gradient in species diversity from estuaries to open ocean, studies reveal that more often a few species tend to occur in greater abundance in the stable environments also (Haridas *et al.*, 1960; Madhupratap *et al.*, 1961; Nair *et al.*, 1961). Birch (1961) analysing the marine benthic communities also came to the conclusion that Mac Arthur's theory need not always hold true.

In the estuaries Copepoda almost always showed the highest numerical dominance. Although 51 species belonging to this group were recorded only a few generally dominated the assemblage. Other groups/species predominated only rarely.

In the Cochin backwaters, Paracalanus crassirostris was the dominant species at the mouth area during the early postmonsoon period (November). This species was replaced by Paracalanus aculeatus during the late postmonsoon and early premonsoon months. Acartia gauthieri dominated all other species, all through the rest of the premonsoon period (Fig. 30 A). Other high saline species which are common in these waters like Acartia sinilis, Acartia hutchinsoni, A. sinuata, A. pacifica, Pseudodiaptomus sarasinoides, P. jayakari and species of the family Centropagidae though occurred in considerable numbers are dwarfed by A. gauthieri. During the peak monsoon month (July) Pseudodiaptomus snyderi which preferred stratified waters is the dominant species. Acartia plumosa, a medium saline species was common during late monsoon. This species with Acartiella karlingi predominated in the middle reaches during the peak salinity regime. Though other high saline species of Acartiidae penetrated into these areas during this period they did not dominate. During the monsoon period A. sinuata is the dominant species in the interior of the estuary.

Acartia gauthieri showed absolute dominance during most of the months of the year at Neendakara estuary. During the peak premonsoon period (April-May) Lucicutia

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hanseni and Paratya haidari predominated (Fig. 30 A). At Kallai also A. sentryi was the dominant species during the pre- and post- monsoon. But during low saline period, naup larvae, Acartiella keralensis and A. grimaldi replaced the high saline species. Almost similar situation existed at Kappanada estuary also (Fig. 30 B).

Similar to Cochin backwaters, various high saline species occurred in higher abundance during the pre-monsoon and postmonsoon months in these estuaries also. But Acartia sentryi almost always outnumbered these species. The low saline species, Acartiella grimaldi was absent in Mandakara estuary, and species like A. keralensis and Acartia plumosa occurred in low numbers. This was because, very low saline condition did not exist at the mouth area of this estuary. Even at the peak of the monsoon salinity was 6.7% and 14.6% at the surface and bottom respectively.

In the estuaries of Mahe and Beypore dominance of copepods was not as high as in other estuaries (Fig. 30 B). Acartia sentryi dominated only during peak premonsoon at Mahe while at Beypore it dominated during early premonsoon and postmonsoon. Paradiaptomus monandala was the dominant species



Mahe and Oithona nana at Beypore. Brachiurus zoea and caridean larvae were dominant during early pre-monsoon and monsoon season and the amphipod Coryphina triangula was abundant in postmonsoon at Mahe. Eutima mammalis, a hydromedusa and the sargostid Lucifer hassani were the dominant species during the peak saline months at Beypore. Zoea, larvae of carideans and the cladoceran Bosmina longirostris stepped into dominance in some months during monsoon. The ctenophore Fluxibrachia globosa dominated in late postmonsoon.

Acartia australis was the dominant species during the early premonsoon at the Thottappilly and Veli lakes (Fig. 30 C). At Thottappilly it was replaced by medium saline species like Acartia plumosa and Acartia borealensis during late premonsoon. Only some caridean larvae were present during the early monsoon period. Very low saline diaptomids like Halodiaptomus cinctus and Allediaptomus mirabilis were dominating copepods during late monsoon and postmonsoon months. But in the Veli lake high saline species like Acartia sentena, A. spinicarpa and Pseudodiaptomus aurivilli dominated during the premonsoon months. Though low saline copepods were present

During the postmonsoon months, caridean larvae dominated.

Zooplankton populations in the estuaries are rich during the saline period. Most of the common species that occurred during this period were able to tolerate a considerable range of salinity variations and dominance is probably achieved at optimum salinity and when other environmental factors are also conducive.

Salinity recovery is faster at the mouth area of the estuaries during postmonsoon season and is slow towards the middle and upper reaches. Broadly, the successional pattern showed three series, the low saline forms dominated the entire estuary (but in low numbers) during monsoon and the head region during postmonsoon. The high saline forms dominated the mouth area during postmonsoon and the middle reaches during premonsoon. The medium saline species were abundant in middle reaches during early premonsoon and at the upper reaches during later premonsoon. The medium saline species replaces the low saline forms towards the head as salinity recovers and later high saline species also invade this area in late premonsoon when salinity increases further.

In general *Acartia sentrum* was the dominant species and this along with a few other species like *Acartia similis*, *Paracartia grossirostris*, *Acartia hirsuta*, *A. spinicauda* and *Paradiploantenna garrismantui* formed bulk of the population during saline period. Although the carnivorous groups like hydromedusae and ctenophora were not numerically dominant, the ecological dominance of these groups cannot be overlooked.

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7. SPECIES DIVERSITY.

The data most basic to the understanding of community structure are the number of species present and their relative abundance. In nature, we find that some communities are rich in the number of species (like perrenial forests of tropics, oceanic environment etc) while certain others (like estuaries) are relatively poor in this respect. Ecologists agree that there is a general increase in the number of species from higher latitudes towards the tropics. However, the reasons for variations are far from clear. Several theories with respect to this have been suggested, but no single explanation is sufficiently cogent to pinpoint the variations.

Within the tropical aquatic environment itself we find that there is a decreasing gradient in diversity from oceanic to neritic and to estuarine habitats. Mac Arthur (1965) stated that "the number of species within a habitat can be expected to increase with productivity (sometimes), with structural complexity of the habitat, lack of seasonality in resources and

the degree of specialization". Various theories involving time (older communities have more numbers of species), spatial heterogeneity (more heterogeneous physical environment supports more species), competition (leads to narrow niches), predation (reduces competition and allows more prey species), environmental stability (more species in stabler environments) and productivity (greater production results in greater diversity) have been put forward. However these theories, apart from lack of general stability of the environment do not explain the lower diversity in the estuaries.

Ecologists argue that negative feed back in ecosystems damps perturbations and the stability or the resilience of the environment contributes to higher diversity. The idea that ecosystems possess such stability was introduced by Mac Arthur (1955). Ecologists' subsequent efforts to verify this hypothesis became complicated by attempts to define stability and diversity in operational ways (Patten and Odum, 1981). "Increased stability with increased species is difficult to demonstrate.... Models reduce the intricate beauty and awesome complexity of a piece of living nature to what is by comparison

a flat pallid image of reality. An ecosystem model, no matter how sophisticated or difficult to produce is but a shadow of its prototype" (Patten *et al.*, 1975).

Environmental conditions in tropical estuaries are highly fluctuating unlike marine environment. A more or less stable condition in estuaries is attained only during the saline pronounced months and zooplankton species are more diverse during this period compared to other seasons.

Two indices, 'D' (Margalef, 1969) and 'E' (Heip, 1974) were used to evaluate the diversity of zooplankton in the estuaries. The formula used for the index 'D' was,

$$D = \frac{S-1}{\log_e N}$$
, where S is the number of species and N is the number of individuals of all species and for E was

$$E = \frac{e^H - 1}{S-1}$$
, where S is the number of species.

The index 'D' is relatively stable compared to α (Fisher *et al.*, 1943) but this index also possess the drawback that it is affected by the sample size. The evenness index is the ratio between

the actual diversity and the maximum possible diversity which occurs when all species are equally abundant and ranges from 0 to 1. In short it gives the evenness in distribution of species within the sample. Evenness is the inverse of dominance.

The number of species and the indices 'E' and 'D' for the eight estuaries are given in Table 9. Diversity value, 'D', was higher during the salinity regime and lowest during the peak monsoon months. At Neendakara the values remained high throughout the year. It ranged between 3.2 and 7.0. Salinity was also high at the mouth of this estuary in most months. In the Cochin backwaters these values ranged between 2.2 and 5.3 comparatively higher values being observed during the time when salinity recovery started. Both marine and true estuarine forms occur at the mouth during this period. The values were comparatively lower in the Karapuzha estuary.

The index 'E' was highest at Thottappilly lake and to some extent at Vali also. In the other estuaries higher values were observed in certain months especially when the species number and zooplankton population were low. Thus during this period although the number of species was less, their distribution within the total numbers was more or less even.

The number of species which occurred showed more or less a direct correlation with salinity when the distribution in the upstream areas of Cochin backwaters was analyzed (Fig. 31). There was a progressive decrease in the number of species along the salinity gradient towards the upper reaches. During July when the system was practically fresh water, only very few organisms tolerating low saline conditions thrived in the estuaries.

In almost all the estuaries, the greatest diversity occurs near the mouth where a wide range of neritic species also appear. Some of the higher values (D values) observed in some estuaries especially at Neendakara (7.0 in April, 6.1 in September) are because of stray occurrences of some of the coastal and neritic species. Thus if the estuarine and euryhaline species which are common in the estuaries only are taken into account and the less tolerant neritic species omitted, the indices would be still low.

Species diversity in estuarine zooplankton was low compared to coastal or oceanic waters. The average diversity index ' D ' ranged between 1.5 and 3.5 in these estuaries. It ranged from 3.1 to 7.7 in a nearshore environment (Haridas *et al.*, 1960) from 8.2 to 12.9 in

the coastal and oceanic waters of the Bay of Bengal (Nair *et al.*, 1981) and from 3.2 to 8.1 in the Andaman Sea (Madhupratap *et al.*, 1981). Thus there is a progressive gradient in the zooplankton diversity from estuarine environment to open ocean. A similar trend was observed for the tropical benthic communities by Sanders (1969). He maintains that estuaries are in principle physically controlled environment, unlike the open ocean which is more stable and develop biologically accommodated communities. The number of species present diminish continuously along the stress gradient from a stable environment to habitats where conditions are fluctuating and finally when the stress conditions become greater than the adaptive abilities of the organisms, an abiotic condition is reached.

Diversification has several important implications for the community. Ricklefs (1973) says that many species can exploit different kinds of resources more efficiently because the evolutionary independence of reproductively isolated populations allows specialisation. Further, diversity creates heterogeneity in the environment which provides the basis for increased diversification of life forms. Whether species exploit more ecological roles in areas of high diversity in view of

the greater variety of ecological opportunities or because species diversity ^{is} to avoid competition, or both, is still open to question.

Productivity or spatial heterogeneity are certainly not the factors affecting the species diversity in the estuaries. Analogous situations where diversity tend to decrease with higher standing population have been reported (Deevey, 1971; Hair et al., 1961). Phytoplankton diversity has been found to be high in oligotrophic areas compared to eutrophic areas (Peterson, 1975). It can only be speculated that lack of stability or time to diversify (only the intermonsoon period allows some diversification) and the constant physical changes of the environment lead to a lesser diversity in the estuaries.

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8. ZOOPLANKTON ASSEMBLAGES.

A variety of zooplankton organisms inhabit the estuarine environment competing or accommodating each other. The degree of coexistence and competition depends on the requirement and availability of food, space etc. in the habitat. Earlier informations on the coexistence of *Acartia* spp. and other copepods in the Cochin backwaters have come from Trenter and Abraham (1971); Madhupratap *et al.* (1975) and Madhupratap (1980). In this chapter the pattern of association of common groups and species of zooplankton in the eight estuaries are ^{discussed} attempted.

Correlation matrices for the groups and species were formed after converting their numbers to their respective logarithmic values. The formula used for obtaining the correlation coefficient was

$$r = \frac{\sum (x - \bar{x})(y - \bar{y})}{n \sigma_x \sigma_y}$$

The results are presented in Tables 10 and 11.

Correlation at group levels showed that most of the groups were positively correlated with each other in these estuaries except at Kandalakara and Beypore.

The carnivorous forms like hydromedusae, ctenophores and chaetognaths were significantly positively correlated with each other in most of the estuaries. Highly significant relation between ^{them} (P = < 0.001) was observed at Mahe. At Neendakara, Cochin and Beypore, Copepoda showed significant positive correlation only with a few groups while in the other estuaries copepods occurred along with the other groups over the same period.

Cladocera in general did not show significant relationship with other taxa. They were negatively correlated with most of the groups at Veli, Cochin and Beypore. This is because of the rather peculiar distribution exhibited by this group being more common during certain months during the low salinity period. However, they exhibited significant positive correlations with sergestids, nauplii and decapods at Karapuzha and Mahe.

With regard to the decapods and sergestids, they showed significant positive relations with other groups at varying levels in Veli, Karapuzha, Mahe and Kallai. But at Neendakara, Cochin and Beypore their association with other groups were not as significant as in other estuaries. Similar trends of associations were noticed for the macroplanktonic forms like nauplii larvae, polychaete larvae and cirripede larvae.

Appendicularia showed significant positive correlations with other groups at Koraput, Mahe and Mallai and negative relations at Neendakara and Cochin. Fish eggs exhibited more positive relations compared to fish larvae in most of the estuaries.

Estuaries generally become rich in zooplankton population during the saline period and most groups thrive in this season. Thus it is natural that most of the groups were positively inter-related. However, the groups consist of species which have different ecological significance. Thus the pattern of associations observed at group level provide only limited information. The associations of the common copepod species is treated separately. The more or less consistent positive correlations for most groups confirm that the seasonal fluctuations of these groups are consistent in the estuaries studied.

Although 51 species of copepods were identified from these estuaries only the common species were taken to examine the pattern of associations. They included species preferring low, medium and high saline conditions.

In general, the various high saline species showed significant positive correlations with each other although variations existed in the level of signifi-

ence in different estuaries. Highly significant correlations were observed among Acartia gartmani, A. guineensis, A. hutchinsoni, A. erythroneura, Centropages alcocki, Pseudodiaptomus surivilli and Acartia similis in the Kallai estuary. Similar relations existed in the Mahe estuary also but A. erythroneura was absent in these waters. In Cochin backwaters, A. gartmani, the most common species, showed positive but not significant correlations with other species. However, it was correlated with Pseudodiaptomus serricaudatus, P. surivilli ($P < 0.001$) and Acartia similis.

At Karapuzha highly significant correlations were observed among Acartia gartmani, A. guineensis, A. hutchinsoni, Centropages alcocki, Pseudodiaptomus serricaudatus and P. surivilli. Acartia pacifica and Acartia similis were negatively correlated with all other species.

Centropages alcocki showed significant correlations with higher saline species at Karapuzha, Kallai and Mahe but exhibited negative relations in other estuaries. Similarly Pseudodiaptomus surivilli also showed significant correlations with high saline forms in some estuaries but negative relation in others.

Acrocalanus girilis, another successful species in the estuarine waters ^{was} found to exist together with all the high saline forms in all the estuaries except at Korapuzha where ^{it} ^{was} they were associated with Acartia pacifica only.

Pseudodiaptomus sarsi, a highly tolerant species to wide salinity fluctuations was found to associate with both high saline and medium saline species.

Acartia plumosa and Acartiella keralensis, the two medium saline species were significantly positively correlated ($P < 0.001$) with each other in all estuaries except at Kallai. They were also observed to associate at higher levels of significance with A. bilobata at Neendakara estuary. The common low saline species Acartiella graveyxi almost always stood separate from other species exhibiting negative correlations except at Veli lake.

The pattern of correlations among the common copepod species shows that the low saline species Acartiella graveyxi and the medium saline species A. keralensis and Acartia plumosa occupy niches which stand separate from the high saline forms. Since species in these two categories are few and the biotope is sufficiently heterogeneous no biological factors could be limiting their flourishing in the estuaries. They are separated both

temporally and spatially from the high saline forms. Only physical forces like strong currents or chemical parameters such as salinity affect their survival.

On the other hand, the various high saline species exist in large abundance over the same span. These species in general showed significant positive correlations in most estuaries with minor variations. Spatially also they have similar distribution and this leads to either coexistence or competition among them. These results are in compliance with the earlier findings by Madhupratap (1980) from Cochin backwaters. Conceptually, if these species coexist, it would call for some degree of ecological differentiation between them to avoid niche overlap.

Earlier ecological theories, which are held valid to some extent even today, imply that competition cannot survive indefinitely. The competitive exclusion principle put forward by Gause (1934) is supported by Hardin (1960) who states that complete competitors cannot coexist. Slobodkin (1961) also holds the view that no two species can indefinitely continue to occupy the same ecological niche.

In aquatic communities, especially in microscopic zooplankton organisms, it is difficult to establish the

nature of ecological differentiations since visual observations are virtually impossible. Trentor and Abraham (1971) had attempted to seek the differentiations among the coexisting copepod species of the family Acartidae from the Cechin baculaters based on the structure of the mandibles. However, in their opinion "the differences which do exist are not sufficient to establish niche separation".

The absolute validity of competitive exclusion theory is, however, under criticism (Smith *et al.*, 1973; Hayward, 1980). While niche overlap is difficult to establish in marine organisms, several studies from the land have proved to be of inconclusive results (Rusterholz, 1981). The concept of a guild (a guild is a group of species which uses the same environment resources in a similar fashion, Root, 1967) is attaining considerable impetus in recent studies. Among the guild subtle differences in feeding mechanisms can alleviate intense competition by harvesting a portion of the resource spectrum unavailable to other species. Hutchinson (1961, 1965) and Halburd (1977) feel that species coexist, but not in equilibrium because equilibrium is attained only after the better competitors have excluded the poorer ones. Such equilibrium conditions are rarely

met with in nature. In coral reef fish communities where the diversity is very high, there is considerable overlap in space and food requirements among the coexisting species (Sale, 1977). Recent experimental studies on several coexisting copepod species have shown that they feed upon the same size range of particles when presented with natural prey, and that the preferred size varies in response to particle size distribution in the habitat (Foulet, 1978). ^{our} With our present knowledge, ecological differentiation among marine/estuarine zooplankton can perhaps only be defined empirically.

Studies (Cawey and Corner, 1963; Conover, 1966; Charvin, 1978; Rispar, 1978; Hayward, 1980) indicate that the spectrum of food used by copepods include detritus and bacteria as well, apart from the conventional phytoplankton-zooplankton link. The high primary production in the estuarine environments, availability of large amount of organic detritus and associated bacterial load provide the estuarine zooplankton with sufficient food resources. This also explains the large abundance of zooplankton in the estuaries compared to the marine environment. This availability of food and the probable variations that exist in feeding habits would allow the various high saline species to live as competitors or

even allow coexistence. Moreover the peculiarity of the tropical estuaries allow them to expand their niches spatially as the salinity increases towards the interiors. Succession of the high saline species starts from the mouth areas of the estuaries at the cessation of the monsoons and as salinity increases they invade the upper reaches utilising the resources available in these regions. Anyway, the periodical monsoons which washes out the high saline species out of the estuaries relieves them of the necessity to compete or co-exist for long, till a ^{the unlikely} situation where resources would run out.

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9. SUMMARY.

1. Ecology of zooplankton of eight estuarine systems of Kerala spread over a stretch of 500 km of the coastline was studied based on year-round collections made during 1978. The hydrographic data and zooplankton samples were taken from one fixed station at the mouth of each estuary. The middle and upper reaches of the Cochin backwaters were sampled seasonally. Of the various estuaries studied, those at Veli and Thottappilly have only seasonal connection with the sea when the sand-bar breaks during the monsoon. Veli lake is situated near Trivandrum and Thottappilly is located north of Alleppey. The other estuaries studied have perennial connections with the Arabian Sea viz. Mandoor or Ashtamudi estuary at Quilon, Cochin backwaters at Cochin, Korapuzha, Kallai, and Beypore estuaries at Calicut, and Mahé estuary near Talli-cherry. The general environment of these estuarine systems and other major features are reviewed in the context of other estuarine studies.

2. Both the southwest and northeast monsoons exert their influence on these estuaries. Salinity, the single-most important factor in the biological process in estuaries showed wide variations over the year. Conditions at the mouths ranged from almost marine to near freshwater during the different seasons except at Neendakara where the water column remained slightly saline even during the peak monsoon period. The cold less oxygenated high saline water at the bottom layers at the Cochin backwaters during certain months could be identified as up-welled Arabian Sea water entering the channel.

3. The water column was well mixed during the premonsoon period (January-April) in these estuaries. Steep vertical gradients in salinity, temperature and oxygen were noticed in the Cochin backwaters during the monsoon period (May-October). Such vertical stratification of the water column was less pronounced in other estuaries, especially at Neendakara and Mahé.

4. Veli and Thottappilly lakes differed from the other estuaries in the general hydrobiological aspects. The system remained practically freshwater during most of the year at Thottappilly. At

was low but bottom salinity

Veil, the surface salinity registered higher values during the premonsoon and postmonsoon seasons. The high saline water at the bottom in this lake in the absence of free connection with the sea could be explained partly by the dynamics of a coastal aquifer and also by spill over during intense wave action. The latter probably brings in the high saline fauna encountered in this lake.

5. Zooplankton biomass and total zooplankton counts varied significantly between seasons and estuaries. Highest biomass and total numbers of zooplankton were recorded from the Cechin backwaters; Korapuzha and Kallai estuaries ranked next. Maximum population was usually observed in April but the month when peak biomass was recorded differed from place to place showing total numbers and biomass were not always correlated. In general estuaries have a high standing stock and have higher turnover rates compared to the adjoining sea.

6. Seventy three species of zooplankton belonging to sixteen groups were identified from these estuaries. The species composition in the 8 estuaries studied were more or less similar.

However, some species did not occur in all the estuaries. Variations in the population of different species were also noticeable between estuaries. Maximum number of species was recorded from Cochin backwaters. Neendakara though poor in biomass and total numbers ranked second in species richness. Copepoda was usually the most abundant group followed by naupliar larvae and Sargastidae.

7. Carnivorous forms like Hydromedusae, Ctenophora and Chaetognaths were present during the high saline period. The common hydromedusae occurring in these waters were essentially brackish water forms, whereas Ctenophora and Chaetognaths comprised only of marine species. Chaetognaths were more abundant at Cochin backwaters than in other estuaries. These three groups together constituted only 4.3% of the total numbers, but their feeding exerted profound influence on other zooplankton groups especially Copepoda.

Cladocerans which are common in the neritic and coastal waters were curiously absent in the estuaries during the high saline periods. They.

on the other hand appeared during the low salinity regime. Higher primary production and lack of competition during the monsoon period seem to offer favourable conditions to them.

Copepoda^{WVS}, the dominant group in these estuaries constituting 67.7% of the total counts. Fifty one species belonging to 18 genera were identified. Families Acartiidae, Pseudodiaptomidae and Paracalanidae formed the bulk of the Copepod population. They together accounted for 65 to 92% of the total copepods in these estuaries except at Thottappilly lake. In this lake they formed only 51%, the rest being low saline species belonging to Diaptomidae and Cyclopida. *Acartia centura*, *A. spinicauda*, *A. bilobata*, *Paracalanus girilla*, *Pseudodiaptomus sarasinoides*, *P. annandalei*, *Centropages alcocki* and *Leptocera nectinata* were the common high saline copepod species in the estuaries. *A. plumosa* and *Acartiella hexalepis* preferred medium salinity and were the dominant forms in the middle and upper reaches of Cochin backwaters. *Acartiella gouveia* thrived well during the low salinity regime. *Acartia centura* emerged as the most abundant species in the three-way analysis of variance performed on the copepod species.

Unlike other estuaries a mixed assemblage of low, medium and high saline species of Copepods were observed at times at Thottappilly and Veli lakes. In the former where the salinity was low throughout the year, low saline species were relatively more abundant. The presence of marine species like Tamora stylifera, Tortanus gracilis and Acartia nannionna at Veli lake which lacks a permanent connection to the sea is a little intriguing. They might have come into this lake through the spill over during intense wave action and were able to survive because of the higher saline bottom water.

Decapod larvae constituted the majority of invertebrate larval forms ranking next to copepods in overall abundance. Zoa larvae were very common occurring in all seasons with peaks usually during the premonsoon months. Larvae of penaeid and caridean decapods at various stages of development including those of commercially important species were observed in all estuaries almost throughout the year. Janitor hansonii was the most common sergestid. Fish eggs and larvae were also common mostly belonging to families Ambassidae, Mugilidae and Gobidae.

Other groups like Siphonophora, Ostracoda, Comataea, Mysidacea, Appendicularia etc. were observed during the higher saline months but are of less importance in the estuarine ecology.

8. The zooplankton of the estuaries comprise of truly estuarine, euryhaline marine, a few stenohaline marine and fresh water species. Among the true estuarine species three clines - high saline, medium saline and low saline - were recognized. The three attain peak populations in different seasons depending on the environmental conditions.

9. The high saline species flourish during the premonsoon period. They are more abundant at the lower reaches but do invade the upper reaches along with salinity incursion. The medium saline species also flourish well during the saline period, but in the middle and upper reaches where salinity is optimum. Low saline species thrive at the upper reaches during postmonsoon months and throughout the estuary during monsoon period. Thus monsoonal inundation, tidal incursion and associated changes in salinity are the main factors controlling the zooplankton of the estuaries.

10. The occurrence of mixed assemblages of high and low saline copepod species in Veli and Thattappilly lakes which have limited connection to the sea, is very interesting. This debilitates the general idea that salinity is the factor controlling the occurrence of various species in the estuaries, though it does control the abundance of various estuarine species.

11. In these estuaries, phytoplankton, detritus and associated bacterial load form the main sources of food for the zooplankton. The species of the families Acartiidae, Pseudodiaptomidae and Paracalanidae which contributed to the bulk of the copepod population are mainly herbivorous, omnivorous or detritivorous. In general, food is not a limiting factor for the survival of zooplankton in these estuaries. The coefficient of energy transfer from the primary and secondary trophic level is low in the Cochin backwaters. The underutilised excess production both at primary and secondary levels probably contribute to the richness of the bottom communities and productivity of coastal waters.

12. In the tropical estuaries heavy discharge of fresh water during monsoon season wipes out all but a few zooplankton organisms. Repopulation of the estuaries start with the salinity recovery. The successional sequence that follows could be seen from the relative numerical abundance culminating in high zooplankton population during the saline period before the following monsoon disrupts further progress. Among the numerous species that occur in the estuaries a few dominate the zooplankton assemblage. The pattern of succession showed three series consisting of high saline, medium saline and low saline forms. The high saline forms dominated the mouth area during the premonsoon. The medium saline species were abundant at the middle reaches during early premonsoon and at the upper reaches during late premonsoon. The low saline species dominate during the monsoon period. Although the broad scheme of succession was similar individual variations did exist in different estuaries.

13. Species diversity of the estuaries was low compared to coastal and oceanic realms. Diversity increased in estuaries in the dry season when conditions tend to be more stable. The higher evenness of the species - population index observed at Thottappilly and Veli lakes was due to lesser numbers of

species corresponding to a thin population. Productivity and spatial heterogeneity being of little consequence, it is speculated that lack of stability leads to the lower diversity in estuaries.

14. Analysis of association of groups of zooplankton in the estuaries revealed that most of the groups exhibited significant correlations with each other. However, this is but natural since all the major groups occur in high numbers during the high saline months.

The common high saline copepod species showed high degree of correlations between themselves. The medium saline species like Acartia plumosa and Acartiella hutchinsonii showed significant correlation between each other, but was negatively correlated with most of the high saline species. Acartiella graueli, the low saline species almost always stood separate exhibiting negative correlation with all other species.

Considerable niche overlap seems to occur between the common high saline species. The ecological differentiation between the species to allow coexistence is not lucid. Perhaps the availability of large amount of food provide the estuarine zooplankton sufficient

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resources to survive as competitors. The high saline species are eliminated by periodic monsoons^b before competition reaches the point where resources would run out.

10. BIBLIOGRAPHY.

- *Alcock, A., 1911 Occurrence of freshwater molluscs in Indian Streams. *Nature*, 27: 214.
- *Annandale, H., 1907 The fauna of Brackish Ponds at Port Canning, Lower Bengal. 1. Introduction and preliminary account of the fauna. *Bull. Ind. Mus.*, 1.
- Annandale, H., & S. Kemp, 1915 Introduction to the fauna of Chilka Lake. *Bull. Ind. Mus.*, 5: 1-20.
- Ansari, Z.A., 1974 Macrobenthic production in Venbanad Lake. *Mahasagar - Bull. natn. Inst. Oceanogr.*, 7: 197-200.
- Bense, K., 1959 On upwelling and bottom trawling off the South West Coast of India. *J. mar. biol. Ass. India*, 1: 33-67.
- Bhargava, R.N.S. & S.N. Deyvedi, 1974 Diurnal variations in Phytoplankton pigments in Zuari estuary, Goa. *Indian J. mar. Sci.*, 3: 142-145.
- Bhattathiri, P.M.A., V.P. Devassy & K. Radhakrishna, 1980 Primary production in the Bay of Bengal during Southwest monsoon of 1978. *Mahasagar - Bull. natn. Inst. Oceanogr.*, 13 (4): 315-323.
- Birch, D.W., 1981 Dominance in marine ecosystems. *Amer. Natur.*, 118: 262-274.
- Bowden, K.F., 1960 Circulation and mixing in the Mersey estuary. I.A.S.H. *Comm. of Surface Waters*, Publ. 21: 352-360.
- _____ 1963 The mixing processes of a tidal estuary. *Intern. J. Air Water Pollution*, 7: 343-356.
- ^Y
_A Bowles, K.A., 1960 Statistical theory and methodology in science and engineering. John Wiley & Sons, New York.

- Caspers, H., 1967 Estuaries: Analysis of definitions and Biological considerations. In: *Estuaries, A.A.S.E. Publ., No. 93*, Washington D.C. : 6-8.
- Chandrasehan, P., 1963 Studies on zooplankton of the Godavari estuary. Ph.D. Thesis, Andhra University, Waltair: 1-163.
- Chandrasehan, P. & T.S.S. Rao, 1972 Tidal cycle studies in relation to zooplankton distribution in the Godavari estuary. *Proc. Indian Acad. Sci.*, 74: 23-31.
- Cheriyon, T., L.V.G. Rao & K.K. Varma, 1974 Hydrography and suspended sediment load of oceanic and estuarine waters adjoining Marmagoa Harbour during early Summer. *Indian J. Mar. Sci.*, 3: 99-106.
- Cheriyon, T., L.V.G. Rao & K.K. Varma, 1975 Variations in physical characteristics of the waters of Zuari estuary. *Indian J. Mar. Sci.*, 4: 5-10.
- Charvin, H.B., 1970 Assimilation of particulate organic carbon by estuarine and coastal Copepods. *Mar. Biol.*, 9: 265-275.
- Conover, R.J., 1966 Assimilation of organic matter by zooplankton. *Limnol. Oceanogr.*, 11: 338-345.
- Conover, R.J. & E.D.S. Conner, 1968 Respiration and nitrogen excretion by some marine zooplankton in relation to their life cycle. *J. Mar. Biol. Ass. U.K.*, 48: 49-75.
- Cooper, Jr. H.H., 1959 A hypothesis concerning dynamic balance of fresh water and salt water in a coastal aquifer. *J. Geophys. Sci.*, 6 (4): 461-467.
- Cowey, C.B. & E.D.S. Conner, 1963 On the nutrition and metabolism of zooplankton. II. The relationship between the marine Copepod *Calanus helgolandicus* and particulate material in Plymouth Sea water in terms of amino acid composition. *J. Mar. Biol. Ass. U.K.*, 43: 495-511.

- Daniel, R. & A. Daniel, 1963 On the siphonophores of the Bay of Bengal. 1. Madras coast. *J. MAR. BIOL. ASS. INDIA*, 5: 185-220.
- Derbshire, M., 1967 The surface waters of the coast of Kerala, South West India. *INDIAN J. MAR. SCI.*, 14: 295-320.
- Das, P.K., C.S. Murty & V.V.R. Varadachari, 1972 Flow characteristics of Coimbatore Canal connecting Mandovi and Zuari estuaries. *INDIAN J. MAR. SCI.*, 1: 93-102.
- Day, J.H., 1951 The ecology of South African estuaries. Part I. General considerations. *Trans. MAR. SCI. S. AFRICA*, 11: 53-91.
- _____, 1967 The biology of Kynna estuary, South Africa. In: *Estuaries, A.A.A.S. Publ.*, 2, Washington D.C.: 397-407.
- Day, J.H., H.A.H. Millard & A.D. Harrison, 1952 The ecology of South African estuaries. Part III. Kynna, a clear open estuary. *Trans. Roy. Soc. S. Africa*, 22: 367-413.
- Day, J.H., H.A.H. Millard & G.J. Brookhuysen, 1954 The ecology of South African estuaries. Part IV. The St. Lucia System. *Trans. Roy. Soc. S. Africa*, 24: 129-156.
- Day, J.H. & J.F.C. Morgans, 1956 The ecology of South African estuaries. Part VII. The biology of Durban Bay. *Ann. Natal. Mus.*, 11: 259-312.
- Deevey, G.E., 1971 The annual cycle in quantity and composition of the zooplankton of Sargasso Sea off Bermuda. 1. The upper 500 m. *Limnol. Oceanogr.*, 16: 219-240.
- Della Croce, N. & P. Venugopal, 1972 Distribution of marine Cladocerans in the Indian Ocean. *Mar. Biol.*, 15: 132-138.
- Devassy, V.P. & P.M.A. Bhattachari, 1974 Phytoplankton ecology of the Cochin Bacterioplankton. *Indian J. MAR. SCI.*, 3: 46-50.

- Devassy, V.P., 1979 Succession of organisms following
P.M.A. Bhattathiri & Trichodinium phenomenon.
S.Z. Qasin, Indian J. MAR. Sci., 8: 89-93.
-
- Fisher R.A., 1943 The relation between the number of
A.S. Corbet & species and the number of individuals
C.B. Williams, in a random sample of an animal
population. J. Anim. Ecol.,
12: 42-58.
- -----
THE FRESHWATER FOR EXISTENCE,
Williams and Wilkins, Baltimore:
163 pp.
- George, M.J., 1938 Observations on the plankton of
Cochin backwaters. Indian J. Fish.,
5: 375-401.
- George, M.J. & 1963 Surface salinity of Cochin back-
K.N. Krishna Kartha, water with reference to tide.
J. MAR. Biol. Ass. India, 5: 178-184.
- Glover, R.E., 1959 The pattern of freshwater flow in
a coastal aquifer. J. Geophys. Res.,
64 (4): 457-458.
- Gopinathan, C.K. & 1971 Silting in Navigational Channels of
S.Z. Qasin, the Cochin harbour area.
J. MAR. Biol. Ass. India, 13: 14-26.
- Gowani, S.C. & 1974 Ecology of Mandovi and Zuari
S.Y.S. Singhal, estuaries. Plankton community in
relation to hydrographic conditions
during monsoon months, 1972.
Indian J. MAR. Sci., 3: 51-57.
- Gowani, S.C. & 1977 Plankton studies in the estuarine
Srivakumar, A., system of Goa. Prog. Symp. Natn
Water Zool. Sci. Publ. UNESCO/IO:
226-241.

- Grindley, J.R., 1960 Estuarine plankton. In: Estuarine Ecology, (Ed.) Day, J.H. A.A. Balkema, Rotterdam, Cape Town: 117-146.
- Grindley, J.R. & T. Wooldrige, 1974 The plankton of Richards Bay. Hydrobiol. Bull., 9: 201-212.
- Hardin, G., 1960 The competitive exclusion principle. Science, 131: 1292-1297.
- Haridas, P., M. Madhupratap & T.S.S. Rao, 1973 Salinity, Temperature, Oxygen and zooplankton biomass of the backwaters from Cochin to Alleppey. Indian J. Mar. Sci., 2: 94-102.
- Haridas, P., P. Gopala Menon & M. Madhupratap, 1980 Annual variations in zooplankton from a polluted coastal environment. Mahasagar - Bull. natn. Inst. Océanogr., 13 (3): 239-248.
- Haridas, P. & T.S.S. Rao, 1981 Distribution of pelagic harpacticoid copepods from the Indian Ocean. Mahasagar - Bull. natn. Inst. Océanogr., 14 (2): 151-154.
- Hayward, T.L., 1980 Spatial and temporal feeding patterns of North Pacific Central Gyre. Mar. Biol., 58: 295-309.
- Hedgepeth, J.W., 1957 Treatise on marine ecology and paleoecology, Vol.1, The Geological Society of America, Memoir, 67.
- Heinle, D.R. & D.A. Flemer, 1975 Carbon requirements of a population of the estuarine Copepod Parhyalella affinis. Mar. Biol., 31: 235-247.
- Heinle, D.R., R.P. Harris, J.F. Ustach & D.A. Flemer, 1977 Detritus as food for estuarine Copepods. Mar. Biol., 40: 341-353.
- Heinrich, A.K., 1962 The life histories of plankton animals and seasonal cycles of plankton communities in the Oceans. J. Cons. Intern. Expl. Mer., 27: 15-24.

- Keip, C., 1974 A new index measuring evenness.
J. MAR. BIOL. ASS. U.K.
54: 355-357.
- Hiett, R.W., 1944 Food chains and food cycle in
the Hawaiian fish ponds.
Part I. The food and feeding
habits of mullet (*Mullus gambusia*),
milk fish (*Chanos chanos*) and
the ten pounder (*Kinga pichosta*).
Trans. Am. Fish. Soc. 74 (2):
250-261.
- Hulbert, E.M., 1977 Coexistence, equilibrium and
nutrient sharing among phytoplankton
species of the Gulf of Maine.
AMER. NATUR. 111: 967-980.
- Hutchinson, G.E., 1961 The paradox of plankton.
AMER. NATUR. 75: 137-145.
- _____., 1965 The ecological theatre and ev-
olutionary play. Yale University
Press, New Haven.
- I.C.A.R., 1964 Final report on hydrobiological
and faunistic survey of Godavari
estuarine system. *Rep. of I.C.A.R.*
Andhra University: 1-54.
- Jacob George,
K.S. Purushan &
N. Madhupratap, 1975 Distribution of planktonic ctenophores
along Southwest coast of India.
Indian J. MAR. Sci. 4: 201-202.
- Jeffries, H.P., 1962a Copepod indicator species in
estuaries. *Ecology*, 43: 730-733.
- _____., 1962b Salinity-Space distribution of
estuarine copepod genus *Eurytemora*.
Int. Rev. Hydrobiol. 47: 291-300.
- _____., 1962c Succession of two *Acartia* species
in estuaries. *Limnol. Oceanogr.*
7: 354-364.
- _____., 1962d Environmental characteristics of
Raritan Bay, a polluted estuary.
Limnol. Oceanogr. 7: 21-31.

- Jeffries, H.P., 1964 Comparative studies on estuarine zooplankton. *Limnol. Oceanogr.* 9: 348-358.
- John, V., & K.M. Alexander, 1968 A preliminary report on the hydrobiology of Moyere river for the year 1964. *Hydrobiologia*, 31: 492-496.
- Joseph, P.S., 1974 Nutrient distribution in the Cochin harbour and in its vicinity. *Indian J. Mar. Sci.*, 3: 28-32.
- Karunakaran, V., 1962 Development pattern of the Kerala economy and its impact on the environment. *Proc. Seminar on Development and Environment, Cochin*: 1-9.
- Ketchum, B.H., 1951 The exchange of fresh and salt waters in tidal estuaries. *J. Mar. Res.*, 10: 18-38.
- _____., 1954 Relation between circulation and planktonic population in estuaries. *Ecology*, 35: 191-200.
- Khosla, A.N., 1951 Appraisal of water resources: Analysis and utilisation of data. *Proc. U.N. Sci. Conf. Conseq. Util. Resources*: 4.
- Kinne, G., 1967 Physiology of estuarine organisms with special reference to salinity and temperature: General aspects. *Estuaria, A.A.A.S. Publ. No. 93*, Washington D.C.: 525-540.
- Konkur, R.G., M.S. Kusuma & B. Neelakantan, 1961 Plankton biomass in the Kali estuary, Karwar. Paper presented at the Seminar on Estuaries - Their Physics, Chemistry, Biology, Geology and Engineering aspects, 7-11 Dec., 1961, NIO, Goa (Abstract).
- Krishnamoorthy, K., 1961 Daily variations in marine plankton from Porto Novo, S. India. *J. Mar. Sci. India*, 13: 180-187.

- Krishnamoorthy, K. & V. Sundarraj, 1973 A survey of the environmental features in a section of the Vellar-Coleroon estuarine system, South India. *Ind. Biol.*, 21: 229-237.
- Nairan, C.V., 1972 Ecology of benthos in a tropical estuary. *Prog. Indian Natn. Sci. Acad.*, 28: 154-161.
- _____, 1977 Contributions to estuarine biology. Papers presented at the Third All Indian Symposium on Estuarine Biology, Cochin, Dept. of Marine Science, 1-1004.
- Lauff, G.H., 1967 Introduction. *Estuaries*, A.A.A.S. Publ. No. 51, Washington D.C., v-vi.
- Lala, S.N. & P.B. Gosw., 1935 Some common hydromedusae of the Bombay harbour. *J. Univ. Bombay*, 3 (5): 90-101.
- Mac Arthur, R.H., 1955 Fluctuations of animal populations and a measure of community stability. *Ecology*, 36: 533-536.
- _____, 1965 Patterns of species diversity. *Biol. Rev.*, 40: 510-533.
- _____, 1969 Patterns of communities in the tropics. *Biol. J. Linn. Soc.*, 1: 19-30.
- _____, 1972 *Geographical Ecology*. Harper and Row, New York.
- Madhupratap, M., 1978 Studies on the ecology of zooplankton of Cochin backwaters. *Mahasagar - Bull. Natn. Inst. Oceanogr.*, 21 (1 & 2): 45-56.
- _____, 1979 Distribution, community structure and species succession of Copepods from Cochin backwaters. *Indian J. Mar. Sci.*, 8: 1-8.
- _____, 1980 Ecological relations of coexisting copepod species from Cochin backwaters. *Mahasagar - Bull. Natn. Inst. Oceanogr.*, 23: 45-52.

- Madhupratap, M., 1981 Cladocera in the estuarine and coastal waters of south west coast of India. Mishra - Bull. natn. Inst. Oceanogr., 14: 215-219.
- Madhupratap, M. & P. Haridas, 1975 Composition and variations in the abundance of zooplankton of the backwaters from Cochin to Alleppey. Indian J. Mar. Sci., 4: 77-85.
- Madhupratap, M., T.S.S. Rao & P. Haridas, 1977 Secondary production in the Cochin backwaters, a tropical monsoonal estuary. Proc. Symp. Warm Water Zool. Sci. Publ. UNESCO/NIO, 515-519.
- Madhupratap, M., P. Venugopal & P. Haridas, 1979 Biochemical studies on some estuarine zooplankton species. Indian J. Mar. Sci., 8: 155-158.
- Madhupratap, M., S.R.S. Hair, C.T. Achuthankutty & V.R. Hair, 1981 Major crustacean groups and total zooplankton diversity around Andaman-Nicobar Islands. Indian J. Mar. Sci., 10: 266-269.
- Madhupratap, M., C.T. Achuthankutty, S.R.S. Hair & V.R. Hair, 1981 Zooplankton abundance in the Andaman Sea. Indian J. Mar. Sci., 10: 258-261.
- Manikoth, S. & K.Y.M. Salih, 1974 Distribution characteristics of nutrients in the estuarine complex of Cochin. Indian J. Mar. Sci., 3: 125-130.
- Margalef, R., 1968 Perspectives in Ecological Theory. The University of Chicago Press, 111 pp.
- Mathew, T. & N. Balakrishnan Nair, 1980 Phytoplankton of the Ashtamudy Estuary, Kerala. Indian J. Mar. Sci., 9: 253-257.
- Menon, H.R., P. Venugopal & S.C. Geewadi, 1971 Total biomass and faunistic composition of the zooplankton in the Cochin backwaters. J. Mar. Biol. Ass. India, 13: 220-225.

- Marthy, P.S.N., & M. Varghaya, 1972a Studies on the sediments of Vembanad Lake, Kerala State. Part I. Distribution of organic matter. Indian J. Mar. Sci., 1: 45-51.
- Marthy, P.S.N., & M. Varghaya, 1972b Studies on the sediments of Vembanad Lake, Kerala State. Part II. Distribution of Phosphorus. Indian J. Mar. Sci., 1: 106-115.
- Maragan, T., O. Divakaran, N. Balakrishnan Nair, & K.G. Padmanabhan, 1960 Distribution and seasonal variation of benthic fauna of the Veli lake, South west coast of India. Indian J. Mar. Sci., 2 (3): 184-188.
- *Nair, K.K., 1951 Medusae of the Trivandrum coast. Part I. Systematics. Bull. Cent. Res. Inst. Univ. Trivandrum (Ser. C. Nat. Sci), 2 (1): 47-75.
- Nair, K.K.C. & D.J. Tranter, 1971 Zooplankton distribution along salinity gradient in the Cochin backwater before and after monsoon. J. Mar. Biol. Ass. India, 13: 203-210.
- Nair, V.R., 1971 Seasonal fluctuations of Chaetognaths in the Cochin backwater. J. Mar. Biol. Ass. India, 13: 226-233.
- , 1973 Breeding and growth of Chaetognaths in the Cochin backwaters. ICRE Handbook, 3: 87-96.
- Nair, V.R., G. Peter & V.T. Paulinose, 1977 Zooplankton studies in the Indian Ocean, I. From Bay of Bengal during Southwest monsoon period. Maharajah - Bull. Natn. Inst. Oceanogr., 10: 43-54.
- Nair, V.R., G. Peter & V.T. Paulinose, 1978 Zooplankton studies in the Indian Ocean, II. Maharajah - Bull. Natn. Inst. Oceanogr., 11: 35-43.

- Nair, V.R., S.N. Gadhya, M. Jiyalalram & B.N. Dasai, 1981 Biomass and composition of zooplankton in Auranga, Ambika Purna and Mindola estuaries, of South Gujarat. *Indian J. Mar. Sci.*, 10: 116-122.
- Nair, S.R.S., Vijayalakshmi R. Nair, C.T. Achuthankutty & M. Madhuratap, 1981 Zooplankton composition and diversity in western Bay of Bengal. *J. Plankton. Res.*, 2 (4): 493-508.
- Natarajan, R., (Ed.) 1973 Recent researches in Estuarine Biology. Proceedings of Symposium on estuarine biology held at Annamalai University. Hindustan Publishing Corporation, New Delhi: 321 pp.
- Noale, I.M. & I.A.E. Bayly, 1974 Studies on the ecology of zooplankton of four estuaries in Victoria. *Aust. J. Mar. Freshwat. Res.*, 25: 337-350.
- Oden, E.P., 1971 Estuarine Ecology. ^{In:} Fundamentals of Ecology. W.B. Sanders Company: 352-362.
- Parikkar, N.K., 1951 Physiological aspects of adaptation to estuarine conditions. Prog. Indo-Pac. Fish. Council, 1950, Sec. 3: 168-175.
- Parikkar, N.K. & R.G. Aiyar, 1937 The brackish water fauna of Madras. Prog. Ind. Acad. Sci., 6: 284-337.
- Parulekar, A.H., S.H. Dabvedi & V.K. Dhargalkar, 1973 Ecology of clam beds in Mandovi, Cambarjua Canal and Zuari estuarine system of Goa. *Indian J. Mar. Sci.*, 2: 122-126.
- Parulekar, A.H., V.K. Dhargalkar & S.Y.S. Singhal, 1980 Benthic studies in Goa estuaries. Part III. Annual cycle of macrofaunal distribution, production and trophic relations. *Indian J. Mar. Sci.*, 9: 189-200.
- Patten, B.C. & 43 authors, 1975 Total ecosystem model for a cove in Lake Temosa. In: B.C. Patten (Ed.). Systems analysis and simulations in ecology. Vol. 1, Academic Press, New York: 205-421.

- Fettan, B.C. & E.P. Ghm, 1981 The cybernetic nature of Ecosystems. *ANNA. SAHUK.*, 118: 886-895.
- Pillai, P.P., 1971 Studies on the estuarine copepods of India. *J. MAR. BIOL. ASS. India*, 13: 162-172.
- Pillai, P.P. & M.A. Pillai, 1973 Tidal influence in the diel variations of zooplankton with special reference to Copepods in the Cochin backwaters. *J. MAR. BIOL. ASS. India*, 15: 411-417.
- Pillai, P.P., S.S. Qasim & A.K.K. Nair, 1973 Copepod component of zooplankton in a tropical estuary. *INDIAN J. MAR. SCI.*, 2: 38-46.
- Foulet, S.A., 1978 Comparison between five coexisting species of marine Copepods feeding on naturally occurring particulate matter. *Limnol. Oceanogr.*, 23: 1126-1143.
- Prichard, D.W., 1953a Salinity distribution and circulation in the Chesapeake Bay estuarine system. *J. MAR. RES.*, 11: 106-123.
- _____., 1953b Estuarine hydrography. *Adv. Geophys.*, 1, Academic Press Inc., New York: 243-280.
- _____., 1954 A study of the salt balance in a coastal plain estuary. *J. MAR. RES.*, 12: 133-144.
- _____., 1956 The dynamic structure of a coastal plain estuary. *J. MAR. RES.*, 15: 33-42.
- _____., 1967a What is an estuary: Physical view point. *Estuaries*, A.A.A.S. Publ. No.83, Washington D.C. : 3-5.
- Purushan, K.S., T. Balachandran & M. Sakthivel, 1974 Zooplankton abundance off the Kerala coast during February and April, 1970. *Mahasagar - Bull. Natn. Inst. Oceanogr.*, 7: 165-175.

- Oasin, S.Z., 1970 Some problems related to the food chain in a tropical estuary. Marine Food Chains, Oliver and Boyd, Edinburgh: 45-51.
- Oasin, S.Z. & C.V.G. Reddy, 1967 The estimation of plant pigments of Cochin backwater during the monsoon months. Bull. MK. Sci., 17: 95-110.
- Oasin, S.Z., P.M.A. Bhattachari & S.A.H. Abidi, 1968 Solar radiation and its penetration in a tropical estuary. J. gen. MK. Biol. Ecol., 7: 87-103.
- Oasin, S.Z. & C.K. Gopinathan, 1969 Tidal cycle and environmental features of Cochin backwater (A tropical estuary). Prog. Ind. Acad. Sci., 69: 336-348.
- Oasin, S.Z., S. Wallershausen, P.M.A. Bhattachari & S.A.H. Abidi, 1969 Organic production in a tropical estuary. J. gen. MK. Biol. Ecol., 7: 87-103.
- Oasin, S.Z., P.M.A. Bhattachari & V.P. Devassy, 1972 The influence of salinity on the rate of photosynthesis and abundance of some tropical zooplankton. MK. Biol., 12: 200-206.
- Oasin, S.Z., S. Vijayaraghavan, K.J. Joseph & V.K. Balachandran, 1974 Contribution of microplankton and nanoplankton in the waters of a tropical estuary. Indian J. MK. Sci., 3: 146-149.
- Oasin, S.Z. & M. Madhupratap, 1979 Changing ecology of Cochin backwaters. In: Contributions to Marine Sciences dedicated to Dr. C.V. Kurian, University of Cochin: 137-142.
- Radhakrishna, K., V.P. Devassy, P.M.A. Bhattachari & R.M.S. Bhargava, 1978a Primary productivity in the North-Eastern Arabian Sea. Indian J. MK. Sci., 7(2): 137-139.
- Radhakrishna, K., V.P. Devassy, R.M.S. Bhargava, & P.M.A. Bhattachari, 1978b Primary production in the Northern Arabian Sea. Indian J. MK. Sci., 7: 271-275.

- Ramanirthan, C.P. & 1960 Hydrographical features of the
R. Jayaraman, continental shelf waters off
Cochin during the years 1958 and
1959. *J. MAR. BIOL. ASS. INDIA*,
2 (2): 199-207.
- Ramanirthan, C.P. & 1963 Some aspects of the hydrographical
R. Jayaraman, conditions of the backwaters
around Willington Island (Cochin).
J. MAR. BIOL. ASS. INDIA, 5:170-177.
- Rameswathy, V.D., 1965 Comparative hydrographical studies
K. Krishnamoorthy & of the nearshore and estuarine
R. Seshadri, waters, Porto Novo, S. India.
J. Annamalai Univ., 25: 154-164.
- Rangarajan, K., 1959 Light penetration in the inshore
waters of Porto Novo. *PROG. IND.*
APPL. SCI., 49: 271-279.
- _____., 1973 Siphonophores obtained during the
cruises of R.V. VARUNA from the
west coast of India and the
Laccadive Sea. *J. MAR. BIOL. ASS.*
INDIA, 15: 125-159.
- Rao, K.K., 1974 Ecology of Mandovi and Zuari
estuaries, Goa: Distribution of
foraminifera assemblages.
Indian J. MAR. SCI., 3: 61-66.
- Rao, K.L., 1975 India's water health.
Orient Longman Limited,
New Delhi, 1-267.
- Rao, T.S.S., 1977 Salinity and distribution of
brackish water zooplankton in
Indian estuaries. *PROG. ENVIR.*
WATER WATER ZOOPL. SCI. Publ.
UNESCO/WHO: 194-304.
- Rao, T.S.S., 1975 Distribution of zooplankton in
M. Madhupratap & space and time in a tropical
P. Haridas, estuary. Paper presented at the
Symposium on Estuarine Biology.
Kanyakulam, 4-6, February, 1975.
- Raymond, J.E.G. & 1958 Quantitative studies on the zoo-
R.G.A. Currie, plankton of Southampton water.
ANN. REP. CHALLENGER SOC., 1: X.

- Raymond, J.E.G., 1969 Studies on marine zooplankton. R.T. Sreenivasagam & J.K.B. Raymond, IV. Investigations on *Microsetina*-phagous nauplius (N. Sars). *Deep Sea Res.* 16: 141-156.
- Reddy, C.V.G. & 1972 Phosphate regenerative activity V.N. Shankaranarayana, in the muds of a tropical estuary. *Indian J. Mar. Sci.* 1: 57-60.
- Ramani, K.N., 1980a Sediments of Cochin backwaters in P. Venugopal, K. Saraladevi & R.V. Unnithan, relation to pollution. *Indian J. Mar. Sci.* 9: 111-114.
- Ramani, K.N., 1980b Retting of coconut husk as a source of organic pollution in the Cochin backwaters. (Abstract). *Seminars on Human Environment*, BARC, Bombay.
- Ramani, K.N., 1981 Sediments of a retting yard. P. Venugopal, K. Saraladevi & R.V. Unnithan, *Indian J. Mar. Sci.* 10: 41-44.
- Ricklefs, R.E., 1973 *Ecology*, Nelson, London, 861 pp.
- Rieper, M., 1978 Bacteria as food for marine harpacticoid copepods. *Mar. Biol.* 45: 337-345.
- Rochford, D.J., 1951 Studies in Australian estuarine hydrology. 1. Introduction and comparative features. *Aust. J. Mar. Freshwat. Res.* 2: 1-116.
- Root, R.B., 1967 The niche exploitation pattern of the blue grey gnatcatcher. *Ecol. Monogr.* 37: 317-350.
- Rustenhals, K.A., 1981 Competition and structure of an avian foraging guild. *Auk. Harv.* 118: 173-190.
- Sakthivel, M. & 1974 Synchronisation in the occurrence P. Haridas, of *Erichthya* blooms and swarming of *Cyclops scutellaris* Rang (Pteropoda) and *Amphibia viviparica* Dana (Cladocera) in the area off Cochin. *Maharaja - Bull. Natn. Inst. Oceanogr.* 7 (1+2), 61-67

- Sale, P.F., 1977 Maintenance of high diversity in coral reef communities. *AMEL. NATUR.* 111: 337-339.
- Sanders, H.L., 1969 Benthic marine diversity and stability - Time hypothesis. Diversity and stability in ecological system, Brookhaven Symposia in Biology: 71-81.
- Sankaranarayanan, V.N., & S.Z. Qasim, 1969 Nutrients of the Cochin backwater in relation to environmental characteristics. *Mar. Biol.* 2: 236-245.
- Santhakumari, V. & N. Vannucci, 1971 Monsoonal fluctuations in the distribution of hydromedusae in the Cochin backwaters, 1968-69. *J. MAR. BIOL. ASS. INDIA*, 11: 211-219.
- Saraladevi, K., P. Venugopal, K.N. Ramani, S. Lalitha & R.V. Unnithan, 1979 Hydrographic features and water quality of Cochin backwaters in relation to industrial pollution. *Indian J. MAR. SCI.* 8: 141-145.
- Sarkar, S.K. & A. Choudhury, 1981 Seasonal variations of zooplankton in a tidal creek (Mooriganga) of Hooghly estuary, Sagar island, Sunderbans, India. Seminar on Estuaries - Their Physics, Chemistry, Biology, Geology and Engineering aspects, IIO, Goa, 7-11 December, 1981. (Abstract).
- Selvakumar, R.A., 1970 Cladoceran swarms in relation to rockshell fishery along the west coast of India. *CHER. SCI.* 12: 481-482.
- Selvakumar, R.A., V.R. Naik & M. Madhupratap, 1980 Seasonal variations in secondary production of the Mandovi-Zuari estuarine system of Goa. *Indian J. MAR. SCI.* 9: 7-9.
- Seshaiya, R.V., 1959 Some aspects of estuarine hydrology and biology. *CHER. SCI.* 11: 54.
- Sewell, R.B.S., 1924 Fauna of the Chilka Lake. Crustacea, Copepoda. *Mem. Indian Mus.* 3: 771-851.

- Shyama, C.S. & K.P. Balakrishnan, 1973 Diurnal variations of some physico-chemical factors in the Cochin backwaters during South-west monsoon. *J. Mar. Biol. Ass. India*, **15**: 391-398.
- Singhal, S.Y.S., 1973 Diurnal variations of some physico-chemical factors in the Zuari estuary of Goa. *Indian J. Mar. Sci.*, **2**: 90-93.
- Slabodkin, L.B., 1961 Growth and regulation of animal populations. Holt, Rinehart & Winston, New York: 184 pp.
- Smith, O.L., H.H. Shugart, R.V. O'Neill, R.S. Booth & D.C. Mc Naught, 1975 Resource competition and an analytical model of zooplankton feeding in phytoplankton. *Mar. Ecol. Prog. Ser.*, **102**: 371-381.
- Sreenivasan, M., 1971 Biology of cheetognaths of the estuarine waters of India. *J. Mar. Biol. Ass. India*, **13**: 173-181.
- Subbaraja, R.C. & K. Krishnamoorthy, 1972 Ecological aspects of plankton production. *Mar. Biol.*, **14**: 25-31.
- Sunderaraj, V. & K. Krishnamoorthy, 1981 The plankton potential of the mudflat and mangrove environments of Forte Novo, Seminar on Estuaries - their physics, chemistry, biology, geology and engineering aspects, 7-11 December, NIO, Goa. (Abstract).
- Suryanarayana Rao, S.V., & F.C. George, 1959 Hydrology of Korapuzha estuary, Malabar, Kerala State. *J. Mar. Biol. Ass. India*, **1** (2): 212-223.
- Taw, H. & D.A. Ritz, 1978 Zooplankton distribution in relation to hydrology of the Derwent River estuary. *Aust. J. Mar. Freshwat. Res.*, **29**: 763-775.
- Tranter, D.J. & S. Abraham, 1971 Coexistence of species of Acartiidae (Copepoda) in the Cochin backwaters, a monsoonal estuarine lagoon. *Mar. Biol.*, **11**: 222-241.

- Unnikhan, R.V.,
M. Vijayan &
K.N. Ramani, 1973 Organic pollution in Cochin
backwaters. *Indian J. Mar. Sci.*,
4: 39-42.
- Vannucci, M.,
V. Santhakumari &
S.P. Das Santos, 1970 The ecology of hydromedusae from
Cochin area. *Mar. Biol.*, 7:
49-58.
- Varkey John, 1971 Hydrobiological studies on the
freshwater conservatories of Kerala
with special reference to physiology
of certain specific groups.
Ph.D. Thesis, University of Kerala,
186 pp.
- Varna, K.K.,
L.V.G. Rao &
T. Cheriyan, 1973 Temporal and spatial variations in
hydrographic conditions of Mandovi
estuary. *Indian J. Mar. Sci.*,
4: 11-17.
- Venugopal, M. &
P.S.N. Murthy, 1974 Studies in the sediments of Vembanad
Lake, Kerala State. Part III. Distri-
bution and interpretation of bottom
sediments. *Indian J. Mar. Sci.*,
3: 16-27.
- Venugopal, P.,
P. Haridas,
M. Madhupratap &
T.S.S. Rao, 1979 Incidence of Red water along South
Kerala coast. *Indian J. Mar. Sci.*,
8: 94-97.
- Venugopal, P.,
K. Saraladevi,
K.N. Ramani &
R.V. Unnikhan 1980 Trace metal levels in the sediments
of Cochin backwaters. (Abstract).
Seminar on Human Environment,
IAC, Bombay.
- Wallersthaus, S., 1969 On the taxonomy of planktonic
Copepoda in the Cochin backwater
(A South Indian estuary).
Versoff. Inst. Meeresforsch. Bremerh.,
11: 245-266.
- _____., 1970 On the taxonomy of some Copepoda
in Cochin backwater (A South Indian
estuary). *Versoff. Inst. Meeresforsch.*
Bremerh., 11: 463-480.

- Wallerstein, S., 1974 Seasonal changes in the zooplankton population in the Cochin backwater (A South Indian estuary). *Hydrobiol. Bull.*, p: 213-223.
- Wickstead, J.H., 1963 The Cladocera of the Zanzibar area of the Indian Ocean, with a note on the comparative catches of two plankton nets. *J. Afr. Zool. Soc. I.*, 22: 164-172.

* Not referred to in original.

T A B L E S

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**Table 1: Annual river discharge from Indian Rivers
(After Khosla, 1951).**

West Coast Rivers	Km³/years
1. Streams from Cape Comorin to Tapti (Excluding Tapti).	114.2
2. Streams from Tapti to Tapti (excluding Tapti)	114.8
3. Tapti	9.1
4. Narmada	49.2
5. Tapi	9.3
6. Sabarmati	4.7
7. Streams of Katin	4.8
8. Luni	0.3
9. Streams of Rutch	3.7
10. Sutlej System	36.7
East Coast Rivers	
1. Rivers from Cape Comorin to Cauvery (excluding Cauvery).	7.9
2. Cauvery	10.0
3. Rivers between Cauvery and Pennar (excluding Pennar)	9.5
4. Rivers between Pennar and Krishna	1.6
5. Krishna System	44.9
6. Godavari System	125.5

Table 1 (Contd.)

	km^3/years
7. Rivers between Godavari and Mahanadi (excluding Mahanadi)	16.1
8. Mahanadi	92.8
9. Brahmani and Baitarni	39.2
10. Subarnarekha and streams from Subarnarekha to Baitarni	20.3
11. Subarnarekha to Damodar (excluding Damodar)	14.0
12. Damodar	13.0
13. Hooghly	17.3
14. Ganges System	489.8
15. Brahmaputra System	381.1
<hr/>	
Total for West Coast Rivers	346.8
Total for East Coast Rivers	1283.0

Table 2 A. Biomass (ml/10³) in the estuarine months in the year 1978.

Months	Velli	Koundalare	Theerappally	Cochin	Kallad	Beypore	Korapuzha	Maha
Premonsoon								
January	0.09	0.04	0.001	1.6	0.07	0.3	0.23	0.4
February	0.03	0.03	0.01	5.8	0.04	0.1	0.14	0.01
March	0.01	0.04	0.001	4.1	0.06	0.5	0.18	0.04
April	1.1	0.1	0.01	0.2	3.5	0.8	1.16	2.1
Monsoon								
May	0.25	0.03	0.001	7.0	1.9	0.5	1.3	0.001
June	0.001	0.05	0.001	0.9	0.2	0.2	0.3	0.03
July	0.02	0.03	0.001	5.1	-	-	0.001	0.01
August	0.01	0.2	0.01	0.8	0.4	0.001	0.001	0.001
September	0.001	0.01	0.001	0.2	0.8	0.001	0.07	0.001
October	0.2	0.04	0.01	1.4	0.2	0.001	0.4	0.001
Postmonsoon								
November	0.01	0.04	-	1.7	0.4	0.5	0.001	0.04
December	0.01	0.2	0.001	2.2	0.5	0.6	0.2	0.02
Annual mean	0.15	0.07	0.003	3.25	0.67	0.28	0.33	0.22
Seasonal average								
Premonsoon	0.31	0.05	0.005	4.9	0.96	0.4	0.42	0.64
Monsoon	0.08	0.06	0.003	2.6	0.60	0.11	0.34	0.007
Postmonsoon	0.01	1.2	0.0005	1.9	0.43	0.5	0.1	0.03

Table 2 B. Counts (No./10m²) of Zooplankton in the estuaries of Kerala coast during 1978.

Months	Velli	Neendakara	Thottappally	Cochin	Kallai	Beypore	Karepuzha	Maha
January	843	405	25	10450	1838	1964	2447	6275
February	591	588	572	41260	611	1068	2075	636
March	82	577	32	37190	845	4403	3353	1414
April	7861	2167	169	104167	30297	10721	12805	11138
May	8800	833	43	40270	15606	7376	12877	78
June	23	1436	43	6873	2957	1313	8402	948
July	1090	928	40	26890	-	-	76	200
August	246	2483	225	16012	28	162	180	30
September	7	155	78	2404	6560	27	285	63
October	2604	1384	1025	5804	810	289	4334	40
November	672	1495	-	8047	1666	13490	9	1645
December	258	3139	27	15839	2926	6636	3503	336
Annual mean	1940	1272	23	26337	5345	3971	4195	1899
SEASONAL MEANS								
Premonsoon	2342	930	199	48291	8398	4589	5170	4863
Monsoon	2162	1136	239	16375	4326	1528	4359	226
Postmonsoon	465	2317	13	11793	2296	10063	1756	990

Table 2 G. Zooplankton biomass (ml/10m³) and total counts (per 10m³) from the mouth to head of Cochin backwaters - 1978 (Values in parenthesis are total counts).

Months	S T A T I O N S						
	1	2	3	4	5	6	7
January	1.6 (10450)	1.3 (8730)	7.2 (4810)	2.9 (3362)	0.9 (2270)	9.6 (950)	0.7 (20)
April	0.2 (104107)	4.1 (12060)	20.0 (13960)	6.0 (46330)	2.0 (8160)	4.2 (1660)	1.1 (5320)
July	5.1 (26090)	0.06 (20)	0.04 (10)	0.05 (5)	0.06 (20)	0.02 (5)	0.02 (2)
November	1.7 (8047)	0.6 (940)	0.4 (290)	0.3 (160)	0.1 (10)	0.1 (3)	0.1 (10)

Table 3. Results of Analysis of Variance.

A. BIOMASS.

Source	SS	df	MS	F
Total	197.930	95		
Months	30.020	11	2.729	3.02**
Estuaries	98.434	7	14.062	15.59***
Error	69.476	77	0.902	

CD for months = 0.9307

CD for estuaries = 0.7309

** - $P < 0.01$

*** - $P < 0.001$

Table 3 (Contd.)

B. Total Burden.

Source	SS	df	ms	F
Total	109.538	95		
Months	23.131	11	2.103	3.33**
Estuaries	37.671	7	5.381	8.5***
Error	48.736	77	0.632	

CD for months = 0.7794

CD for estuaries = 0.6365

** - $P < 0.01$

*** - $P < 0.001$

Table 4. Result of three way ANOVA.

A. GRASSH.

Source	SS	df	MS	F
Total	1467.69	1439		
Months	205.67	11	18.69	33.31**
Estuaries	13.44	7	1.92	3.99**
Groups	197.54	14	14.11	25.14**
Months x estuaries	181.56	77	2.35	4.90**
Months x groups	129.03	154	0.83	1.49**
Estuaries x groups	221.78	98	2.26	4.70**
Error	518.63	1078	0.48	

CD for months = 0.207

CD for groups = 0.2312

CD for estuaries = 0.1579

** Significant at 1% level.

Table 4 (Contd.)

B. Common Canal species

Source	SS	df	MS	F
Total	855.36	1267		
Months	74.64	11	6.78	13.31**
Estuaries	13.96	7	1.99	5.60**
Species	152.91	12	12.74	24.99**
Months x estuaries	85.25	77	1.11	3.11*
Months x species	161.16	132	1.22	2.39*
Estuaries x species	30.81	84	0.36	1.03*
Error	336.59	944	0.35	

CD for months = 0.2287

CD for estuaries = 0.1617

CD for species = 0.2380

** Significant at 1% level

* Significant at 5% level.

Table 5. Seasonal distribution and annual mean (No./10m³) of various groups and their percentages to total counts in the estuaries - 1978.

YHJ

	Hydrozoans	Ctenophores	Cnidarians	Cladocera	Copepods	Amphipods	Decapod larvae	Other invertebrates	Appendicularia larvae	Fish eggs & larvae
Jan - Apr.	68	-	109	117	7345	-	764	318	96	-
May - June	301	-	200	-	3963	-	1300	373	153	-
July - Oct.	-	-	-	14	2484	66	1352	126	84	-
Annual mean.	30.7	-	25.7	22.5	1367.3	6.3	302.7	68.2	27.7	-
% to annual total	1.6	-	1.3	1.1	69.5	0.32	15.6	3.5	1.4	-

1
2
3
4

Table 5 (Contd.)

MERIDAYARA

	Hydro- medusae	Cteno- phora	Chaeto- gnatha	Clado- cnere	Cope- poda	Amphi- phoda	Decapod larvae	Other stidias	Appendi- calaria	Fish eggs & larvae	
Jan - Apr.	97	13	53	32	1341	49	970	61	11	140	
May - June	222	90	159	-	1310	-	287	71	24	-	
July - Oct.	31	4	218	491	2317	448	373	103	227	110	
Nov - Dec.	223	16	56	324	3452	-	112	68	25	74	
Annual mean.	67.7	10.2	40.5	69.7	701.7	45.5	145.2	93.3	28.1	4.3	27.0
% to annual total	3.7	0.8	3.2	5.5	55.2	3.6	11.4	7.3	2.2	0.34	2.1

Table 5 (Contd.)

ZIGTAGPILIA

	Hydro- medusae	Cteno- phore	Cteno- gastera	Clado- cera	Cope- poda	Amphi- poda	Decapod larvae	Other stidias	Inverte- brate larvae	Appendi- cularia	Fish eggs & larvae
Jan - Apr.	20	-	-	579	2	97	23	5	-	-	70
May - June	-	-	-	31	-	52	2	-	-	-	1
July - Oct.	-	-	84	1123	5	44	17	24	-	-	5
Nov - Dec.	-	-	1	17	-	4	1	1	-	-	3
Annual mean	1.6	-	7.1	145.8	0.6	16.4	3.75	2.5	-	-	6.6
% to annual total	0.87	-	3.7	76.8	0.3	8.6	1.9	1.3	-	-	3.5

Table 5 (Contd.)

COELENTERATA

	Hydrozoa	Ctenophora	Cnidaria	Copepoda	Amphipoda	Decapoda	Serpulidae	Other Invertebrates	Appendicularia	Fish eggs & larvae
Jan - Apr.	130	390	290	174090	390	12300	410	480	2680	1110
May - June	770	50	65	41130	100	3710	280	240	20	320
July - Oct.	-	-	20	27300	680	3125	213	90	20	1030
Nov - Dec.	20	20	90	12240	140	5620	20	220	920	110
Annual mean	76.7	37.5	39.7	305.0	169.2	2662.9	76.9	85.6	303.3	214.2
% of annual total	0.29	0.14	0.15	1.2	0.41	7.9	0.29	0.33	1.1	0.81

Table 5 (Contd.)

BALAK

	Hydre- medusae	Cteno- phore	Chaeto- gastra	Clado- cera	Cope- poda	Amphi- pods	Emerged larvae	Other Inverte- brate larvae	Appendi- cularia	Fish eggs & Fish larvae	
Estimation											
Jan - Apr.	905	991	9	-	23377	300	2319	1500	1530	662	600
Estimation											
May - June	197	196	-	-	11044	-	2936	1261	2940	-	1268
July - Oct.	-	-	-	10	1004	4	52	70	23	-	233 893
Estimation											
Nov - Dec.	205	-	263	-	2125	52	690	399	14	-	19
Annual mean	105.6	65.6	22.6	0.8	3212.5	29.7	463.1	269.2	273.6	71.8	173.6
% to annual total	2.0	1.26	0.43	0.02	61.7	0.57	9.3	5.2	7.3	1.4	3.4

Table 5 (Contd.)

INDONESIA

	Hydro- medusae	Cteno- phora	Charo- gaatha	Clado- cera	Cope- pada	Amphi- poda	Decapod larvae	Serge- stidae	Other inverte- brata	Appendi- cularia	Fish eggs & larvae
NUMBER											
Jan - Apr	4662	3553	258	-	5422	39	938	2810	380	64	73
			258								
May - June	128	2080	112	-	1274	-	1633	2987	64	-	226
July - Oct.	3	-	-	312	9	1	144	2	-	-	5
PERCENTAGE											
Nov - Dec.	495	713	289	6	11039	30	4312	2646	-	-	63
Annual mean	440.7	538.7	54.9	26.3	1478.7	5.0	585.6	658.7	37.0	5.3	30.6
% to annual total	11.1	13.3	1.4	0.7	37.2	0.15	16.7	15.6	0.90	0.13	0.77

Table 5 (Contd.)

FAUNA

	Hydre medusae	Ctenophore	Chaetognaths	Cladocera	Copepoda	Ampipoda	Decapod larvae	Sesipidinae	Other invertebrates	Appendicularia	Fish eggs & larvae
Jan - Apr.	420	223	884	30	9718	371	3202	1342	792	773	455
May - June	-	-	77	-	29	79	652	-	38	-	27
July - Oct.	-	-	-	-	36	36	135	3	-	-	3
Nov - Dec.	34	-	-	30	222	1066	118	2	61	32	299
Annual mean	37.8	18.7	75.9	5	803.7	129.3	342.2	112.2	74.2	67.3	65.3
% to annual total	1.9	0.98	3.9	0.26	43.8	6.8	17.9	5.9	3.9	3.5	3.4

Table 6: Systematic list of species identified from the eight estuaries of Kerala. (+ = Present; - = Absent).

Species	Veli	Thottappilly	Neendakara	Cochin	Kerupuzha	Kallal	Boyyappa	Mabe
Phylum - Ccelenterata								
Class - Hydroses								
Order - Hydroida								
Family - Campanularidae								
<i>Obelia</i> sp.	+	-	+	+	+	+	+	+
<i>Blackfordia viridinea</i> Mayer	+	+	+	+	-	+	+	-
Family - Lovenellidae								
<i>Echallota masoni</i> Kramp	-	-	+	-	-	-	-	-
Family - Sirenidae								
<i>Sirena sylvanensis</i> Bruno	+	+	+	+	+	+	+	+
<i>Sirena masoni</i> Kramp	-	-	+	-	-	-	+	-
Family - Batimidae								
<i>Batima summanalis</i> Santhakumari	+	+	+	+	+	+	+	+
<i>Batima masoniana</i> Uchida	-	-	+	-	-	-	-	-

Table 6 (Contd.)

Species	Velli Thottappally	Neendakara	Cochin	Kareepuzha	Kallai	Beypore	Maha
Order - Siphonophora							
Family - Diphyidae							
<i>Elathusa shandersonia</i> Hurley	-	-	+	+	+	+	-
<i>Lapsalis subuloides</i> (Lena & Van Kiemdi, 1971)	-	+	+	+	+	-	+
Phylum - Ctenophora							
Class - Tentaculata							
<i>Eleutherozoa gibbosa</i>	-	+	+	+	+	+	+
Class - Mada							
<i>Bacora</i> sp.	-	+	+	-	-	-	-
Phylum - Chaetognatha							
<i>Sacitta badoti</i> Be'raneck	+	+	+	+	+	+	+
<i>Sacitta inflata</i> Grassi	+	+	+	+	-	+	+
<i>Sacitta sarsenii</i> Gray	-	+	+	-	-	-	-
<i>Sacitta robusta</i> Doncaster	-	-	+	-	-	-	-

Table 6 (Contd.)

Species	Velli Thottappally	Neendakara	Cochin	Kareepuzha	Kallai	Beypore	Maha
Phylum - Arthropoda							
Class - Crustacea							
Order - Cladocera							
<i>Podon polyphemoides</i> Leubart	+	-	-	-	+	+	-
<i>Eubria karunakina</i> Claus	+	+	+	+	-	+	-
<i>Emilia miktonkris</i> Dana	-	+	+	-	-	+	-
<i>Daphnia</i> sp.	+	+	-	-	-	-	-
Order - Ostracoda							
Family - Halocyprididae							
<i>Hydrobia</i> sp. Th. Scott	-	-	+	-	-	-	-
Order - Cepopoda							
Sub Order- Calanoida							
Family - Calanidae							
<i>Scutocalanus</i> sp. (Giesbrecht)	-	-	+	-	-	-	-
<i>Undinula yulcoris</i> (Dana)	-	-	+	-	-	-	-

Table 6 (Contd.)

Species	Veli	Thettappally	Neendakara	Cochin	Korapuzha	Kallal	Boyyere	Mahe
Family - Eucalanidae								
<i>Eucalanus unguis</i> Giesbrecht	-	-	+	+	-	-	-	-
<i>Eucalanus attenuatus</i> (Dana)	-	-	+	-	-	-	-	-
Family - Paracalanidae								
<i>Paracalanus parvulus</i> Giesbrecht	+	+	+	+	+	+	+	+
<i>Paracalanus parvulus</i> bairdii, <i>gambianus</i> Kollerhaus	-	-	-	+	-	-	-	-
<i>Acartalanus similis</i> Sewall	+	+	+	+	+	+	+	+
<i>Acartalanus gibbus</i> Giesbrecht	+	-	+	+	+	-	-	+
Family - Centropagidae								
<i>Centropages alexisii</i> Sewall	+	+	+	+	+	+	+	+
<i>Centropages furcatus</i> (Dana)	-	-	+	+	+	+	+	-
<i>Centropages triandrus</i> Sewall	-	-	+	+	-	-	-	-
<i>Centropages tenuiremis</i> Thompson & Scott	-	-	+	+	+	-	-	-

Table 6 (Contd.)

Species	Vell	Thottappilly	Neendakara	Cochin	Korapuzha	Kallai	Beypore	Maha
Family - Diaptomidae								
<i>Haliolobos sinensis</i> Curney	+	+	-	+	-	-	-	-
<i>Allocladus mirabilis</i> Kiefer	+	+	-	+	-	-	-	-
Family - Pseudodiaptomidae								
<i>Archidiatomus spinosus</i> Machin & Haridas	-	+	-	+	-	-	-	-
<i>Pseudodiatomus annandalei</i> Sewell	+	+	+	+	+	+	+	+
<i>Pseudodiatomus manchani</i> Malayala & Wollershae	+	+	-	+	-	-	-	-
<i>Pseudodiatomus japoni</i> Kallai	+	-	+	+	+	+	+	+
<i>Pseudodiatomus suriyilli</i> Cleve	+	+	+	+	+	+	+	+
<i>Pseudodiatomus marioni</i> Frectl	-	-	+	+	+	+	+	+
<i>Pseudodiatomus scottii</i> (Y. Scott)	+	+	+	+	+	+	+	+
<i>Pseudodiatomus halliense</i> Sewell	-	-	-	+	-	-	-	-

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Table 6 (Contd.)

Species	Velli Thottappilly	Neendakare	Cochin	Kerapuzha	Kallai	Boypore	Maha
Family - Tenebridae							
<i>Tenebra barbifera</i> (Dana)	-	+	+	+	-	-	-
<i>Tenebra stylifera</i> (Dana)	+	-	+	+	-	-	-
Family - Lucicutidae							
<i>Lucicutia flavicornis</i> (Claus)	-	+	-	-	-	-	-
Family - Candacidae							
<i>Candacia hirsuta</i> A. Scott	-	-	+	-	-	-	-
Family - Pentellidae							
<i>Salmonella allardica</i> (Dana)	-	-	+	-	-	-	-
<i>Labidognathus neotropicalis</i> Thompson & Scott	+	-	+	+	+	+	+
<i>Labidognathus senilis</i> (Dana)	-	-	+	-	-	+	-
Family - Acartidae							
<i>Acartia senilis</i> Giesbrecht	+	+	+	+	+	+	+
<i>Acartia senilis</i> Giesbrecht	+	+	+	+	+	+	+
<i>Acartia senilis</i> Giesbrecht	+	-	+	-	+	-	-

Table 6 (Contd.)

Species	Veli	Thottappilly	Meendakara	Cochin	Karapuzha	Kallal	Beypore	Mahe
<i>Ascutia bilobata</i> Abrehan	+	+	+	+	+	+	+	+
<i>Ascutia boothiella</i> Sewall	-	-	+	-	-	-	-	+
<i>Ascutia mediana</i> Dana	+	-	-	+	-	-	-	-
<i>Ascutia plumosa</i> T. Scott	+	+	+	+	+	+	-	+
<i>Ascutia pacifica</i> Steur	+	+	+	+	+	-	+	+
<i>Ascutiella bangalensis</i> Wollersheim	+	+	+	+	+	+	-	-
<i>Ascutiella senaria</i> Sewall	+	+	-	+	+	+	-	-
Family - Tortanidae Tortanidae								172
<i>Tortania senaria</i> (Brady)	+	-	-	-	-	-	-	-
Sub order - Harpacticoida								
Family - Techididae								
<i>Platystrophia senaria</i> (Dana)	+	-	+	+	-	-	-	-
Family - Centrocampa								
<i>Centrocampa senaria</i> Boeck	+	+	+	+	-	-	-	-

Table 6 (Contd.)

Species	Velli	Thottappilly	Neendakara	Cochin	Karepuha	Kallai	Boypore	Maha
Family - Macrosetellidae								
<i>Macrosetella succilla</i> (Dana)	-	-	+	-	-	-	-	+
Sub Order - Cyclopoidea								
Family - Oithonidae								
<i>Oithona lubra</i> Giesbrecht	-	-	+	+	+	+	+	+
<i>Oithona dana</i> Giesbrecht	+	+	+	+	+	+	+	+
<i>Oithona kunda</i> Giesbrecht	-	-	+	+	-	-	-	-
<i>Oithona nimifera</i> Baird	-	-	+	-	-	-	-	-
<i>Oithona navisoma</i> Giesbrecht	-	-	-	+	-	-	-	-
Family - Ceryceus								
<i>Ceryceus</i> sp.	+	-	+	+	-	-	-	-
Family - Sapphirinidae								
<i>Sapphirina</i> sp.	-	-	+	-	-	-	-	-
Order - Mysidacea								
Family - Mysidae								
<i>Rhopalanthalmus indicus</i> Pillai	-	-	+	-	-	-	-	-

Table 6 (Contd.)

Species	Velli	Thettappally	Keendakara	Cochin	Karayana	Kallal	Myppore	Maho
Order - Decapoda								
Family - Penaeidae								
Subfamily - Seryestidae								
<i>Limulus</i> sp.	+	-	-	+	-	-	-	-
<i>Limulus</i> <i>limulus</i> Nobili	+	+	+	+	+	+	+	+
<i>Limulus</i> <i>limulus</i> H.M. Edw.	-	-	-	+	-	-	-	-
Phylum - Chordata								
Subphylum - Urochordata								
Class - Thaliacea								
Family - Salpidae								
<i>Thalia</i> <i>imbricata</i> Fernald	-	-	-	+	-	-	-	-
Class - Appendicularia								
<i>Salpinx</i> sp.	-	-	+	+	+	+	+	+

Table 7: Distribution of Groups/Species (those not represented in diagrams)
 in the different estuaries in 1978 - Nos./10m³ are given in parenthesis.
 * Represented in diagrams; - absent.

Species	Velli	Neendakara	Thettayalily	Cochin	Kallad	Buyyare	Korapuzha	Maha
HYDROPSUSAE								
Blattellidae Blattella	May (75)	*	Mar. (1); Apr. (3)	*	*	*	*	-
Buchelolidae Buchelolus	-	-	-	Dec. (17)	-	-	-	-
Elmidae Elmidae	Apr. (25) May (50)	*	Apr. (4)	*	*	*	*	*
Elmidae Elmidae	-	-	-	*	-	Mar. (19)	-	-
Elmidae Elmidae	Apr. (45) May (175)	*	Apr. (5)	*	*	*	*	*
SIPHONOPHORA								
Diapysa Diapysa	-	-	-	Dec. (14)	Mar. (19); May (294)	Nov. (22)	Apr. (12)	-
Lanella Lanella	-	Dec. (26)	-	Nov. (7)	Apr. (526)	-	Sept. (14)	May (9)

Table 7 (Contd.)

Species	Velli	Mendalure	Theppally	Cochin	Kallad	Bayyore	Karayanna	Mabe
CYNOCHORA								
Elanus leucurus	-	*	-	*	*	*	*	*
Buteo sp.	-	May (7)	-	Mar. (8) Apr. (3)	-	-	-	-
CHASTOGASTRA								
Scolecia bedfordi	Apr. (82) May (200)	*	-	*	Jan. (9) Nov. (54) Dec. (209)	*	*	*
S. senclaria	Apr. (27)	Jan. (3) Aug. (20) Oct. (5) Nov. (8) Dec. (30)	-	*	-	Jan. (2) Nov. (205)	*	Apr. (319)
S. senclaria	-	Apr. (13)	-	*	-	-	-	-
S. subarctica	-	-	-	Jan. (2) Dec. (18)	-	-	-	-
POLYCHAETE LARVAE	Feb. (8) Apr. (27) May (150) Jun. (3) Jul. (13) Aug. (71)	Jan. (5) May (14) Aug. (164) Dec. (30)	Apr. (4) Jul. (16) Sept. (8)	Jan. (10) Mar. (10) Jun. (190) Jul. (40) Sept. (10)	Mar. (13) Apr. (1053) May (96) Oct. (23)	Jan. (4) May (119)	Apr. (8) May (101) Oct. (171)	Jan. (411) Feb. (32) Apr. (53) Dec. (41)

Table 7 (Contd.)

Species	Velli	Meendakare	Thottappilly	Cochin	Kallad	Beypore	Kareppushe	Mahé
CIRRIFORME LARVAE	Jan. (3) Feb. (54) Mar. (4)	Jan. (34) Feb. (17) Mar. (3)	-	Jan. (40) Feb. (360) Mar. (20) Apr. (1404) May (1078) Jun. (200) Dec. (20)	Jan. (21) Mar. (39) Apr. (169) Apr. (52) May (64) Jun. (119)	Feb. (32) Mar. (169) Apr. (52) May (64) Jun. (119)	Mar. (10) Apr. (33) May (800) Jun. (119)	Jan. (276) Mar. (11) Dec. (4)
DECAPOD LARVAE	•	•	•	•	•	•	•	•
OSTRACODA	-	Apr. (7)	-	Apr. (12)	-	-	-	-
BRANCHIOPODA	-	Jan. (4) Feb. (2) Jul. (11) Aug. (226)	-	May (10) Jun. (15) Jul. (157) Nov. (6)	Apr. (8)	Mar. (30)	Mar. (9)	Mar. (3)
ISOPODA	-	-	-	May (12) Jun. (3) Oct. (4) Dec. (4)	-	-	Mar. (9)	-
MYSIDACEA	Jul. (13)	-	-	Jan. (10) Feb. (10) Mar. (20) Apr. (30)	-	-	-	-
SIRENOSTOMATA	Feb. (15) Mar. (3) Apr. (301) May (375) Oct. (126)	•	Jan. (14) Feb. (8) Apr. (13) Sept. (12)	•	•	•	•	•

Table 7 (Contd.)

Species	Velli	Koondakara	Thettappally	Cochin	Kallad	Beypore	Karapuzha	Maha
<i>Lactuca</i> <i>scariola</i>	-	-	*	-	-	-	-	-
COMPOSITAE								
<i>Centropogon</i> <i>indicus</i>	-	-	-	Feb. (3) Mar. (8)	-	-	-	-
<i>Erigeron</i> <i>indicus</i>	-	-	-	Jan. (10)	-	-	-	-
<i>Eucalyptus</i> <i>terrestris</i>	-	Apr. (20)	-	Jan. (7) Apr. (13)	-	-	-	-
<i>Eucalyptus</i> <i>nitens</i>	-	Aug. (20)	-	-	-	-	-	-
<i>Eucalyptus</i> <i>saligna</i>	Mar. (200)	-	Apr. (2)	-	-	-	Dec. (149) Nov. (8) Dec. (4)	-
<i>Eucalyptus</i> <i>camaldulensis</i>	-	-	-	-	-	-	-	-
<i>Aspalathus</i> <i>indica</i>	Apr. (1130) May (250)	-	Apr. (8)	-	-	-	Dec. (567)	-
<i>Aspalathus</i> <i>indica</i>	Dec. (2)	Dec. (136)	-	Apr. (112)	-	-	Dec. (110) Mar. (6)	-
<i>Centropogon</i> <i>indicus</i>	Mar. (329) Jul. (15)	-	Apr. (3)	-	-	Jan. (15) Nov. (188)	-	Apr. (691) Jan. (12)

Table 7 (Contd.)

Species	Velli	Neendakara	Thottaypilly	Cochin	Kallad	Beypore	Karayunha	Maha
<i>Cymbopogon</i> <i>axillaris</i>	-	Mar. (4) Dec. (33)	-	Jan. (16) Dec. (14)	Jan. (5)	Jan. (15)	Apr. (63)	-
<i>S. stramineus</i>	-	Jan. (4) Mar. (2) May (14) Nov. (17)	-	Jan. (12) Dec. (22)	-	-	-	-
<i>S. tenuis</i>	-	Aug. (20)	-	Mar. (63)	-	-	Sept. (9)	-
<i>Halodactylon</i> <i>glaberrimum</i>	Feb. (23)	-	Jan. (3) May (2) Aug. (6) Sept. (11) Oct. (342) Dec. (4)	Jul. (750) Aug. (60)	-	-	-	-
<i>Alloclonchium</i> <i>microclonchium</i>	Feb. (54)	-	Jan. (3) May (2) Aug. (10) Sept. (15) Oct. (552) Dec. (13)	Jul. (68)	-	-	-	-
<i>Amphidactylon</i> <i>microclonchium</i>	-	-	Dec. (2)	Oct. (16)	-	-	-	-
<i>Dactyloctenium</i> <i>spicatum</i>	†	-	Jan. (1) Aug. (8)	-	Mar. (26) May (137)	Jun. (69)	Jun. (795) Nov. (8) Dec. (4)	Mar. (548) Nov. (8) Dec. (4)

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Species	Velli	Kondakara	Thottappally	Cochin	Kallad	Beypore	Karepuzha	Mabe
<i>E. bairdianus</i>	Jan. (44) Mar. (2)	-	Feb. (214) Mar. (12)	Aug. (18)	Jun. (213) Oct. (11)	-	-	-
<i>E. bairdianus</i>	Jul. (13) Dec. (25)	-	Aug. (12) Sept. (4)	-	-	-	-	-
<i>E. jenseni</i>	Mar. (1) Dec. (19)	Jan. (2) Mar. (1)	-	-	Feb. (4)	Feb. (44) Dec. (20)	Feb. (260)	Jan. (69) Mar. (318)
<i>E. maculatus</i>	Apr. (178) Jun. (10) Jul. (63)	Mar. (2) Apr. (247) Aug. (123) Sept. (6)	Apr. (4)	May (40)	Apr. (1228) May (39) Dec. (55)	Jan. (7) Feb. (48) Mar. (50) Apr. (765) May (64) Dec. (145)	Jan. (65) Feb. (109) Mar. (5) May (606) Sept. (9)	Feb. (27) Apr. (32)
<i>E. maculatus</i>	-	Mar. (6) May (14)	-	Jun. (75)	Apr. (702)	Mar. (215)	Mar. (75) Apr. (101) May (28)	Jan. (137) Mar. (4) Apr. (425)
<i>E. maculatus</i>	Jul. (25) Dec. (6)	-	Apr. (7)	-	-	Jan. (79) Nov. (411) Dec. (12)	-	Jan. (69) Feb. (48) Apr. (479) Nov. (16)
<i>E. pallidus</i>	-	-	-	Sept. (5)	-	-	-	-
<i>E. pallidus</i>	-	Mar. (2) Jul. (5) Sept. (1) Dec. (22)	-	Jan. (28) Nov. (5)	-	-	Nov. (3)	-
<i>E. pallidus</i>	Mar. (2) Apr. (55)	-	-	Jan. (38)	-	-	Mar. (15)	-

Table 7 (Contd.)

Species	Velli	Heendakara	Thattappilly	Cochin	Palal	Beypore	Kerayutha	Maha
<i>Lepidocrypta flavicornis</i>	-	Aug. (20)	-	-	-	-	-	-
<i>Carduella leucosticta</i>	-	-	Apr. (3)	-	-	-	-	-
<i>Salpinctes laticaudata</i>	-	-	Jan. (6) Feb. (18) Mar. (25)	-	-	-	-	-
<i>Lobocorella punctata</i>	Jul. (12) Dec. (1)	-	-	-	-	-	-	-
<i>L. scintilla</i>	-	-	May (30)	-	Jan. (15)	-	-	-
<i>Asartia contracta</i>	-	Feb. (75) Apr. (34)	-	-	-	-	-	-
<i>A. spinicornis</i>	-	Apr. (23)	-	-	-	-	-	-
<i>A. senilis</i>	Mar. (12) Dec. (4)	Feb. (12) Mar. (61)	Apr. (740) Dec. (670)	Mar. (6) Apr. (1604)	-	-	-	-
<i>A. bilobata</i>	Mar. (1500)	-	Feb. (94) Apr. (19)	-	-	Apr. (1689)	-	-
<i>A. senilis</i>	-	-	-	-	-	-	Nov. (32)	-
<i>A. senilis</i>	Dec. (4)	-	Apr. (16)	-	-	-	-	-

1961

Table 7 (Contd.)

Species	Velli	Neendakara	Thottappally	Cochin	Kallei	Beypore	Kerapuzha	Mabe
<i>Acartia plumosa</i>	•	Oct. (300) Nov. (128)	Feb. (107) Mar. (7) May (10) Aug. (42)	•	Nov. (10)	-	•	Dec. (22)
<i>A. pacifica</i>	Apr. (822) May (425)	Mar. (4) Aug. (390)	Apr. (2)	•	-	Jan. (7)	Dec. (207)	Nov. (4)
<i>Acartiella karalensis</i>	•	Oct. (65)	Jan. (2) Mar. (3) May (12) Aug. (15)	•	Jan. (850)	-	•	-
<i>A. sumbrici</i>	•	-	Jan. (6) Feb. (133) Mar. (3) May (6) Aug. (30)	•	Jan. (745) Oct. (250)	-	•	-
<i>Tortanus carolinii</i>	Dec. (2)	-	-	-	-	-	-	-
<i>Subercula aculeicornis</i>	Mar. (2)	Jan. (3) Nov. (6) Dec. (52)	-	May (40)	-	-	-	-
<i>Nitocera similis</i>	Feb. (31)	Jan. (1) Feb. (19)	Feb. (12)	Jan. (12) Jan. (67)	-	-	-	-
<i>Macrosetella carolinii</i>	-	-	-	Mar. (6)	-	-	-	Apr. (8)

Table 7 (Contd.)

Species	Velli	Kondakara	Thettappilly	Cochin	Kalad	Baypare	Korapuzha	Maha
<i>Stethana bahadri</i>	-	Jan. (6) Feb. (52) Apr. (27) Nov. (87)	-	Jan. (8) Feb. (16) Apr. (43) Sept. (14) Nov. (5)	Mar. (6) Apr. (970) May (130)	Jan. (19)	May (606)	Jan. (4)
<i>Q. nana</i>	Apr. (110)	Feb. (9) Sept. (26) Nov. (21) Dec. (43)	Apr. (9)	Feb. (12) Apr. (24) Jun. (9) Aug. (7) Nov. (18)	Apr. (610) May (120) Oct. (11) Dec. (27)	Mar. (2354) Jun. (61) Oct. (8)	Jun. (448) Sept. (9) Oct. (113) Dec. (132)	Dec. (2)
<i>Q. flabida</i>	-	-	-	Feb. (7) Apr. (32)	-	-	-	-
<i>Q. nivalis</i>	-	Mar. (2)	-	-	-	-	-	-
<i>Q. basidensis</i>	-	-	-	Feb. (84) Jun. (10) Dec. (235)	-	-	-	-
<i>Campoplex</i> sp.	Mar. (2)	Mar. (6) Nov. (8) Dec. (57)	-	Jan. (136) Feb. (4) Mar. (18)	-	-	-	-
<i>Aspilota</i> sp.	-	-	-	Apr. (14)	-	-	-	-
ANTEROPODA	-	-	May (5) Aug. (5)	-	-	-	-	-

Table 6 : Occurrence of species of Cladocera (Nos./10m³) in different estuaries of Kerala in 1978.
(Months of occurrence are given in parenthesis)

Species	Velli	Neendakara	Thettappally	Cochin	Kallal	Boypore	Korapuzha	Mahe
<i>Bosmina longirostris</i>	10 (Dec.)	10 (Feb.) 70 (Aug.) 10 (Sept.) 320 (Dec.)	30 (Aug.)	20 (Jan.) 1700 (Aug.) 50 (Sept.) 3 (Oct.) 276 (Nov.) 2 (Dec.)	-	920 (Aug.) 110 (Oct.)	30 (Jan.) 60 (Aug.)	20 (Nov.)
<i>Bosmina</i> <i>avirostris</i>	-	350 (Aug.) 70 (Sept.)	-	2 (Jan.) 29 (Jul.) 1788 (Aug.) 12 (Oct.) 75 (Nov.)	-	700 (Aug.) 20 (Oct.)	110 (Aug.)	10 (Nov.)
<i>Bosmina longirostris</i>	10 (Jan.)	-	30 (Oct.)	-	10 (Oct.)	20 (Oct.)	-	-

Table 9 : Number of species observed, evenness index ($E = \frac{S}{S-1}$) and diversity index ($D = \frac{S-1}{\log^2 S}$) in the estuaries studied.

Months	Velli		Thottanally		Meendakara		Cochin	
	No. of species	E	No. of species	D	No. of species	D	No. of species	D
January	9	0.49	8	0.99	26	0.62	23	0.21
February	18	0.71	12	0.61	19	0.72	22	0.22
March	15	0.92	8	0.99	31	0.38	14	0.11
April	18	0.47	17	0.91	22	0.35	22	0.11
May	16	0.64	9	0.99	18	0.62	13	0.13
June	6	0.69	-	-	17	0.53	22	0.44
July	10	0.18	3	0.93	17	0.71	4	0.99
August	10	0.71	15	0.86	21	0.64	17	0.19
September	-	-	10	0.96	21	0.92	9	0.94
October	9	0.70	9	0.29	21	0.39	18	0.86
November	6	0.63	-	-	25	0.45	23	0.69
December	21	0.71	11	0.99	25	0.45	27	0.69

Table 9 (Contd.)

Months	Kaziranga		Kalla		Banyan		Maha					
	No. of species	D	No. of species	D	No. of species	D	No. of species	D				
January	18	0.32	3.1	20	0.50	3.6	29	0.47	5.2	24	0.62	3.6
February	16	0.67	2.8	21	0.59	4.8	11	0.47	2.1	16	0.56	3.5
March	17	0.28	2.8	22	0.62	4.7	25	0.27	3.9	21	0.34	4.0
April	16	0.43	2.1	24	0.60	2.9	20	0.25	2.7	24	0.68	3.3
May	20	0.67	2.6	20	0.64	2.6	17	0.33	2.4	4	0.99	2.2
June	14	0.39	1.9	10	0.71	1.6	11	0.65	2.0	12	0.22	2.5
July	3	0.86	1.0	-	-	-	-	-	-	4	0.89	1.0
August	-	-	-	3	0.99	1.8	-	-	-	-	-	-
September	14	0.74	3.8	2	0.89	0.56	3	0.91	1.4	7	0.93	2.7
October	14	0.44	2.1	7	0.19	1.4	14	0.65	4.1	3	0.91	1.4
November	3	0.99	1.8	5	0.39	1.5	12	0.44	1.5	21	0.35	3.9
December	13	0.65	2.0	15	0.26	2.5	25	0.22	3.7	30	0.78	5.2

(a, p < 0.1; b, p < 0.05; c, p < 0.02; d, p < 0.01; e, p < 0.001) YEM

	Hydromedusae	Ctenophora	Chaetognaths	Cladocera	Copepoda	Aphipoda	Decapoda	Caridea	Sergestidae	Zoea	Polydora	Appendicularia	Fish eggs	Fish larvae	Cirripede larvae
Hydromedusae	0.00														
Ctenophora	0.00														
Chaetognaths	0.95	0.00													
Cladocera	-0.17	0.00	-0.20												
Copepoda	0.78	0.00	0.90	-0.23											
Aphipoda	-0.18	0.00	-0.21	-0.16	-0.28										
Decapoda	0.15	0.00	0.14	-0.12	0.07	0.75									
Caridea	0.55	0.00	0.61	0.40	0.61	-0.33	-0.06								
Sergestidae	0.85	0.00	0.94	-0.23	0.94	-0.27	0.09	0.63							
Zoea	0.93	0.00	0.98	-0.23	0.93	-0.25	0.11	0.63	0.97						
Polydora	0.85	0.00	0.82	-0.21	0.65	0.11	0.16	0.43	0.78	0.78					
Appendicularia	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00				
Fish eggs	0.92	0.00	0.98	-0.21	0.88	-0.21	0.11	0.65	0.93	0.93	0.78	0.00			
Fish larvae	0.85	0.00	0.91	-0.14	0.81	-0.19	0.16	0.95	0.83	0.88	0.78	0.00	0.88		
Cirripede larvae	-0.12	0.00	-0.14	0.55	-0.16	-0.16	-0.14	-0.20	-0.14	-0.15	0.01	0.00	-0.13	-0.15	

-1891-

	Hydromedusa	Ctenophora	Chaetognaths	Cladocera	Copepoda	Amphipoda	Decapoda	Caridea	Sargassinidae	Eggs	Polychaeta	Appendicularia	Fish eggs	Fish larvae	Cladocera larvae
Hydromedusa	0.00														
Ctenophora	0.00	0.00													
Chaetognaths	0.00	0.00	0.00												
Cladocera	0.00	0.00	0.00	0.17											
Copepoda	0.00	0.00	0.00	0.00	0.00										
Amphipoda	0.00	0.00	0.00	0.00	0.00	0.00									
Decapoda	0.00	0.00	0.00	0.00	0.00	0.00	0.00								
Caridea	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00							
Sargassinidae	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00						
Eggs	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00					
Polychaeta	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00				
Appendicularia	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00			
Fish eggs	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
Fish larvae	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	
Cladocera larvae	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00

	Hydrorhiza	Chonophora	Chaetognaths	Cladocera	Copepoda	Amphipoda	Isopoda	Caridea	Serpentaria	Zona	Polychaeta	Appendicularia	Fish eggs	Fish larvae	Cirripede larvae
Hydrorhiza															
Chonophora	+0.61														
Chaetognaths	0.63	0.64													
Cladocera	-0.30	0.47	-0.26												
Copepoda	0.43	0.61	0.76	-0.18											
Amphipoda	-0.23	0.14	-0.01	0.21	0.51										
Isopoda	0.22	0.23	0.65	-0.23	0.73	0.30									
Caridea	-0.04	-0.02	-0.03	-0.17	0.23	0.53	0.09								
Serpentaria	0.43	0.55	0.34	-0.39	0.78	0.34	0.41	0.30							
Zona	0.04	0.43	0.73	-0.14	0.43	0.02	0.53	-0.14	-0.01						
Polychaeta	0.26	0.18	-0.32	-0.12	0.23	0.32	-0.43	0.17	0.14	-0.33					
Appendicularia	-0.04	0.03	0.37	0.03	0.17	-0.10	0.35	-0.53	-0.21	0.63	-0.12				
Fish eggs	0.26	0.38	0.77	0.18	0.37	-0.07	0.47	-0.43	0.31	0.64	-0.43	0.51			
Fish larvae	0.04	0.09	0.05	-0.21	0.53	0.47	0.31	0.93	0.53	-0.02	0.22	-0.29	-0.36		
Cirripede larvae	-0.09	0.06	0.19	0.07	-0.03	-0.05	0.19	-0.43	-0.41	0.57	-0.04	0.63	0.63	-0.43	

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	Hydrorhizans	Chemosphaera	Chaetognaths	Cladocera	Copepoda	Amphipoda	Isopoda	Caridea	Scudicoida	Polychaeta	Appendicularia	Fish eggs	Fish larvae	Cladipoda larvae
Hydrorhizans	0.97													
Chemosphaera	0.19	-0.13												
Chaetognaths	0.00	0.00	0.00											
Cladocera	0.93	0.98	-0.08	0.00										
Copepoda	-0.15	-0.14	-0.07	0.00	-0.11									
Amphipoda	0.32	0.55	-0.14	0.00	0.55	-0.11								
Isopoda	0.93	0.98	-0.11	0.00	0.98	-0.15	0.41							
Caridea	0.01	0.92	0.09	0.00	0.01	-0.16	0.73	0.01						
Scudicoida	0.61	0.02	-0.13	0.00	0.73	-0.15	0.98	0.01	0.92					
Polychaeta	0.93	0.98	-0.09	0.00	0.93	-0.10	0.21	0.98	0.51	0.97				
Appendicularia	0.01	0.01	-0.11	0.00	0.01	0.06	0.11	0.92	0.64	0.41	0.97			
Fish eggs	0.35	0.64	-0.15	0.00	0.53	-0.12	0.98	0.43	0.73	0.93	0.23	0.14		
Fish larvae	0.98	0.73	-0.18	0.00	0.73	-0.07	0.01	0.71	0.73	0.01	0.53	0.01	0.01	
Cladipoda larvae	0.01	0.97	-0.15	0.00	0.93	-0.14	0.73	0.91	0.91	0.01	0.01	0.73	0.73	0.07

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	Hydromedusae	Ctenophora	Chaetognaths	Cladocera	Copepoda	Amphipoda	Decapoda	Caridea	Zoea	Sargassidae	Polychaeta	Appendicularia	Fish eggs	Fish larvae	Cirripede larvae
Hydromedusae	0.65														
Ctenophora	-0.15	0.02													
Chaetognaths	-0.06	-0.06	0.85												
Cladocera	0.55	0.38	-0.26	0.12											
Copepoda	0.58	0.58	0.10	0.00	0.17										
Amphipoda	0.02	0.09	-0.09	-0.03	0.65	-0.11									
Decapoda	0.85	0.55	0.32	0.45	0.45	0.55	0.04								
Caridea	0.02	0.09	-0.15	-0.09	0.65	-0.09	0.95	0.01							
Zoea	0.01	-0.12	-0.03	-0.12	-0.07	-0.14	-0.08	0.05	-0.09						
Sargassidae	0.01	-0.12	-0.04	-0.12	-0.07	-0.14	-0.07	0.05	-0.09	0.95					
Polychaeta	0.25	0.45	-0.09	-0.13	0.06	0.56	0.04	0.16	0.19	-0.20	-0.20				
Appendicularia	0.00	0.40	0.06	-0.14	0.07	0.45	0.29	-0.02	0.45	-0.01	-0.01	0.85			
Fish eggs	0.23	0.27	-0.03	-0.14	0.07	0.27	-0.16	0.25	-0.17	0.65	0.65	-0.01	0.12		
Fish larvae	-0.11	-0.15	-0.17	-0.05	0.45	-0.15	-0.10	-0.11	-0.10	-0.09	-0.09	-0.01	-0.21	-0.02	

Table 10 (Contd.)

1961

	Hydranthes	Stenophora	Chaetognaths	Cladocera	Copepods	Amphipods	Caridean	Serpentidae	Zoa	Polytheca	Appendicularia	Fish eggs	Fish larvae	Cirripede larvae
Hydranthes	0.01	0.51	0.37											
Stenophora				0.17										
Chaetognaths					0.44									
Cladocera	-0.15	0.98	0.17											
Copepods	0.29	0.51	0.61	0.44										
Amphipods	-0.14	-0.16	-0.22	-0.14	-0.15									
Decapoda	0.68	0.02	0.78	-0.11	0.68	-0.17								
Caridean	0.13	0.38	0.21	0.38	0.09	-0.01	-0.27							
Serpentidae	0.48	0.88	0.96	0.72	0.72	-0.24	0.44	0.37						
Zoa	0.53	0.71	0.33	0.63	0.48	-0.20	0.18	0.68	0.83					
Polytheca	0.48	0.01	0.48	-0.08	0.40	0.57	0.67	-0.23	0.22	0.04				
Appendicularia	0.51	0.61	0.86	0.53	0.77	-0.22	0.78	0.12	0.82	0.53	0.53			
Fish eggs	0.27	0.48	0.48	0.41	0.92	-0.28	0.66	-0.12	0.78	0.41	0.28	0.68		
Fish larvae	0.19	-0.25	0.56	-0.27	0.39	0.31	0.41	0.33	0.00	-0.01	0.47	0.12	0.12	
Cirripede larvae	0.61	0.48	0.82	0.29	0.88	-0.22	0.92	-0.09	0.72	0.48	0.61	0.78	0.28	0.28

Table 10 (Contd.)

INDEX

	Hydroneuridae	Ctenophora	Chaetognaths	Cladocera	Copepoda	Amphipoda	Decapoda	Caridea	Sergestidae	Zoea	Polydora	Appendicularia	Fish eggs	Fish larvae	Cirripede larvae
Hydroneuridae	0.83	0.82	0.97												
Ctenophora				Chaetognaths											
Chaetognaths															
Cladocera	-0.08	-0.13	0.03												
Copepoda	0.83	0.93	0.97	0.17											
Amphipoda	0.09	0.14	0.13	0.40	0.13										
Decapoda	0.83	0.98	0.97	-0.14	0.93	0.14									
Caridea	0.74	0.82	0.92	0.36	0.98	0.08	0.82								
Sergestidae	0.58	0.68	0.73	0.61	0.83	0.04	0.58	0.93							
Zoea	0.41	0.42	0.68	0.61	0.83	0.03	0.48	0.88	0.87						
Polydora	0.06	0.00	0.19	0.88	0.34	-0.03	0.00	0.53	0.78	0.81					
Appendicularia	0.74	0.78	0.88	0.42	0.98	0.06	0.73	0.93	0.98	0.82	0.63				
Fish eggs	0.81	0.98	0.97	-0.15	0.91	0.13	0.93	0.81	0.57	0.47	-0.003	0.78			
Fish larvae	0.42	0.39	0.37	0.40	0.39	0.83	0.35	0.32	0.29	0.26	0.11	0.34	0.34		
Cirripede larvae	-0.02	-0.10	0.09	0.87	0.24	-0.07	-0.11	0.47	0.73	0.72	0.98	0.53	-0.11	0.03	

TABLE 11. Correlation matrix for the common estuarine species in the different estuaries - 1978. (a, p < 0.1; b, p < 0.05; c, p < 0.02; d, p < 0.01; e, p < 0.001)

	<i>P. maritima</i>	<i>P. maritima</i>	<i>P. maritima</i>	<i>P. maritima</i>	<i>P. maritima</i>	<i>P. maritima</i>	<i>P. maritima</i>	<i>P. maritima</i>	<i>P. maritima</i>	<i>P. maritima</i>	<i>P. maritima</i>	<i>P. maritima</i>	<i>P. maritima</i>
<i>Acetia sentosa</i>	0.99												
<i>A. minima</i>	0.99	0.00											
<i>A. bilobata</i>	0.99	0.00	0.00										
<i>A. erythroa</i>	0.99	0.00	0.00	0.00									
<i>A. pacifica</i>	0.99	0.99 ^c	0.00	0.00	0.00								
<i>A. plumosa</i>	0.00	0.00	0.00	0.00	0.00	0.00							
<i>A. huxleyana</i>	0.00	0.00	0.00	0.00	0.00	0.00	0.99						
<i>A. annulata</i>	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00				
<i>Centronus alcocki</i>	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00			
<i>Zoodierantona</i> <i>maritima</i>	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00		
<i>Z. mundula</i>	0.99	0.00	0.99	0.00	0.00	0.00	0.99	0.99	0.99	0.99	0.00	0.00	
<i>Z. murvilli</i>	0.99	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00
<i>Acetia sentosa</i>	0.99	0.99	0.00	0.00	0.99	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.99

Table 11 (Contd.)

MINERALIA

	Alkali metals	Alkaline earth	Alkali metals	Alkaline earth	Alkali metals	Alkaline earth	Alkali metals	Alkaline earth	Alkali metals	Alkaline earth	Alkali metals	Alkaline earth
<i>Amelia smithii</i>												
<i>A. minima</i>	-0.13											
<i>A. bilobata</i>	0.41	0.15										
<i>A. striatella</i>	0.78	-0.25	-0.21									
<i>A. insularis</i>	-0.31	0.38	-0.20	-0.10								
<i>A. minuta</i>	0.28	-0.32	0.78	-0.14	-0.13							
<i>A. insularis</i>	0.14	-0.34	0.63	-0.10	-0.10	0.91						
<i>A. insularis</i>	-0.27	-0.13	-0.18	-0.10	-0.10	0.09	0.09					
<i>Centropus alenhi</i>	-0.24	0.65	0.09	-0.27	0.81	-0.02	-0.04	0.24				
<i>Paradolichopus auriculatus</i>	0.78	0.07	0.03	0.98	0.01	-0.12	-0.12	-0.14	0.03			
<i>P. sandaloi</i>	0.48	0.21	0.81	-0.18	-0.17	0.42	0.12	-0.15	0.13	0.10		
<i>P. aurivillii</i>	-0.30	0.23	-0.26	-0.15	0.36	-0.17	-0.13	-0.10	0.15	-0.13	-0.22	
<i>Amomalanus alenhi</i>	0.74	-0.36	-0.09	0.98	-0.16	0.02	0.03	-0.12	-0.32	0.88	-0.11	-0.12

Table 11 (Contd.)

ESCHERICH

	ESCHERICH	ESCHERICH	ESCHERICH	ESCHERICH	ESCHERICH	ESCHERICH	ESCHERICH	ESCHERICH	ESCHERICH
<i>Escherichia</i>									
<i>E. aminicanda</i>	0.43	0.41	0.28	0.30	0.19	-0.23	-0.24	0.15	0.08
<i>E. blubata</i>	0.41	0.41	0.28	0.30	0.19	-0.23	-0.24	0.15	0.43
<i>E. burthous</i>	0.28	0.28	0.28	0.30	0.19	-0.23	-0.24	0.15	0.43
<i>E. pacifica</i>	0.30	0.30	0.28	0.30	0.19	-0.23	-0.24	0.15	0.43
<i>E. blumosa</i>	0.19	0.19	0.28	0.30	0.19	-0.23	-0.24	0.15	0.43
<i>E. burthous</i>	-0.23	-0.23	0.28	0.30	0.19	-0.23	-0.24	0.15	0.43
<i>E. burthous</i>	-0.24	-0.24	0.28	0.30	0.19	-0.23	-0.24	0.15	0.43
<i>Enterobacter aerogenes</i>	0.15	0.15	0.28	0.30	0.19	-0.23	-0.24	0.15	0.43
<i>Enterobacter aerogenes</i>	0.08	0.08	0.28	0.30	0.19	-0.23	-0.24	0.15	0.43
<i>E. amandae</i>	0.43	0.43	0.28	0.30	0.19	-0.23	-0.24	0.15	0.43
<i>E. burthous</i>	0.08	0.08	0.28	0.30	0.19	-0.23	-0.24	0.15	0.43
<i>Enterobacter amandae</i>	0.58	0.58	0.28	0.30	0.19	-0.23	-0.24	0.15	0.43

Table 11 (Contd.)

	0.00	0.76	0.00	0.18	-0.23	-0.26	-0.20	0.63	0.67	-0.22	0.63	0.18
<i>Asartia umbonata</i>	0.00	0.76	0.00	0.18	-0.23	-0.26	-0.20	0.63	0.67	-0.22	0.63	0.18
<i>A. sublineata</i>	0.00	0.52	0.00	0.13	-0.01	-0.16	-0.13	0.97	0.93	-0.13	0.93	0.00
<i>A. bilobata</i>	0.00	0.00	0.00	-0.13	-0.01	-0.02	-0.14	0.00	0.24	0.01	0.00	0.00
<i>A. striatipes</i>	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	-0.11	-0.09	0.97	0.00
<i>A. insidiosa</i>	0.18	-0.13	-0.13	0.00	0.00	0.00	0.00	0.00	-0.11	-0.14	-0.15	-0.13
<i>A. lineata</i>	-0.23	-0.14	-0.01	0.00	-0.09	-0.11	0.97	0.97	0.97	0.97	0.97	-0.09
<i>A. lineolata</i>	-0.26	-0.16	-0.02	0.00	-0.11	0.97	0.97	0.97	0.97	0.97	0.97	-0.12
<i>A. insularis</i>	-0.20	-0.13	-0.14	0.00	-0.09	-0.01	0.12	0.12	0.12	0.12	0.12	0.12
<i>A. sumatrana</i>	0.63	0.97	0.40	0.00	-0.11	-0.11	-0.12	-0.10	-0.10	-0.10	-0.10	-0.10
<i>Centronema alcocki</i>	0.67	0.93	0.24	0.00	-0.11	-0.14	-0.15	-0.13	0.93	0.93	0.93	0.93
<i>Paradialanema</i> <i>sumatranum</i>	-0.22	-0.13	0.01	0.00	-0.09	0.97	0.97	0.97	0.97	0.97	0.97	-0.10
<i>P. sumatranum</i>	0.63	0.97	0.26	0.00	-0.12	-0.13	-0.14	-0.12	0.97	0.97	0.97	0.97
<i>Assemblanema alcocki</i>	0.18	-0.14	-0.15	0.00	0.97	-0.12	-0.12	-0.10	-0.10	-0.10	-0.10	-0.10

STATISTICAL ANALYSIS

STATISTICAL

STATISTICAL

STATISTICAL ANALYSIS

STATISTICAL ANALYSIS

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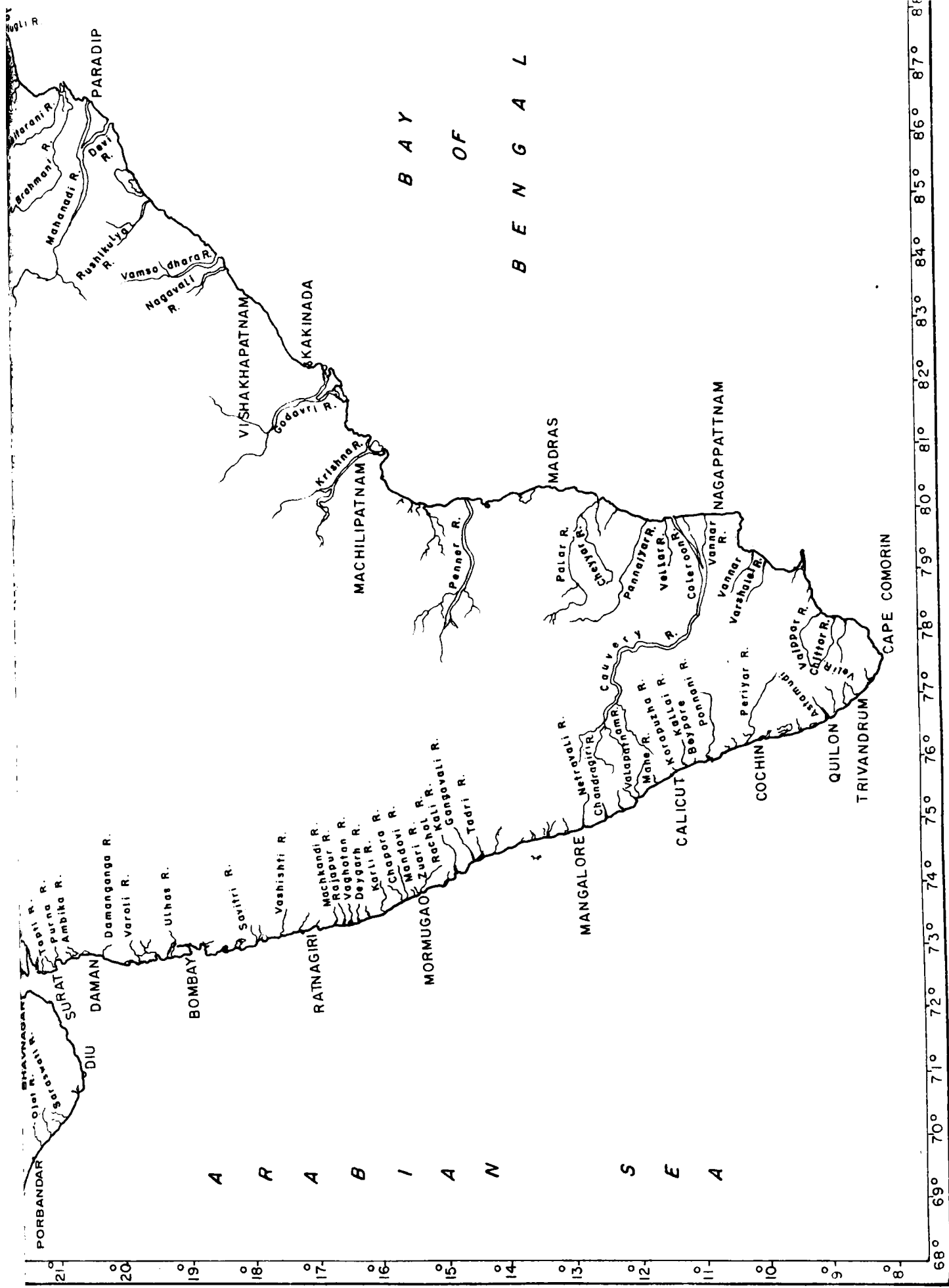
STATISTICAL

STATISTICAL

STATISTICAL

STATISTICAL

Fig. 1. Map showing the estuaries along the east and west coast of India.



A R A B I A N S E A

B A Y
O F
B E N G A L



Fig. 2. Map showing the location of the Stations (indicated by arrows) at the mouth of the estuaries where collections were made.

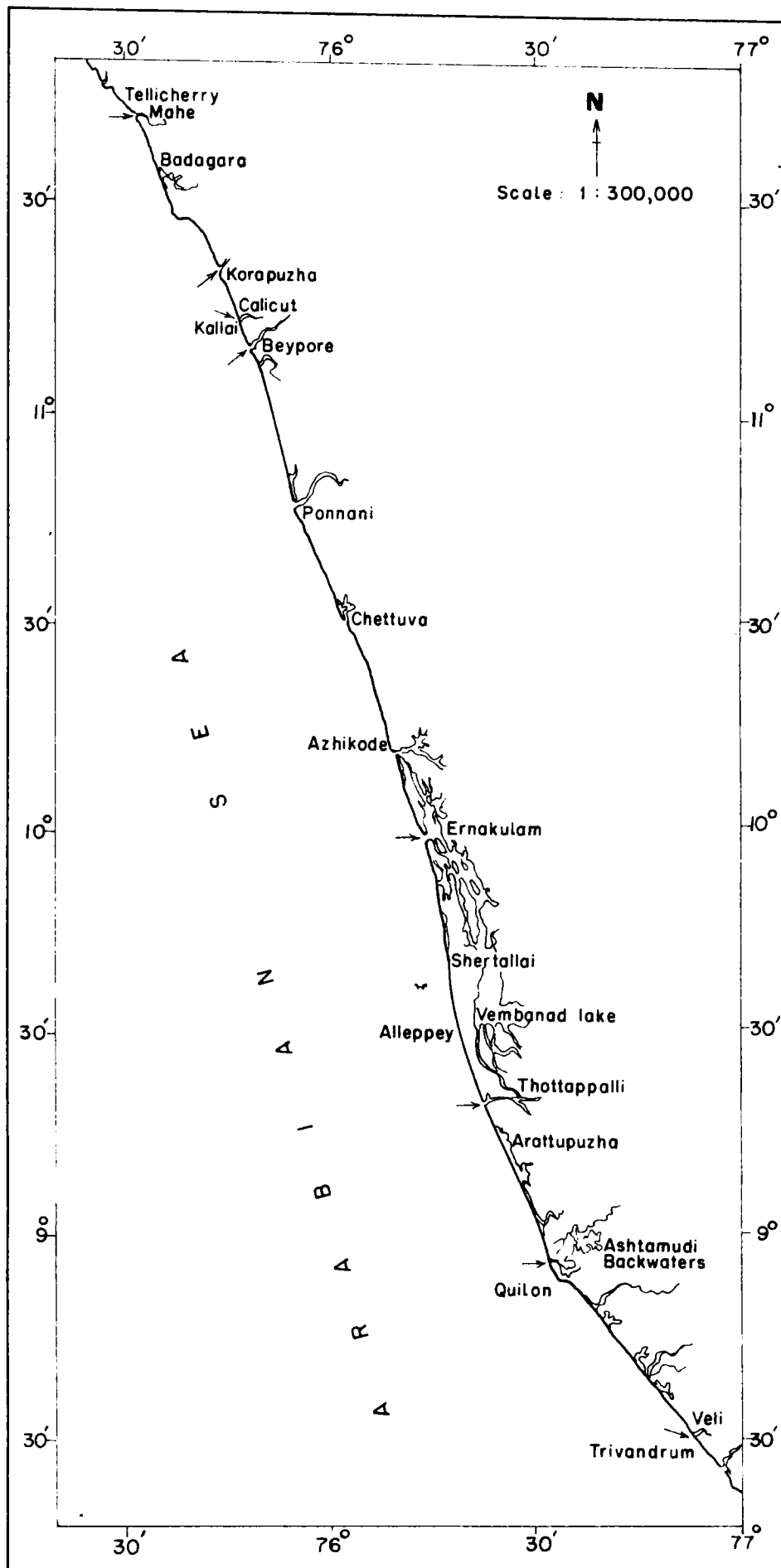


Fig. 2

Fig. 3. Map of Cochis backwaters showing 7 Stations.

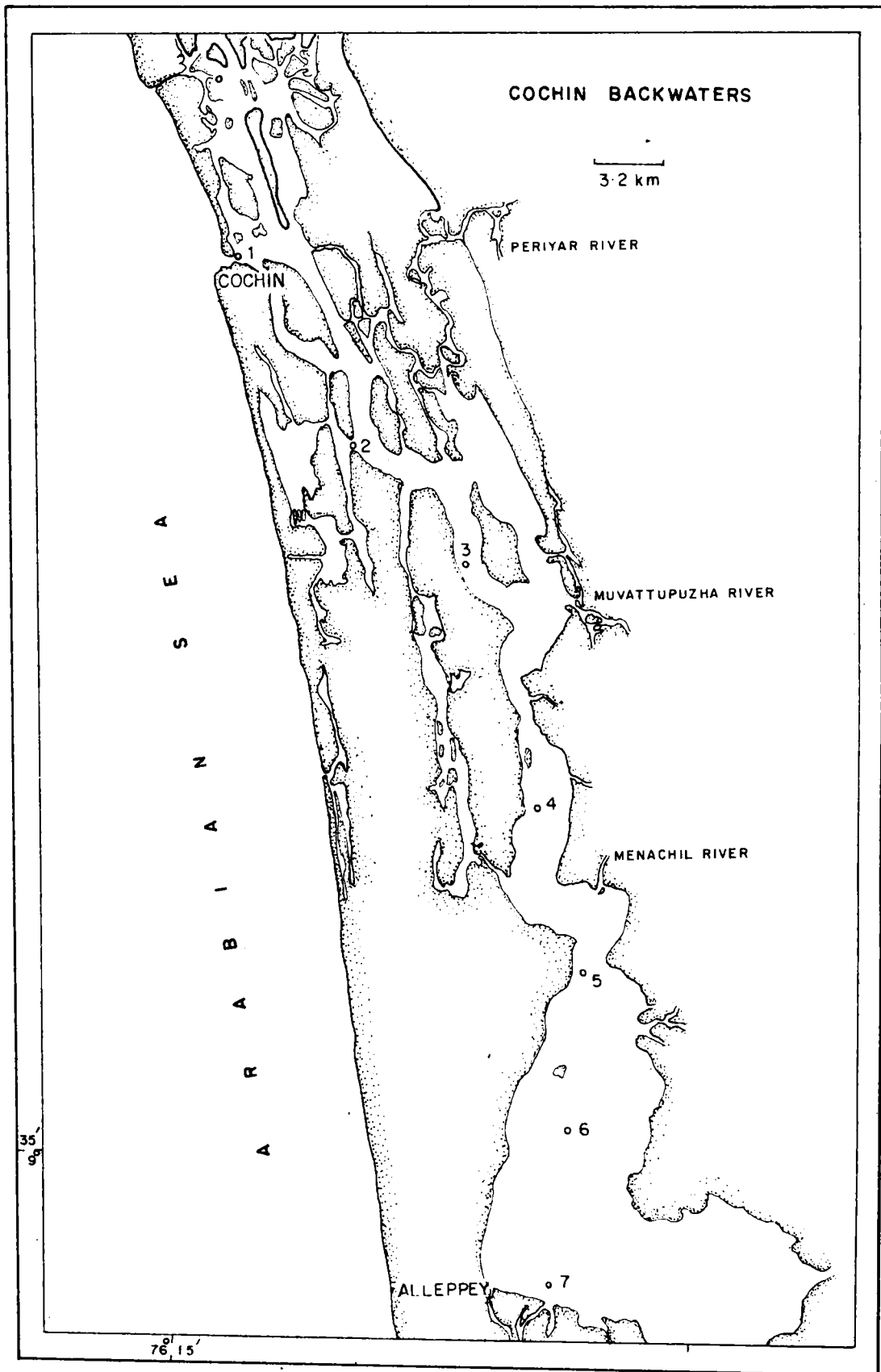


Fig . 3

**Fig. 4 A . Salinity, temperature, oxygen distribution and
rainfall at the mouth of Cochin backwaters
during 1978.**

COCHIN

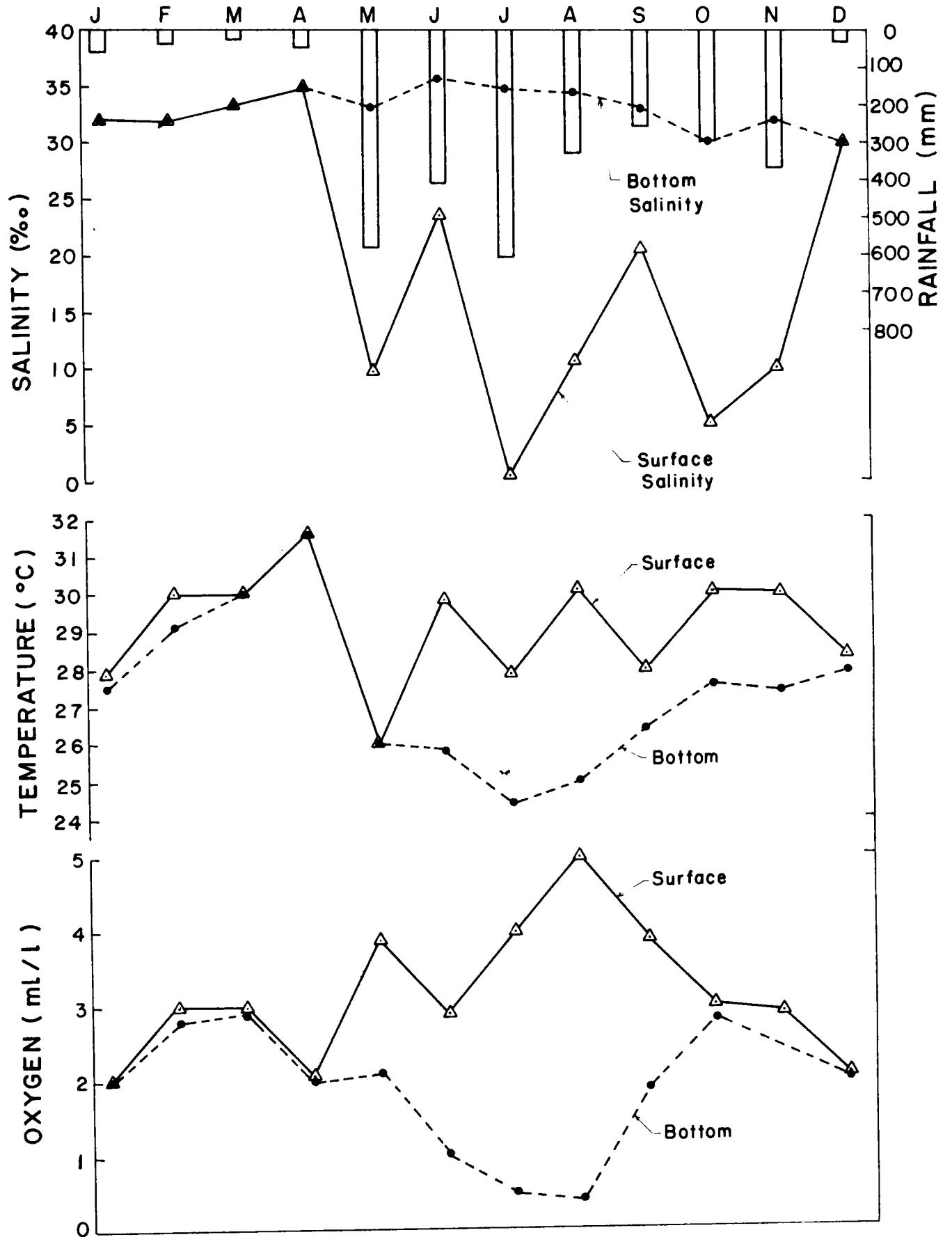


Fig. 4 A

**Fig. 4 B. Salinity distribution to the upper reaches of
Cochin backwaters.**

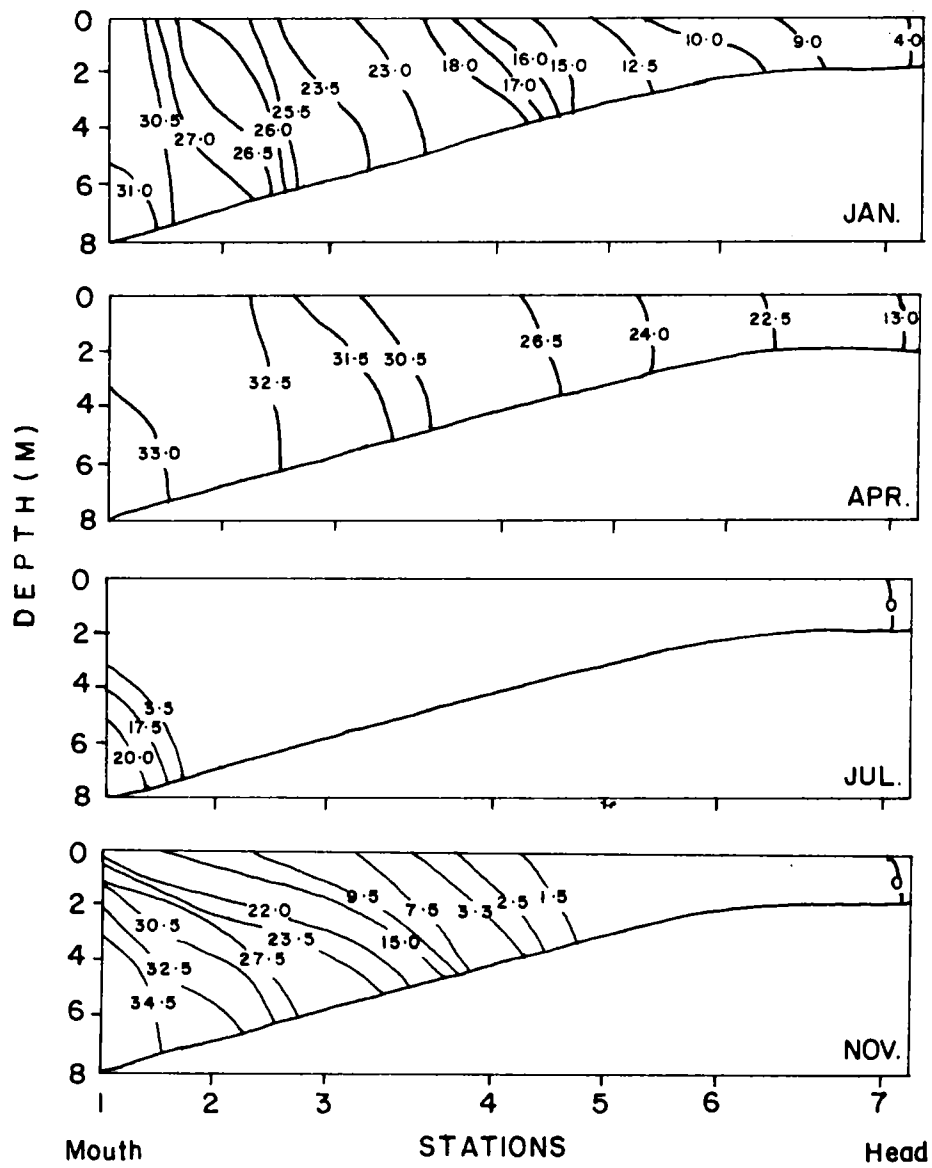


Fig. 4 B

**Fig. 5. Salinity, temperature, oxygen distribution and
rainfall during 1978 at Houndakara.**

Description of lines in Figures 5 to 11 as in Fig. 4 A.

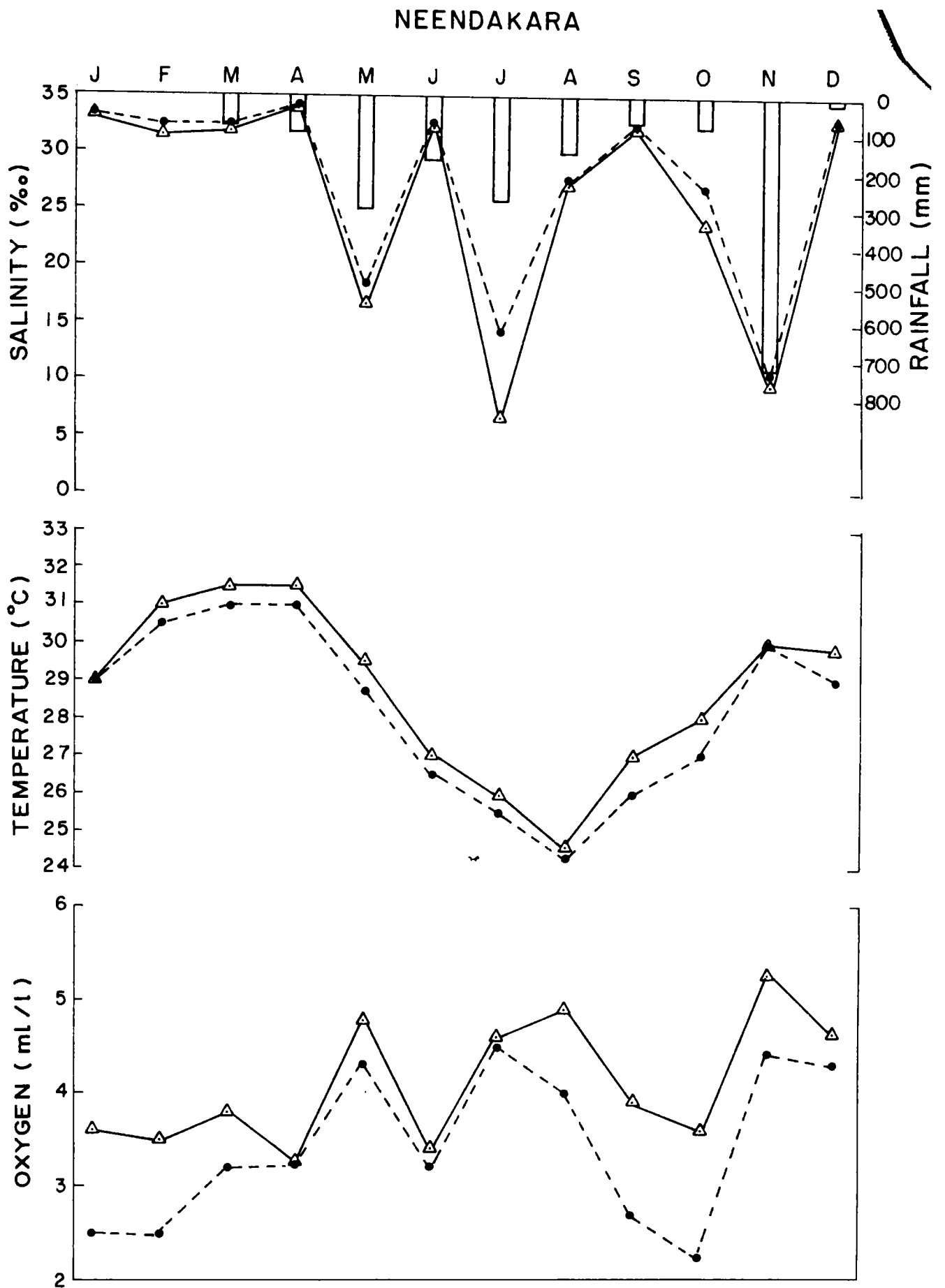


Fig. 5

**Fig. 6 : Salinity, temperature, oxygen distribution
and rainfall during 1978 at Kallai.**

KALLAI

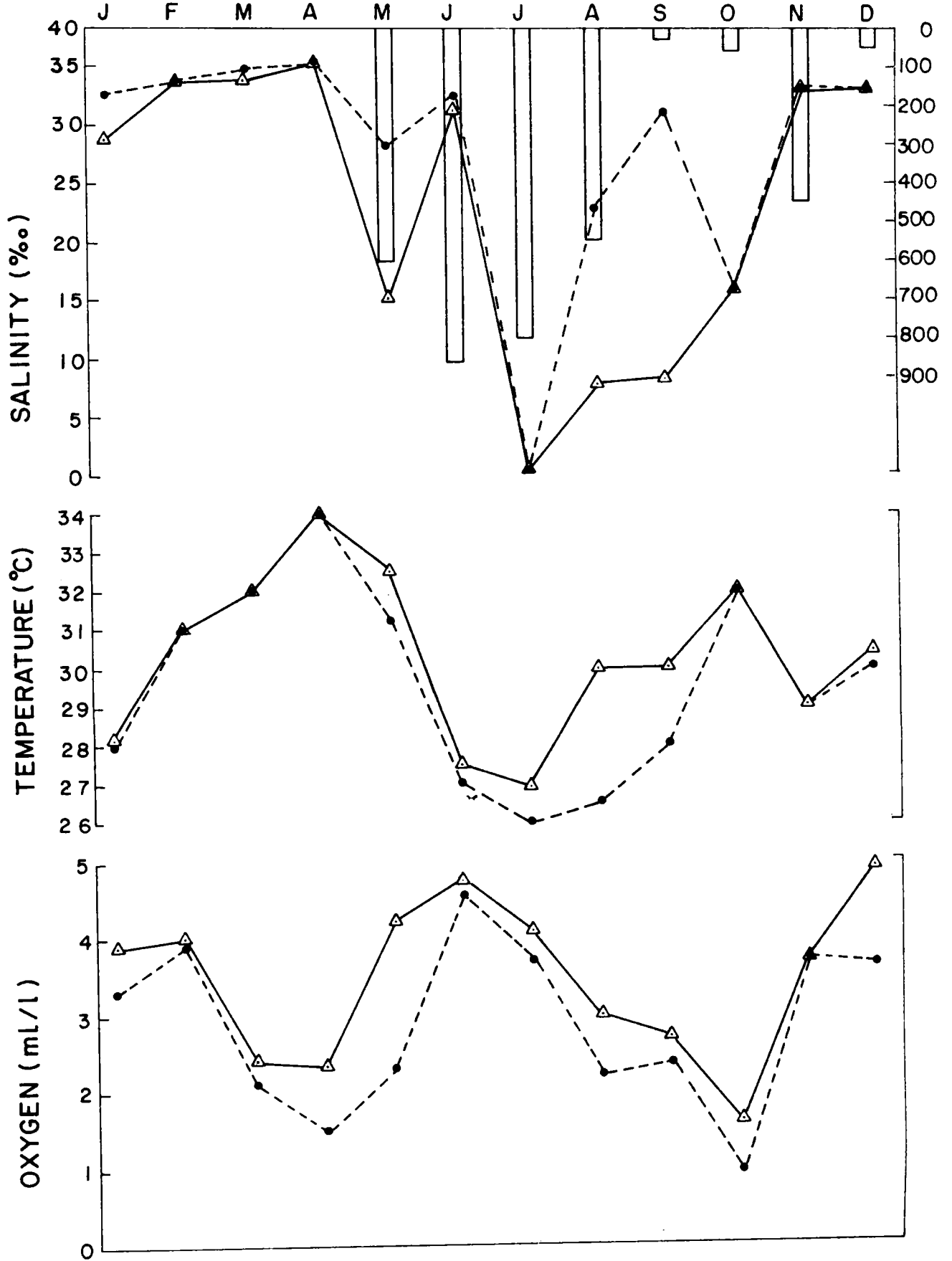


Fig. 6

**Fig. 7 : Salinity, temperature, oxygen distribution
and rainfall during 1978 at Beypore.**

BEYPORE

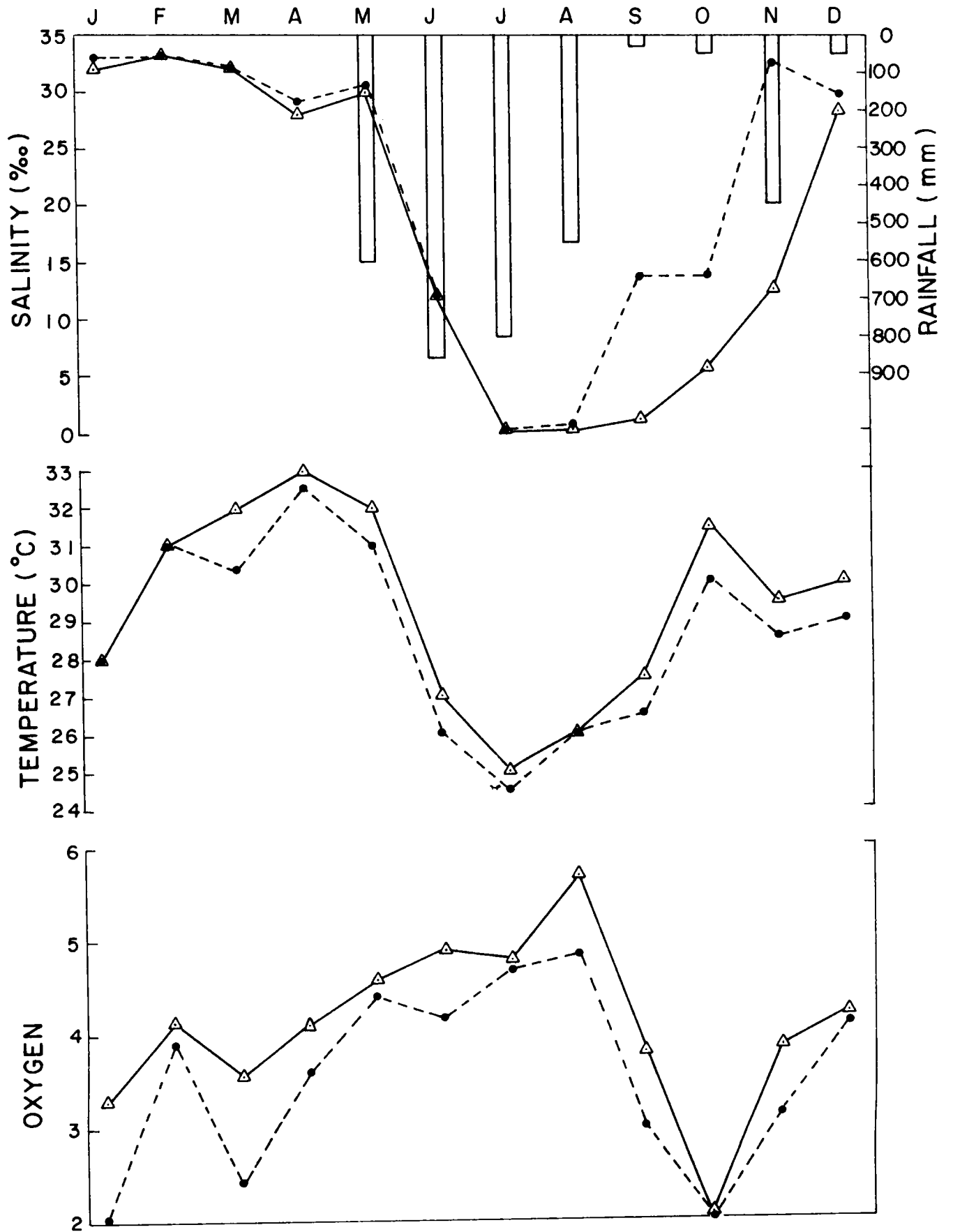


Fig. 7

Fig. 8: Salinity, temperature, oxygen distribution and rainfall during 1978 at Koraputa.

KORAPUZHA

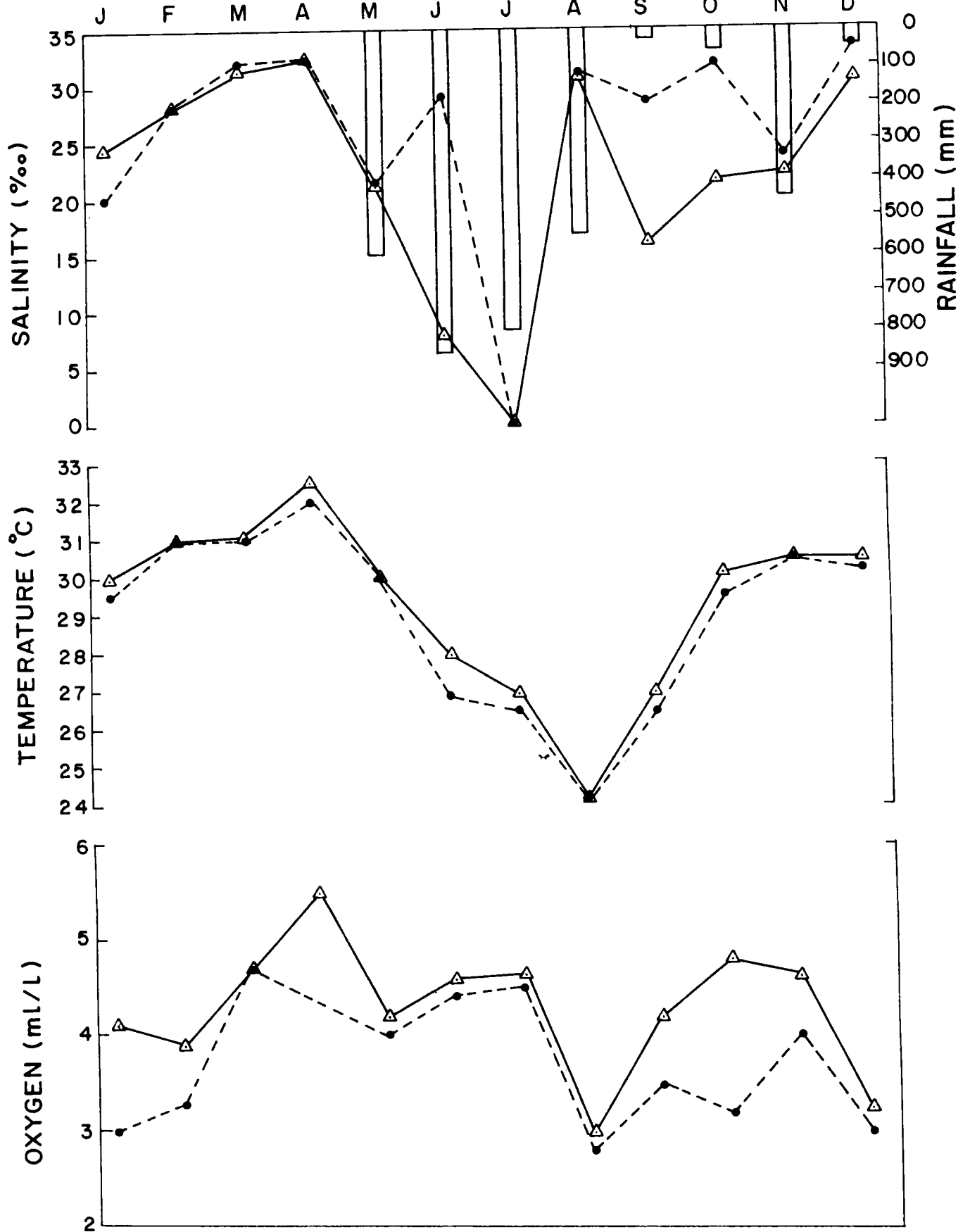


Fig. 8

Fig. 9. Salinity, temperature, oxygen distribution and rainfall during 1970 at Nabe.

MAHE

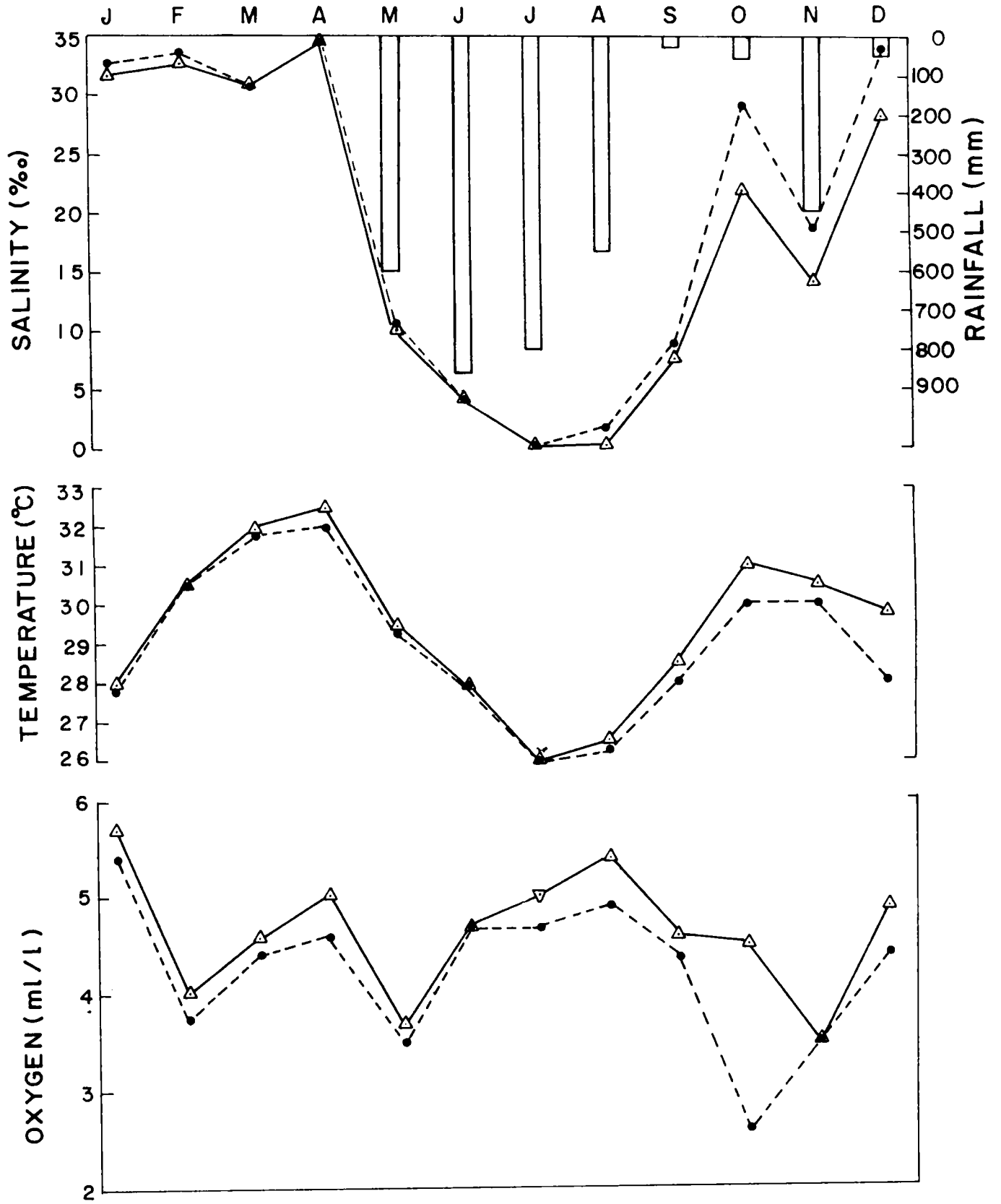


Fig. 9

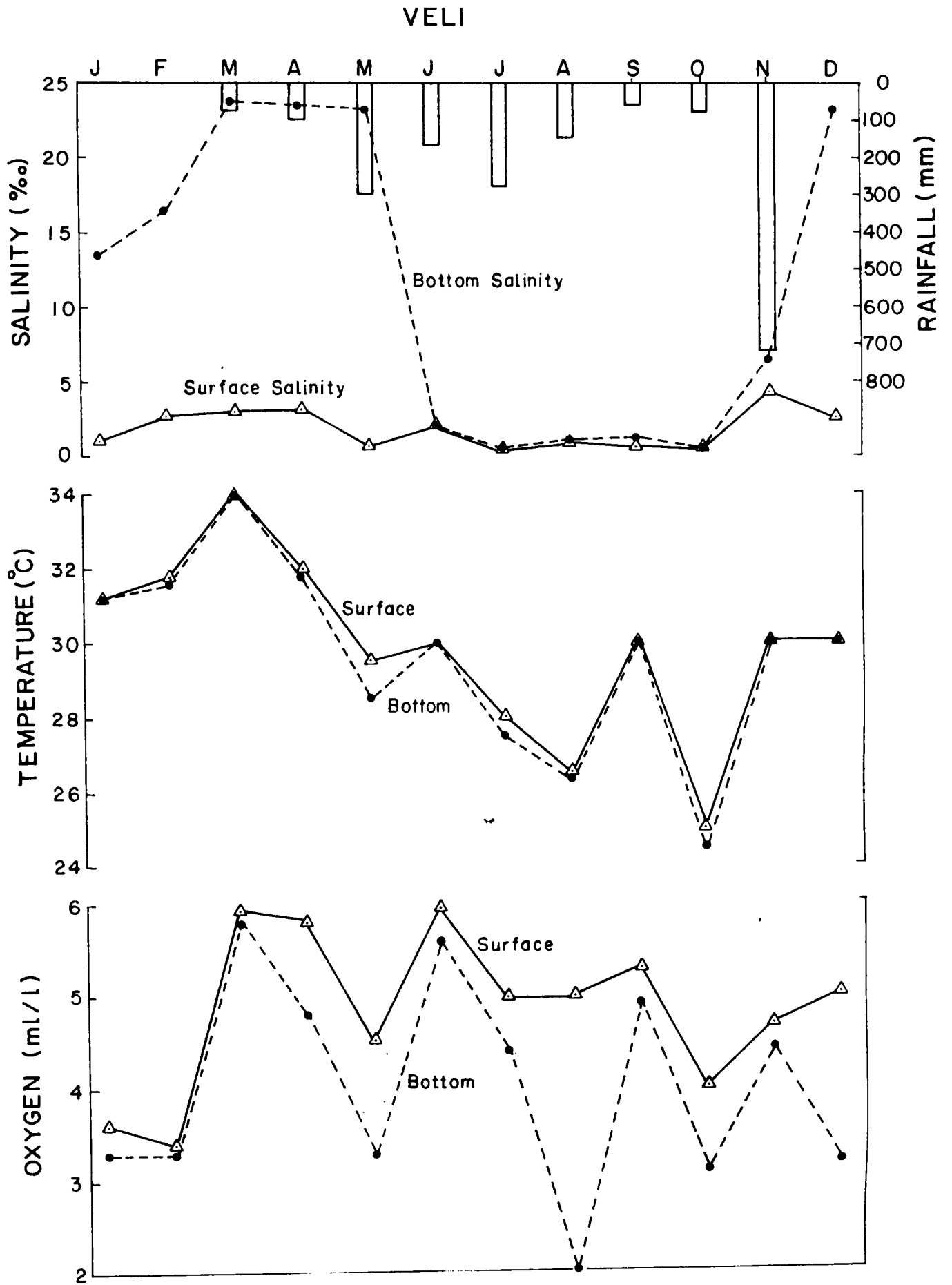


Fig. 10

Fig. 11. Salinity, temperature, oxygen distribution and rainfall during 1978 at Thottappilly.

THOTTAPPILLY

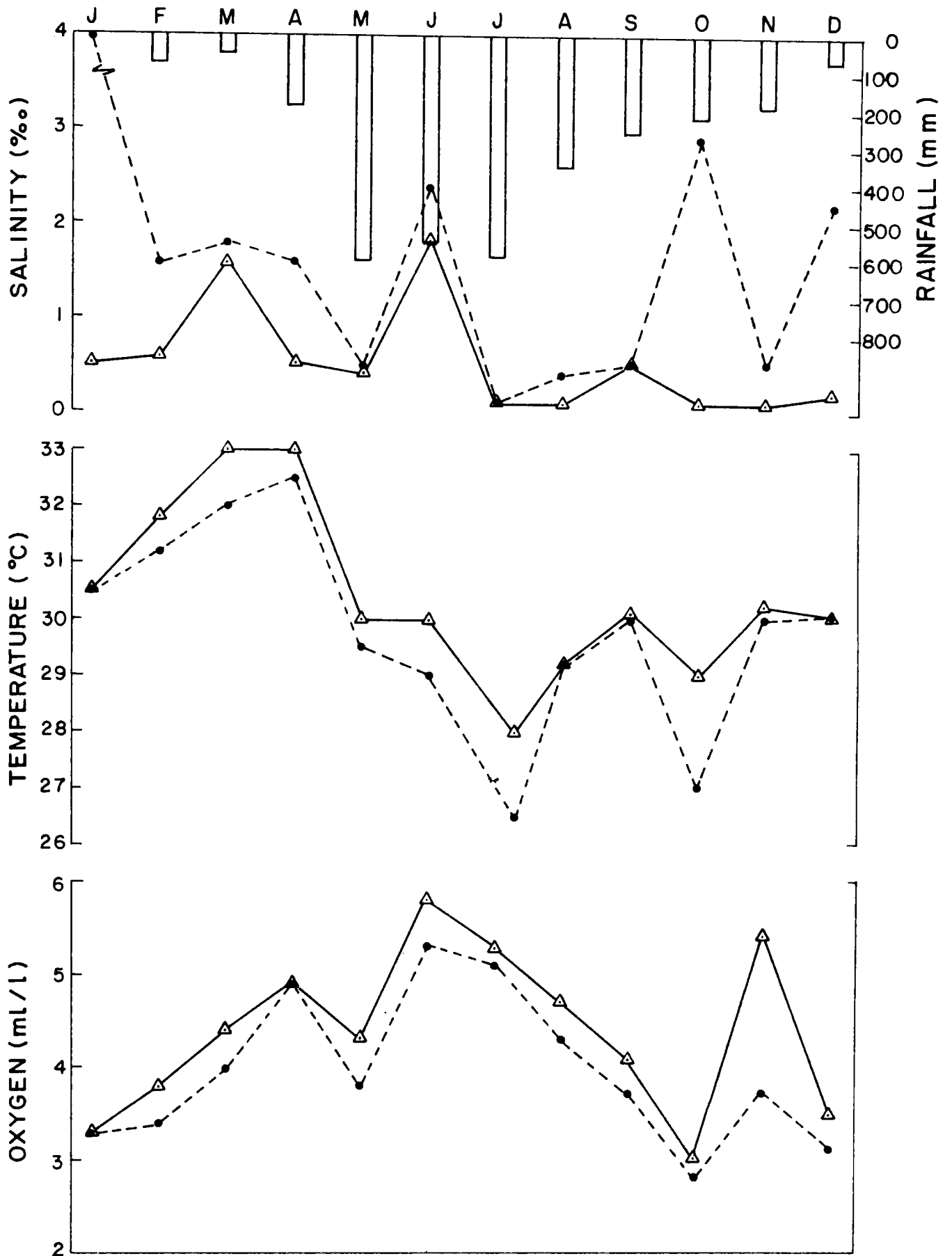


Fig. 11

**Fig. 12. Distribution of zooplankton counts (No./10m³)
in the estuaries during 1978.**

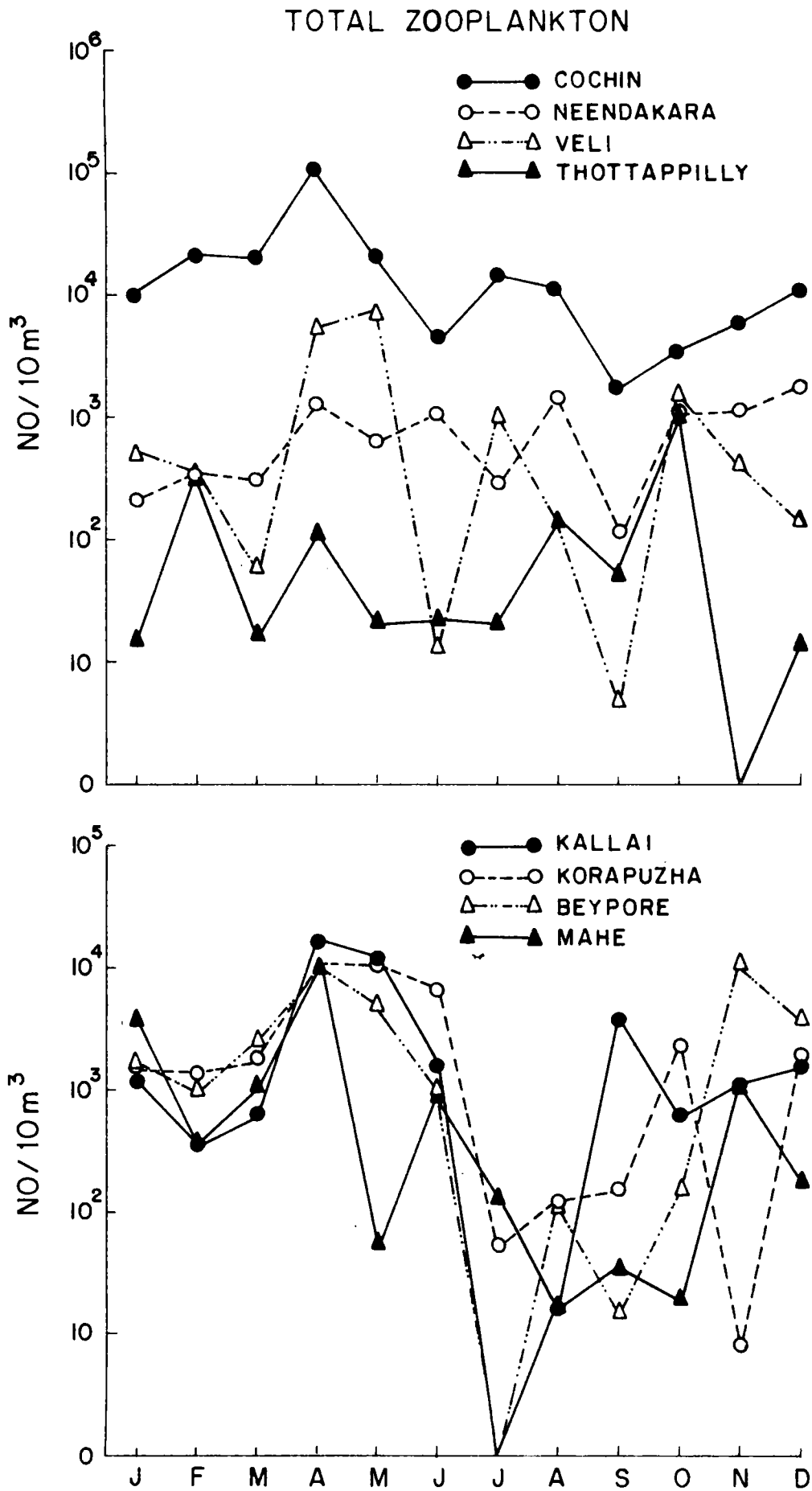


Fig. 12

**Fig. 13. Distribution of common groups at Cochin and
Kandakara estuaries.**

**Description of lines in the bottom figures of
Figs. 13 to 16 as in the upper figures.**

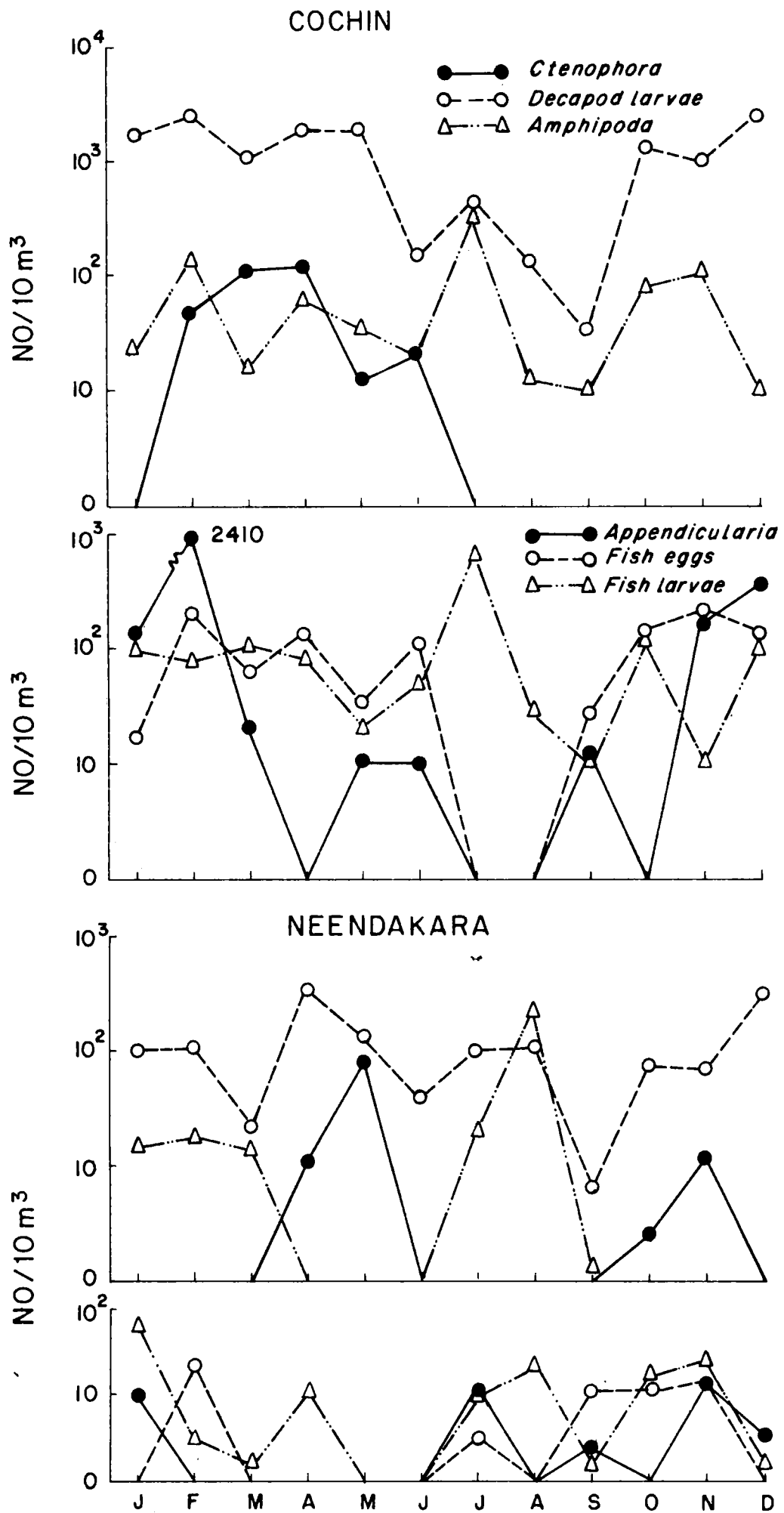


Fig. 13

Fig. 14. Distribution of common groups at Kallai and Kerepuzha estuaries.

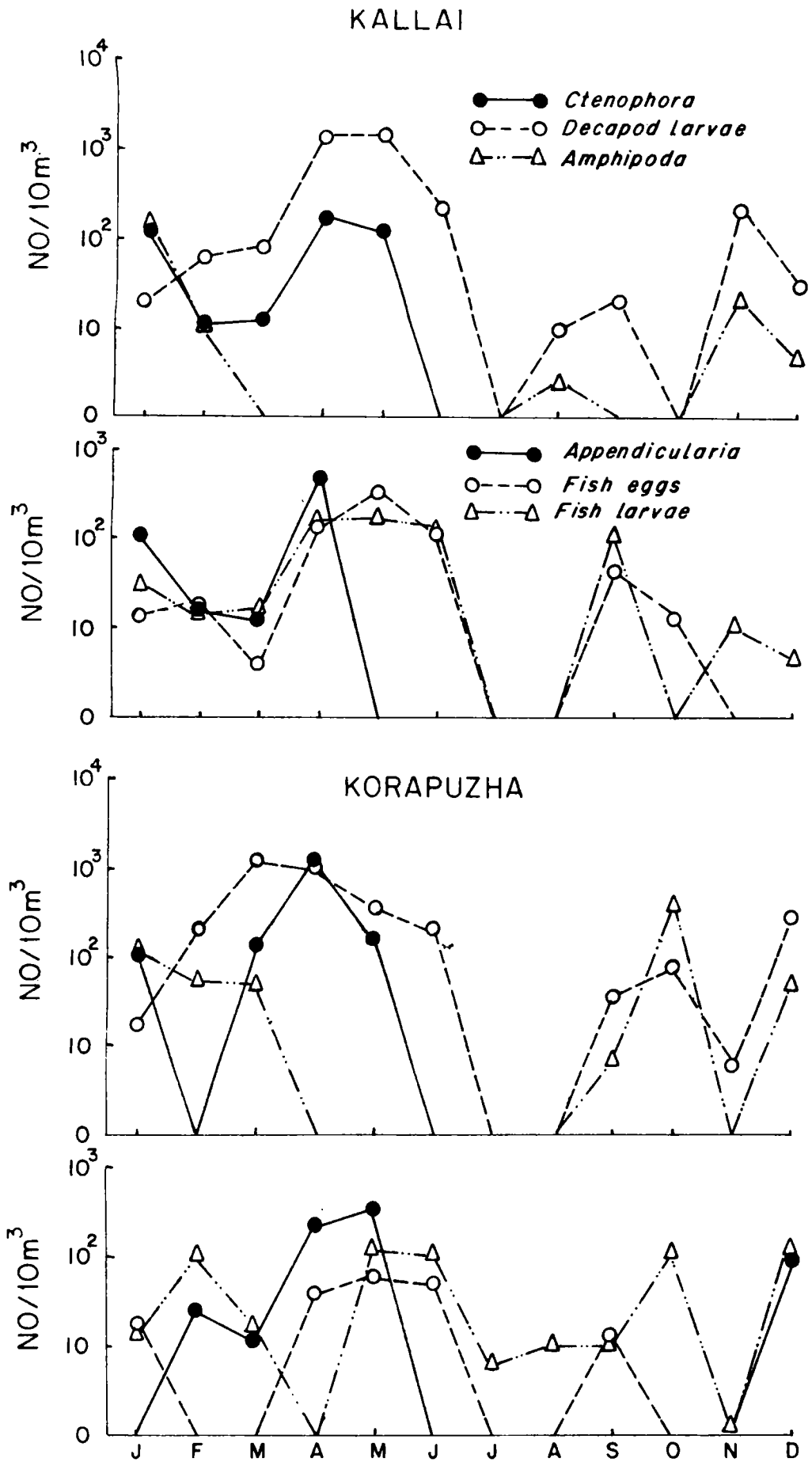


Fig. 14

**Fig. 15. Distribution of common groups at Beypore and
Maha estuaries.**

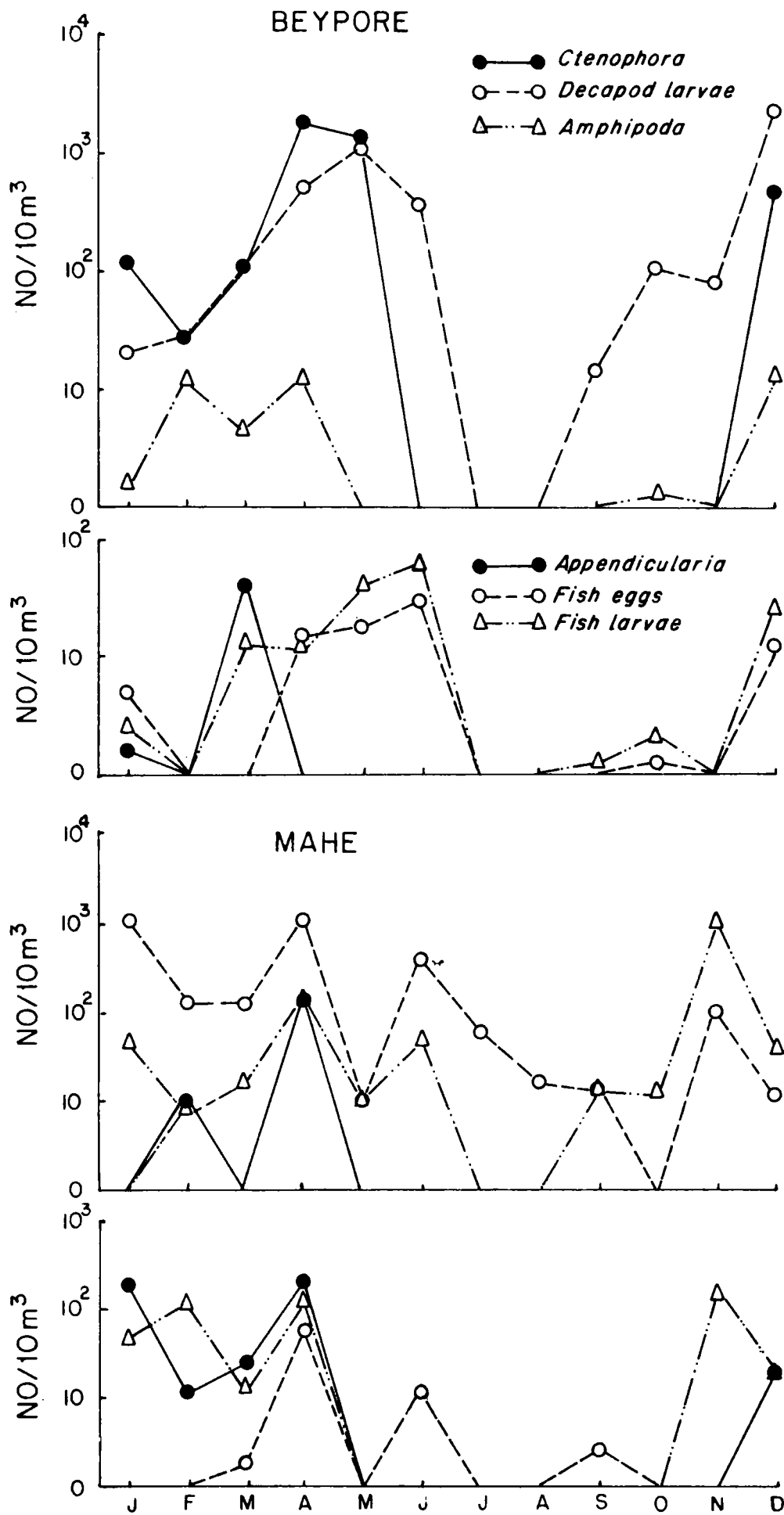


Fig. 15

Fig. 16. Distribution of common groups at Veli and Thottappilly lakes.

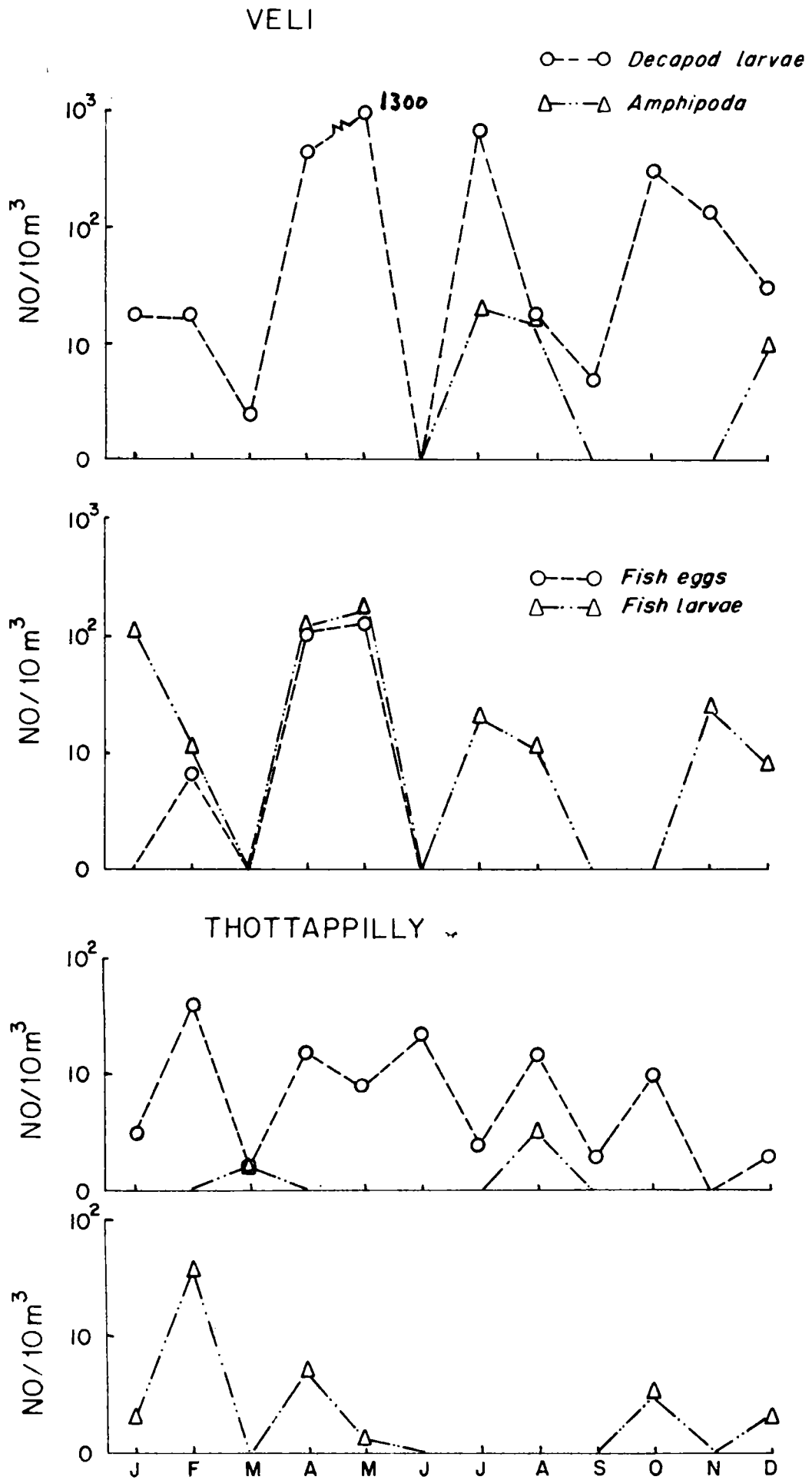


Fig. 16

Fig. 17. Distribution of common groups towards the interior of Cechin backwaters.

COCHIN

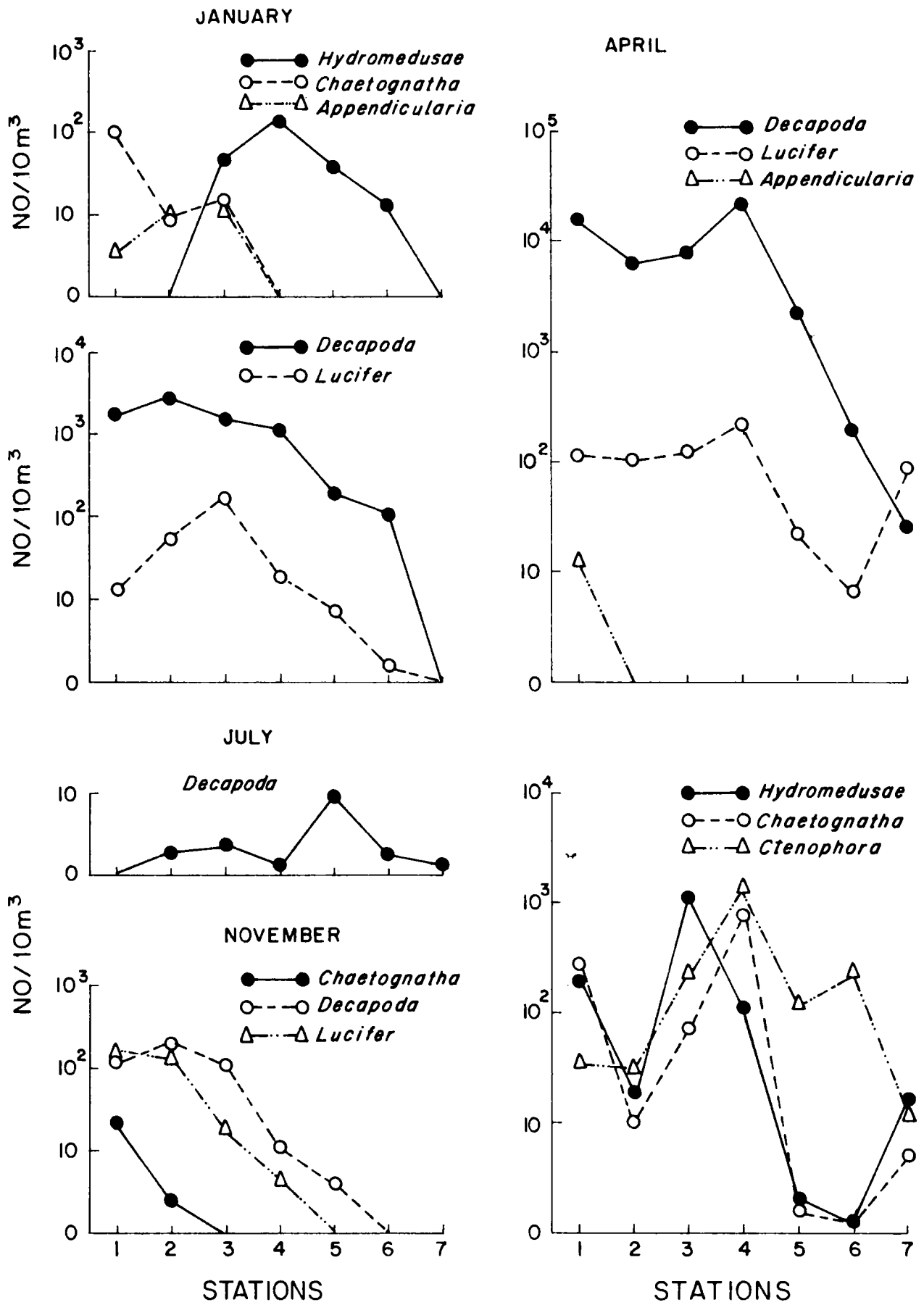


Fig. 17

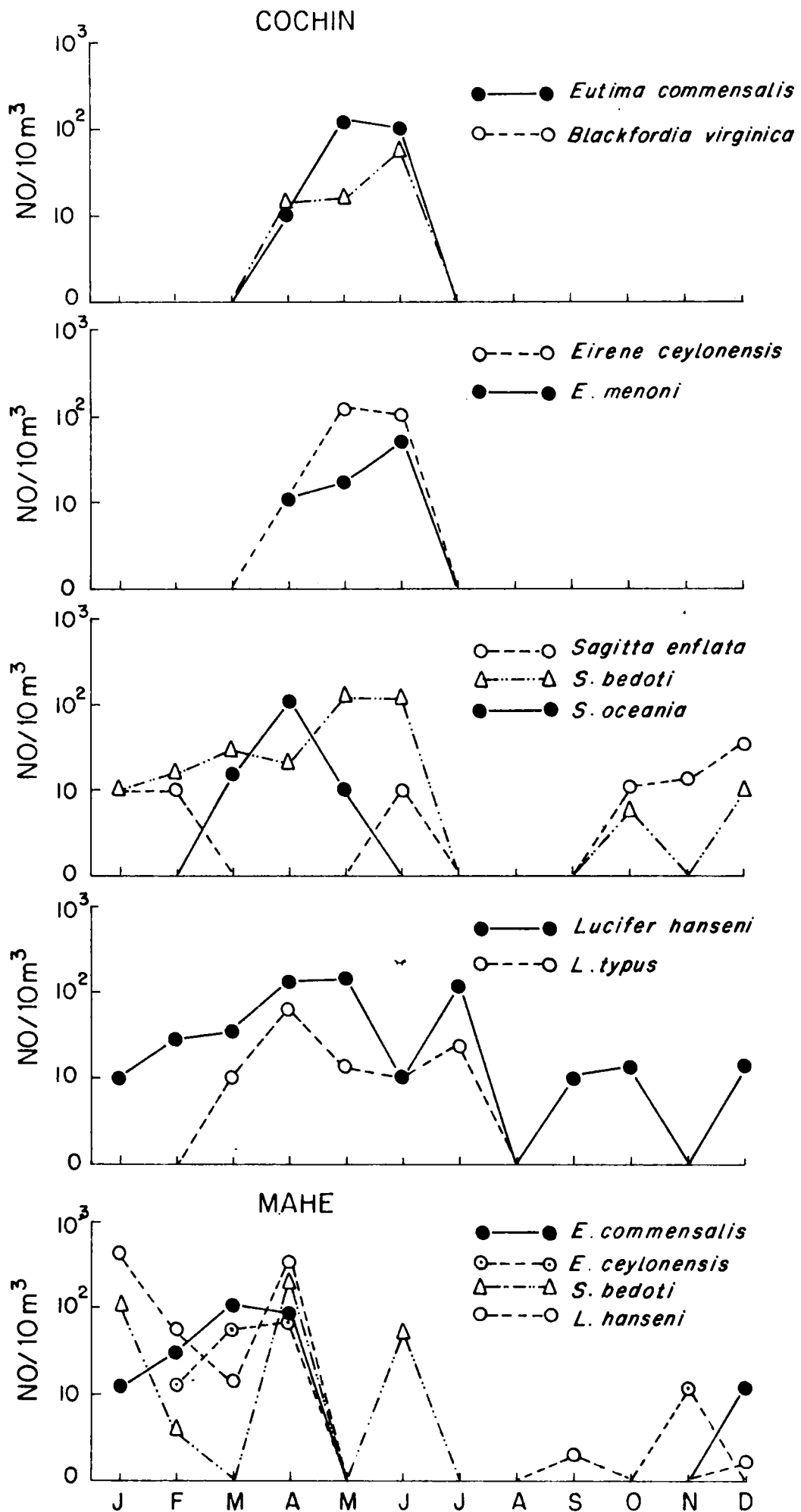


Fig. 18

**Fig. 19. Distribution of common species other than copepods
at Karapuzha and Kallai estuaries.**

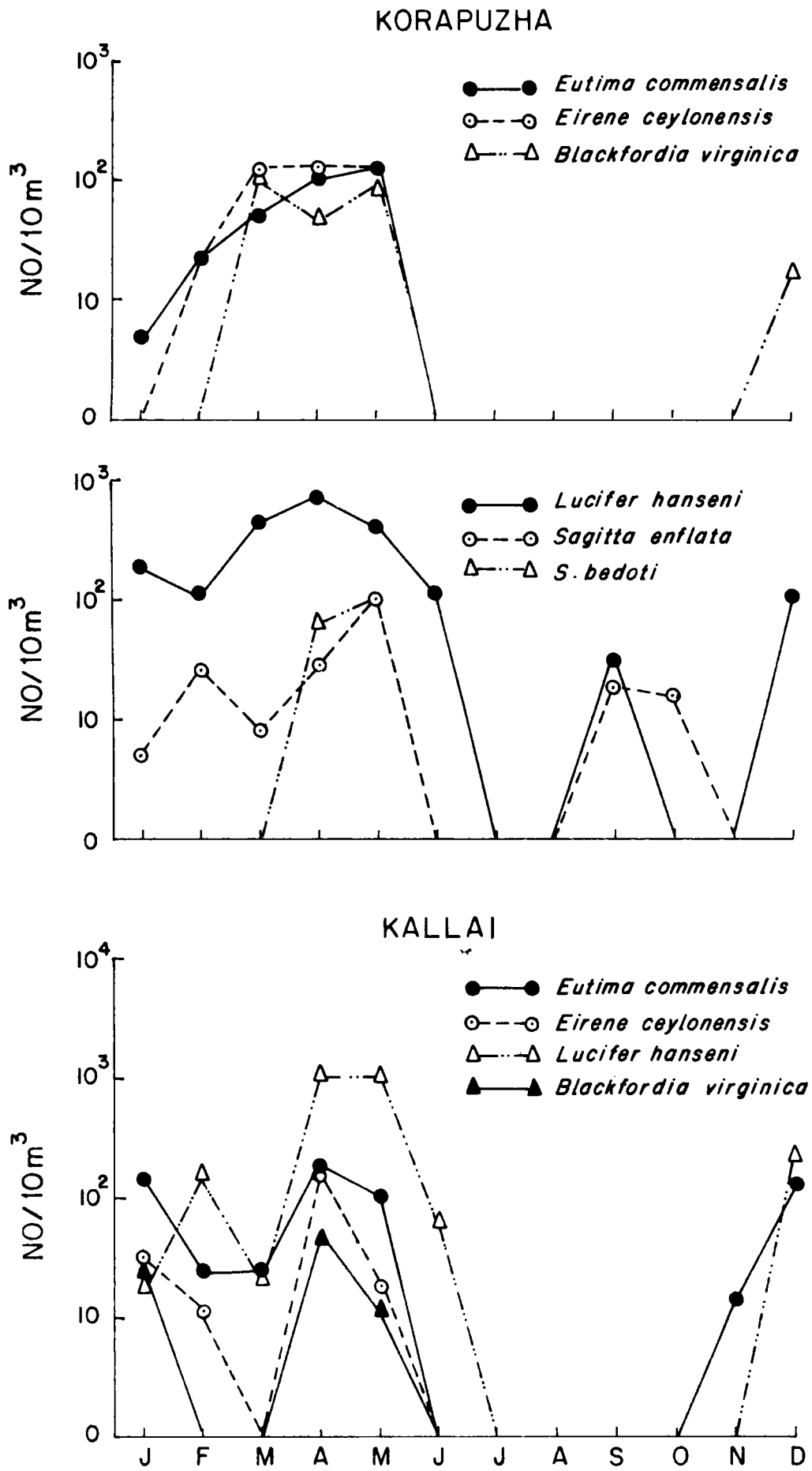


Fig. 19

**Fig. 20. Distribution of common species other than copepods
at Baypare and Neendakara estuaries.**

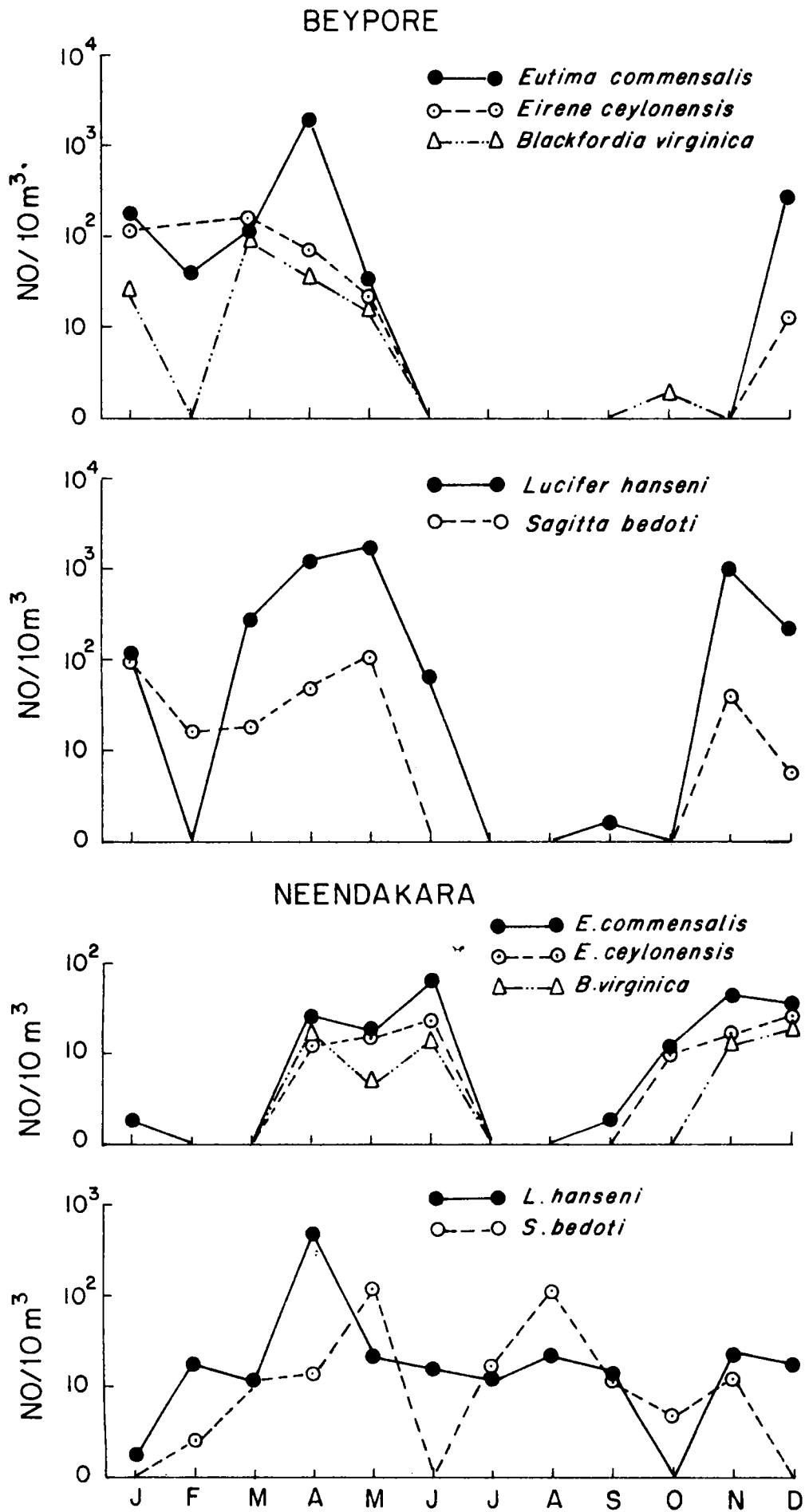


Fig. 20

Fig. 21. Distribution of total Copepoda in the eight estuaries during 1978.

TOTAL COPEPODA

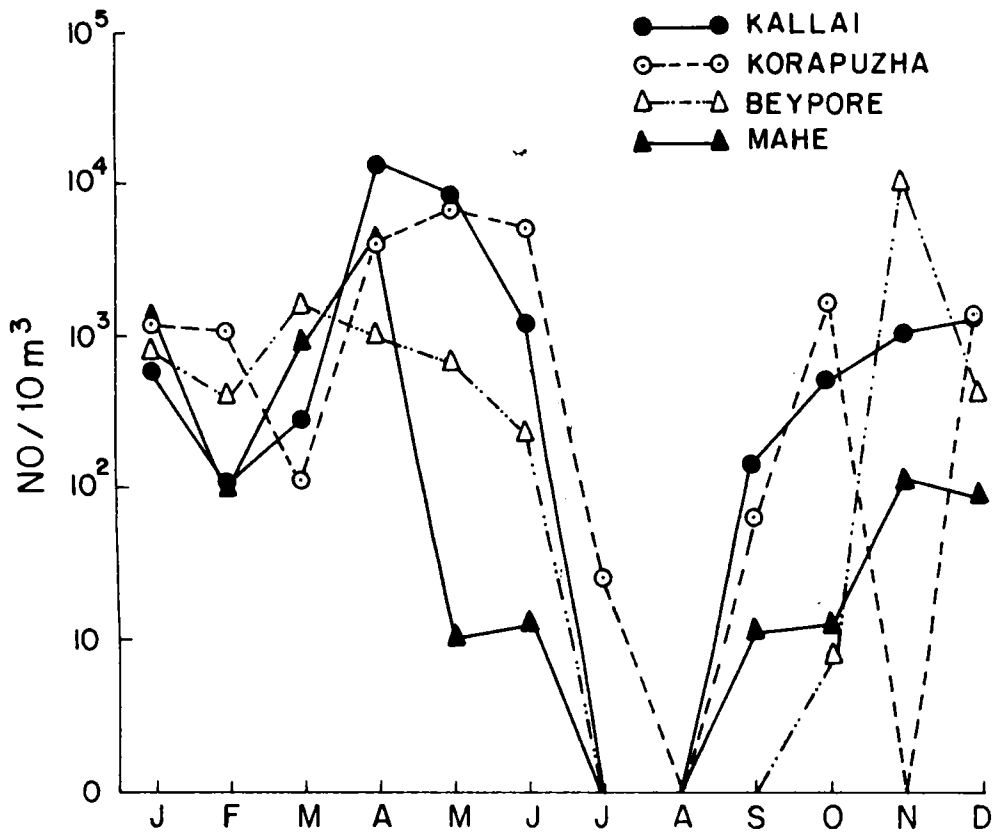
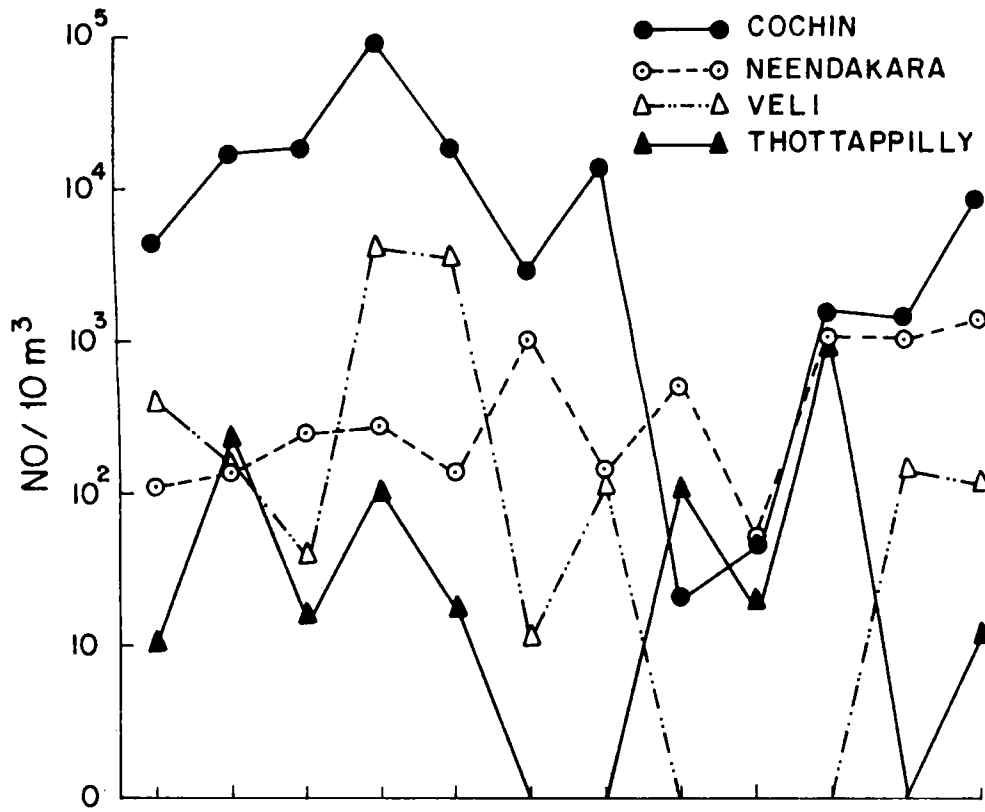


Fig. 21

Fig. 234. Distribution of common cepopod species at the mouth of Cochin backwaters.

COCHIN

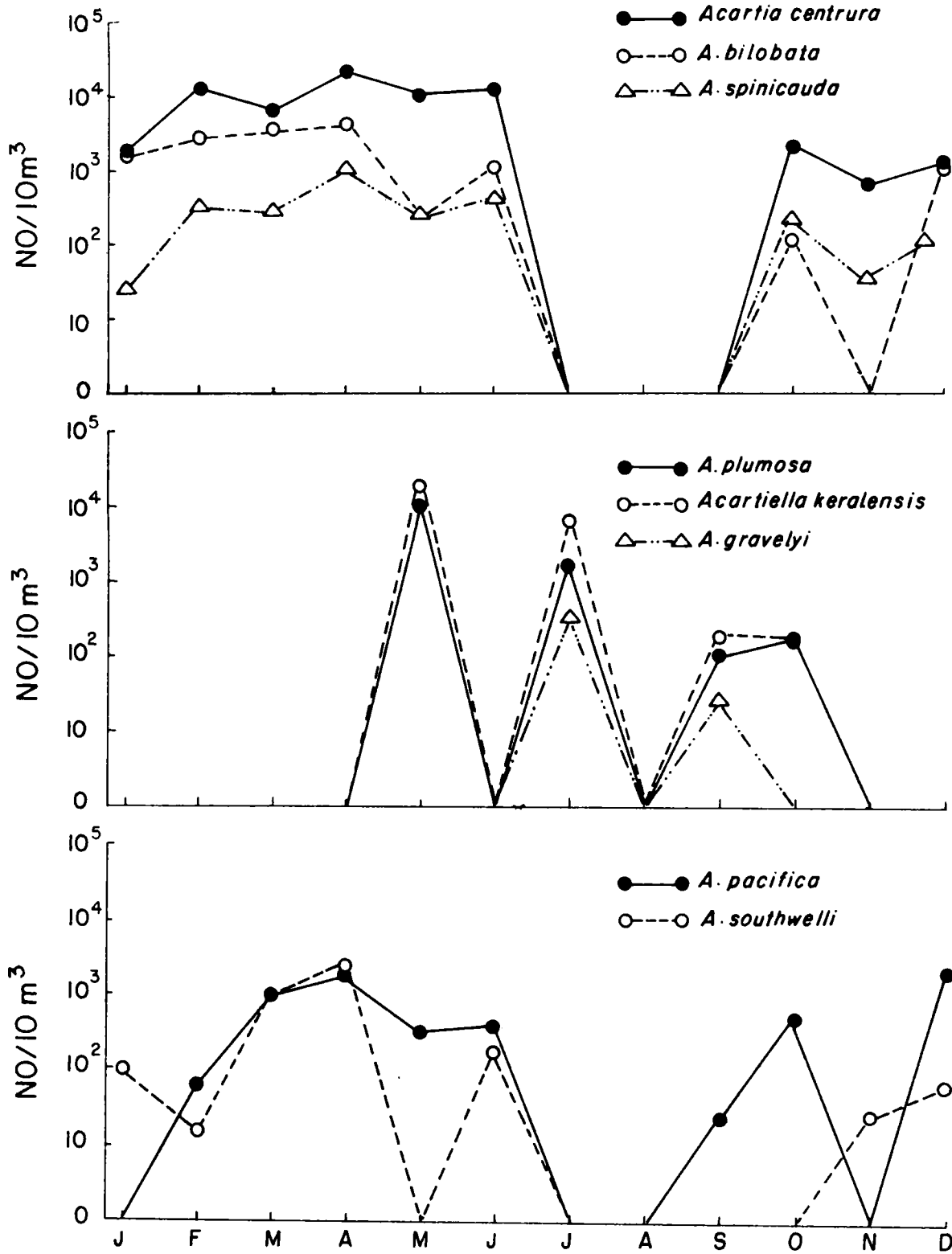


Fig. 22 A

COCHIN

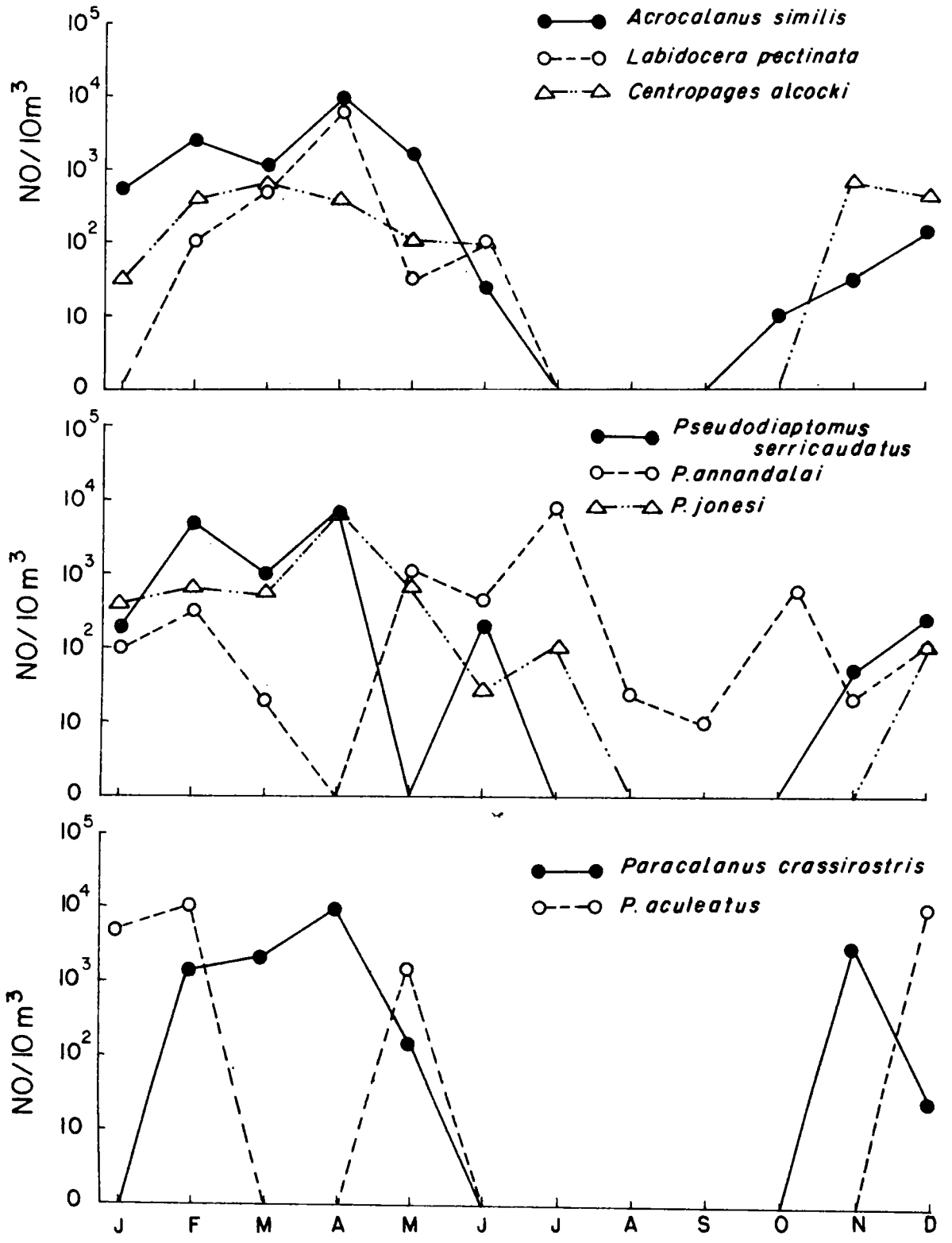


Fig. 22 A

Fig. 22 B. Distribution of common caged species towards the upper reaches of Cochin backwaters.

COCHIN

APRIL

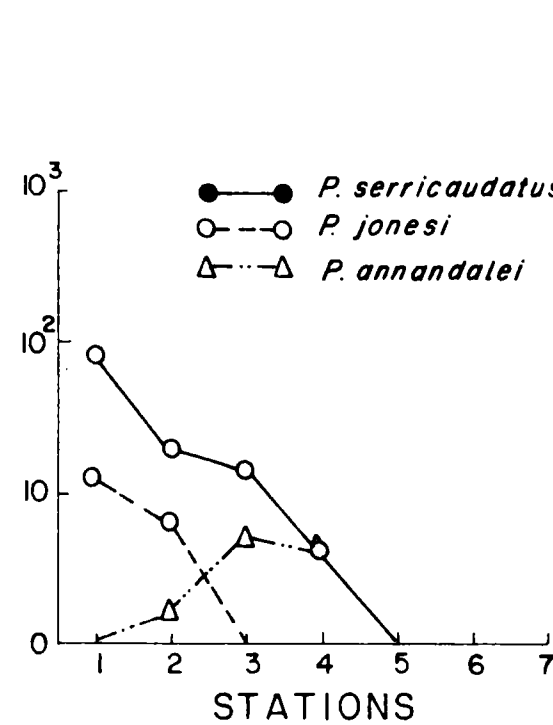
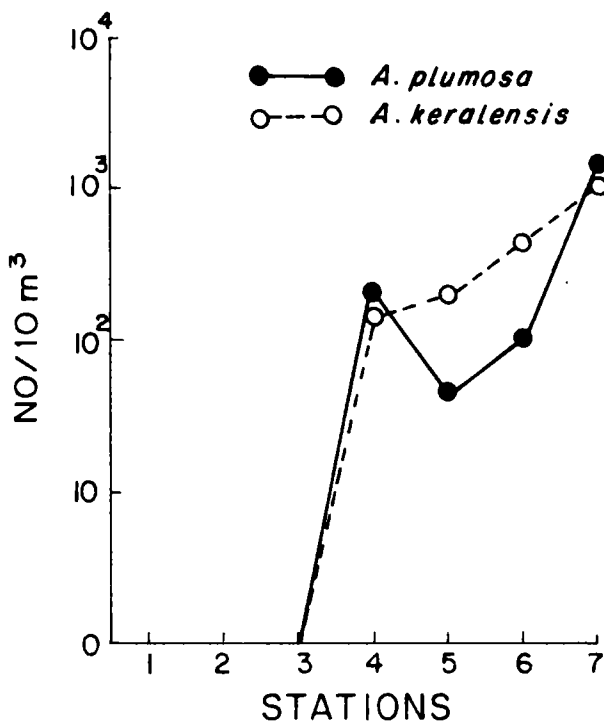
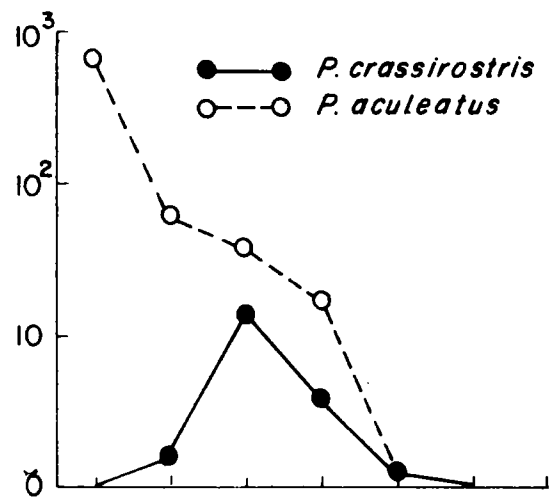
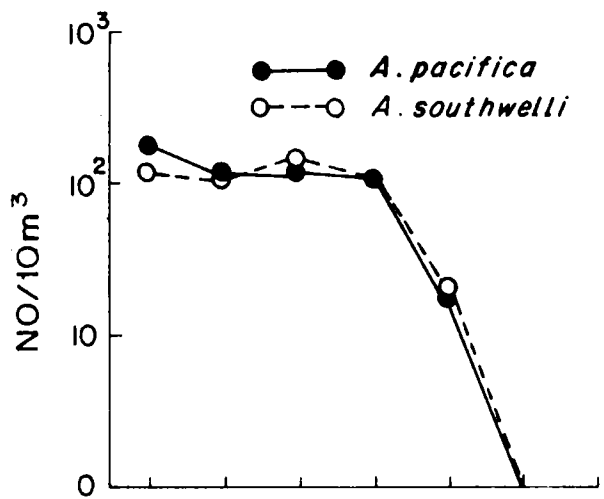
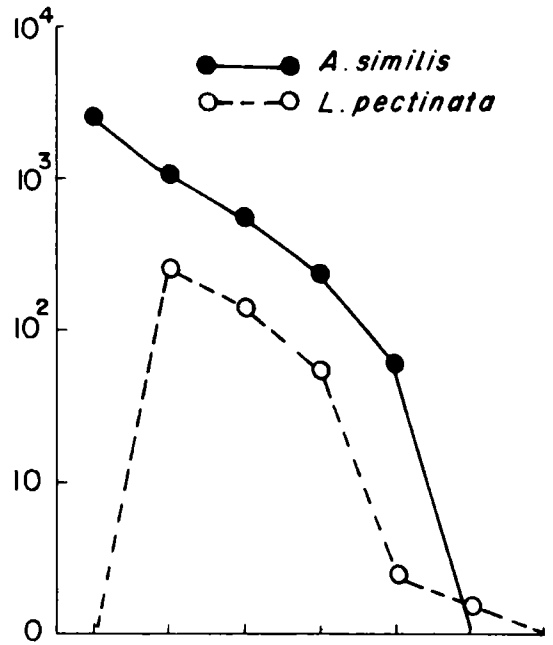
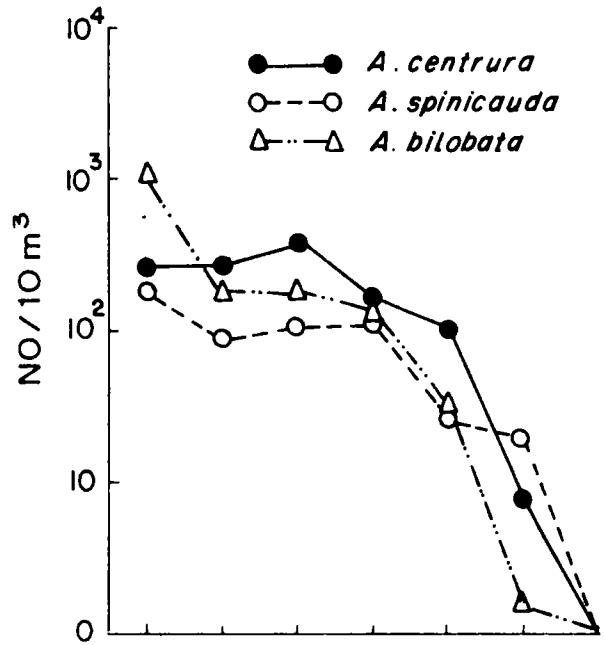


Fig. 22 D

Fig. 22 C. Distribution of common copepod species towards the upper reaches of Cechia backwaters.

COCHIN

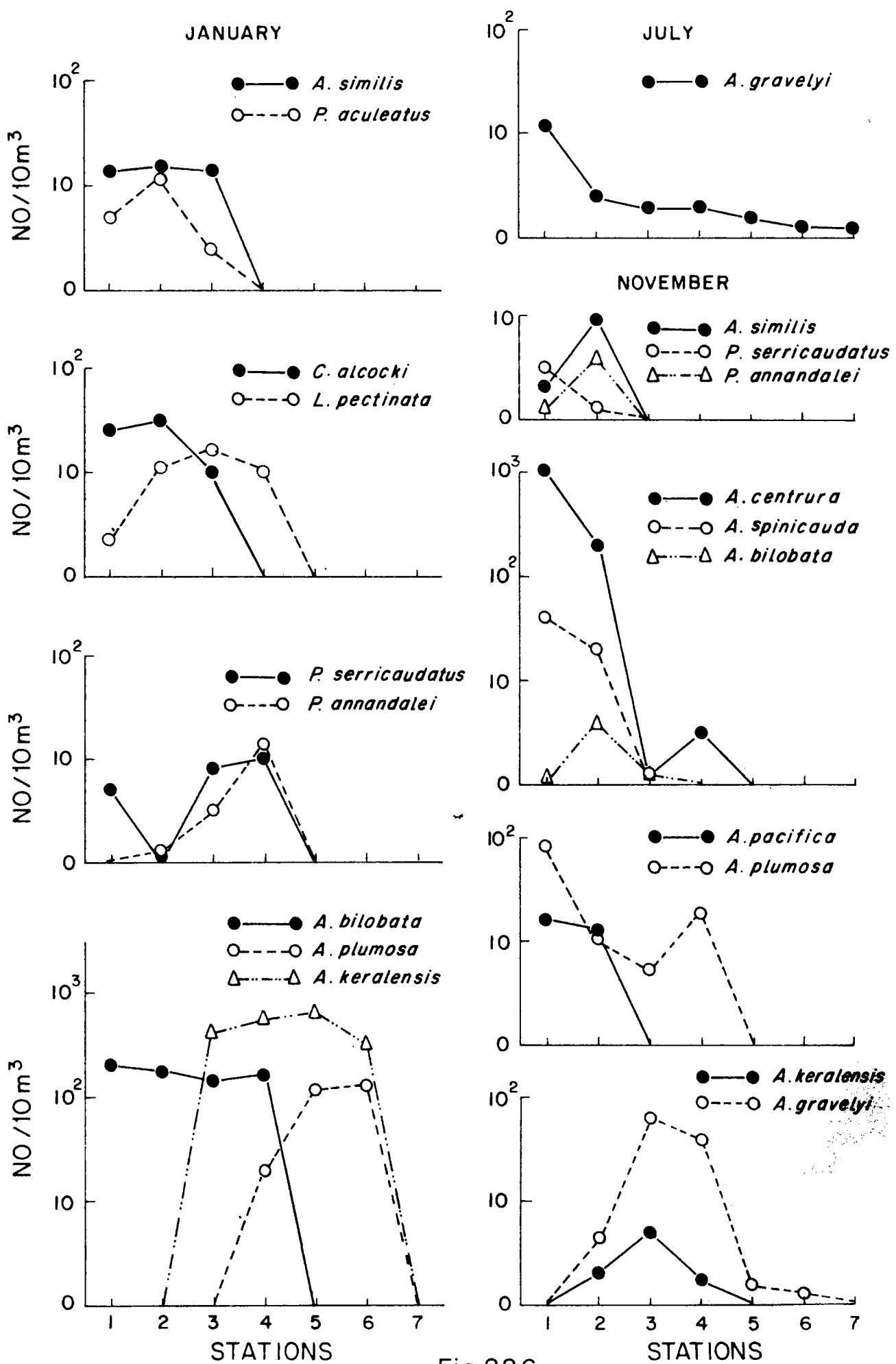


Fig 220

**Fig. 23. Distribution of common copepod species
at Nondakara.**

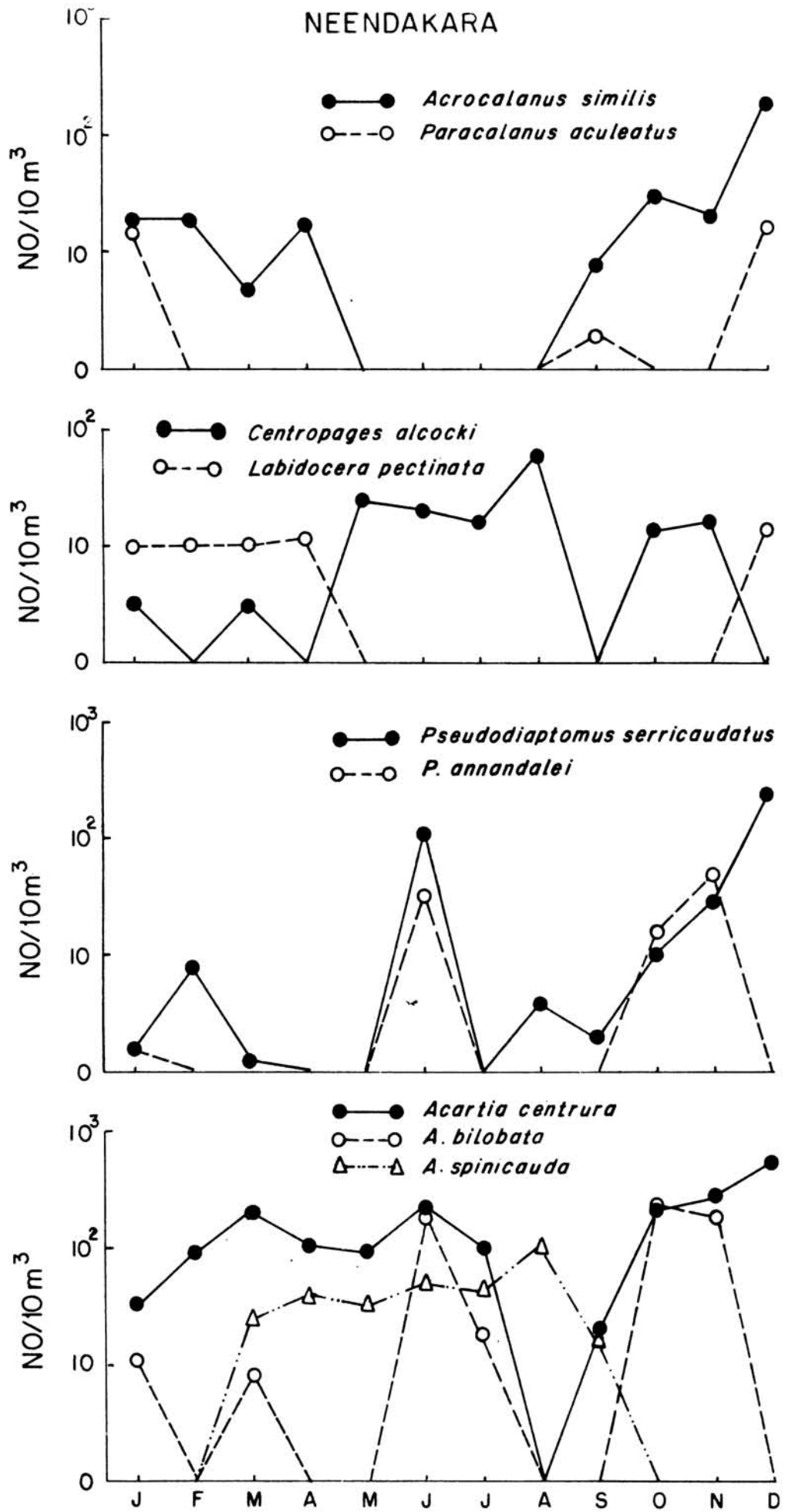
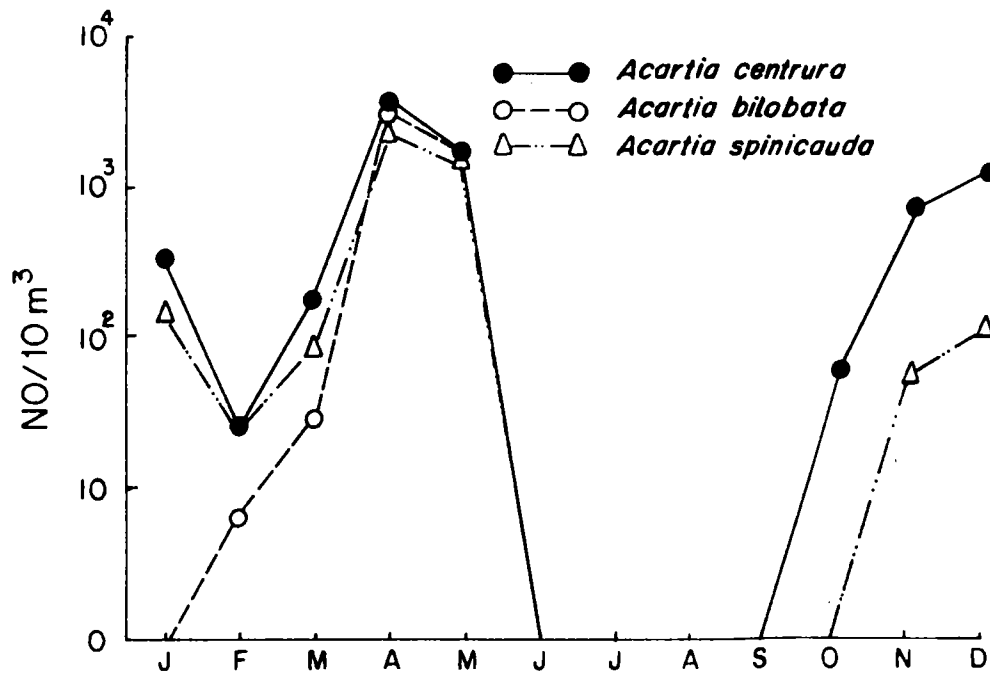
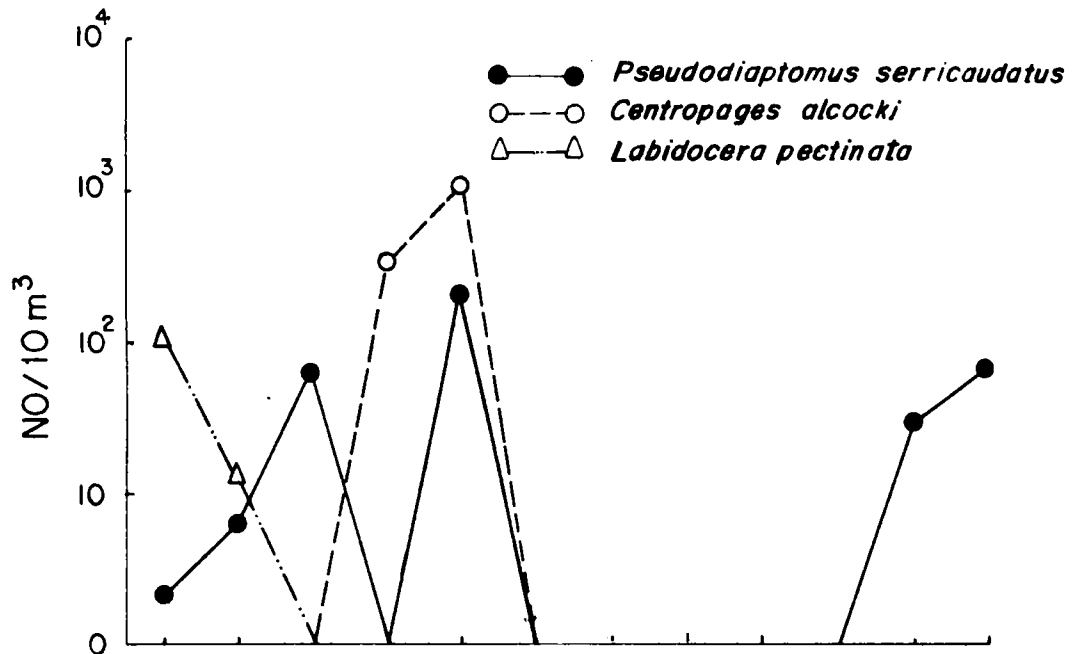
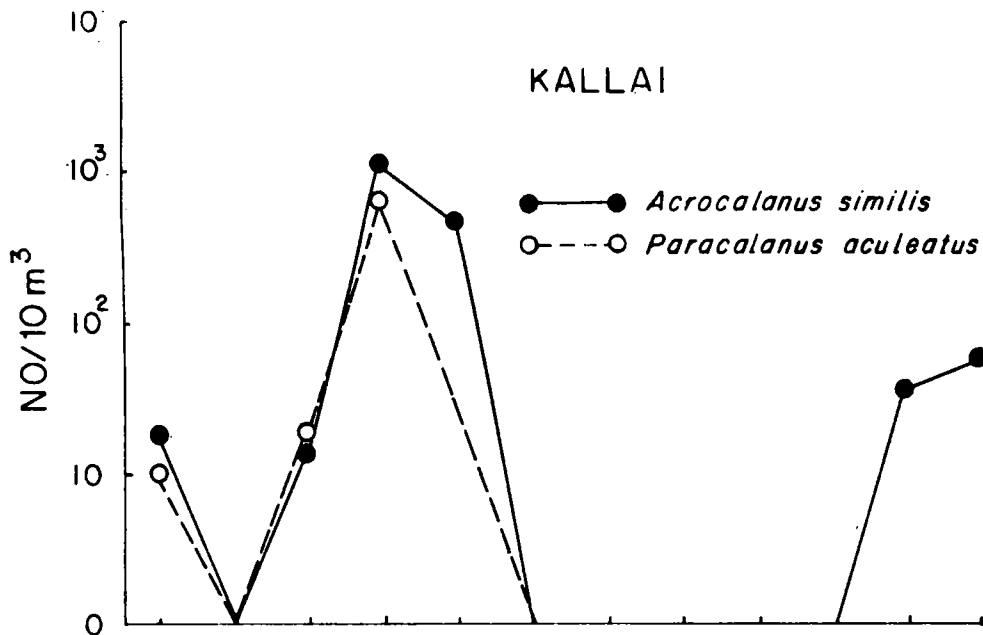


Fig. 23

**Fig. 24. Distribution of common copepod species
at Malindi.**



BEYPORE

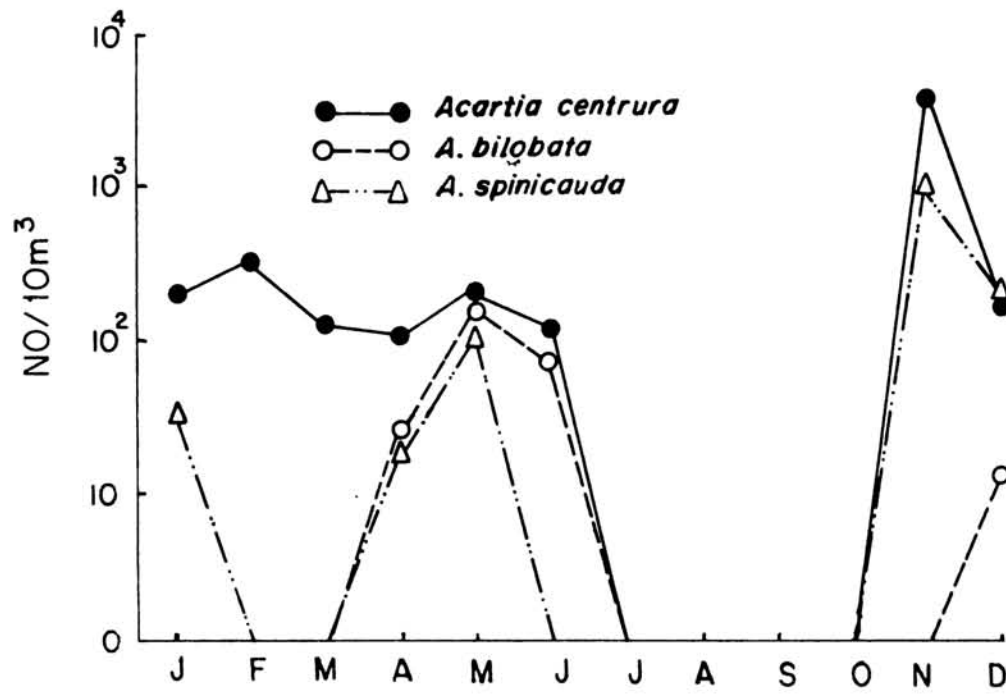
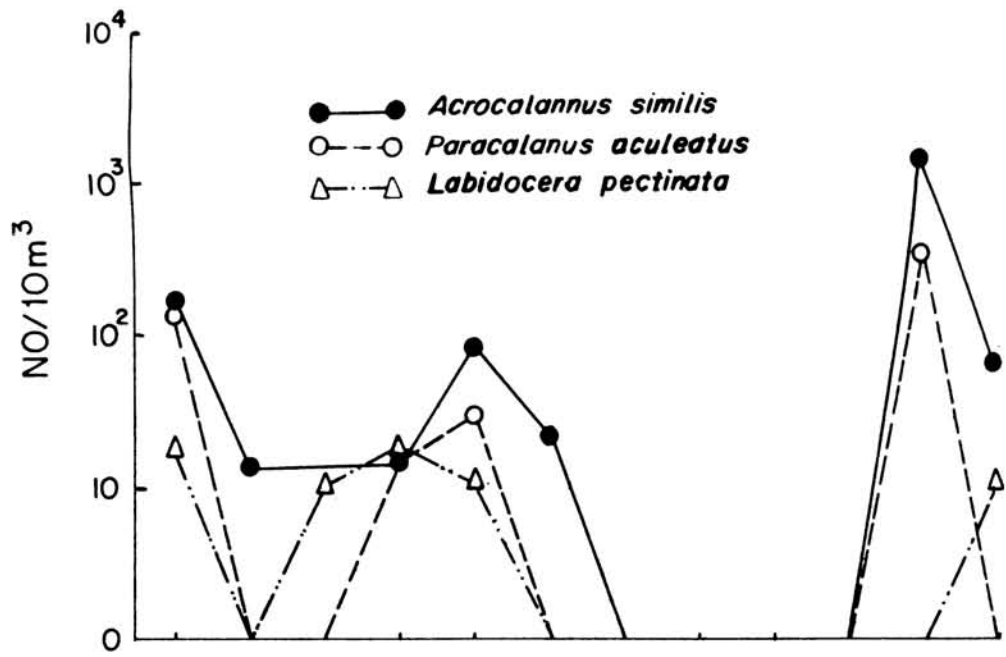


Fig. 25

**Fig. 26. Distribution of common ceped species
at Karapuzha.**

KORAPUZHA

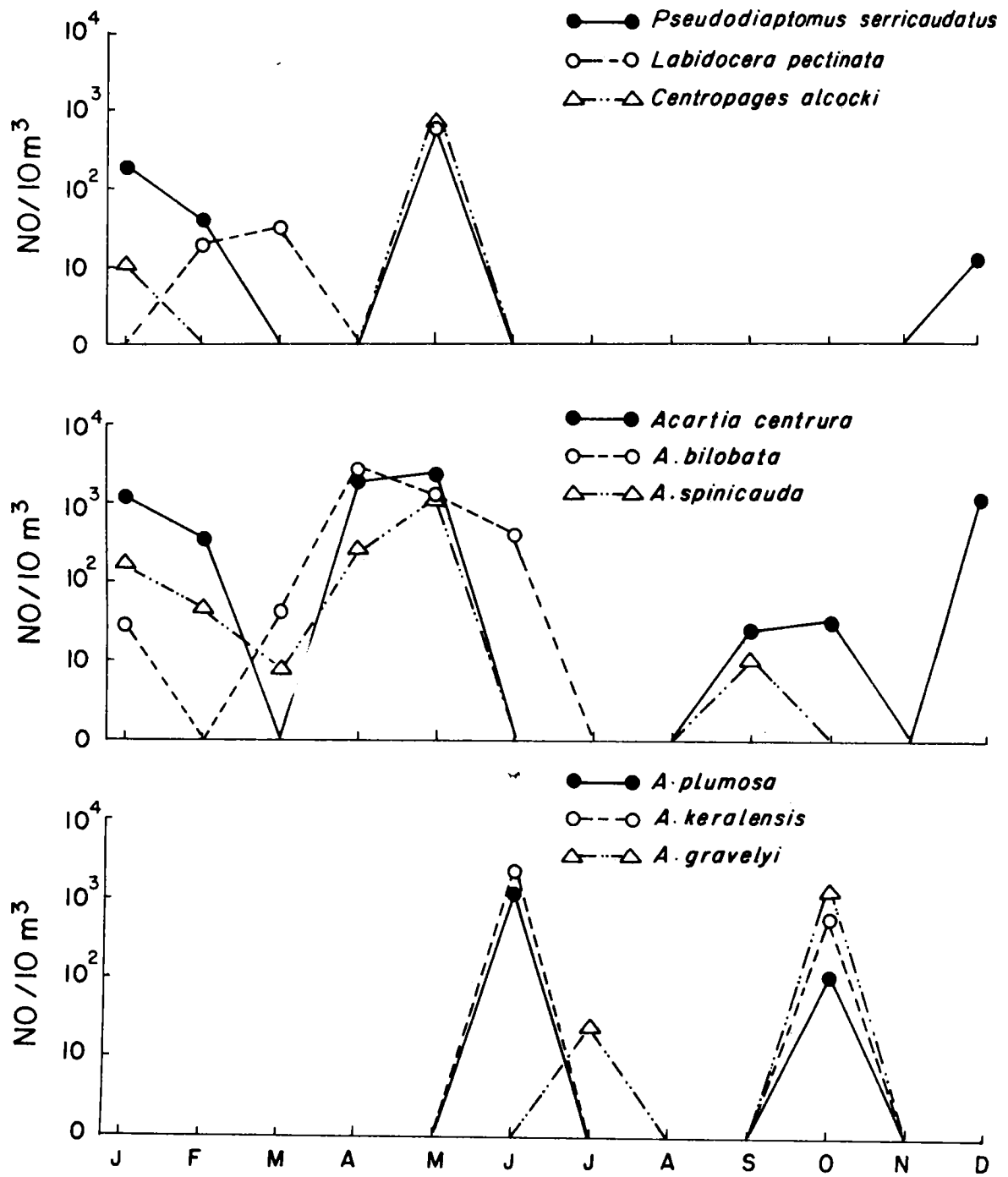


Fig. 26

**Fig. 17. Distribution of common copepod species
at Naha and Vohi.**

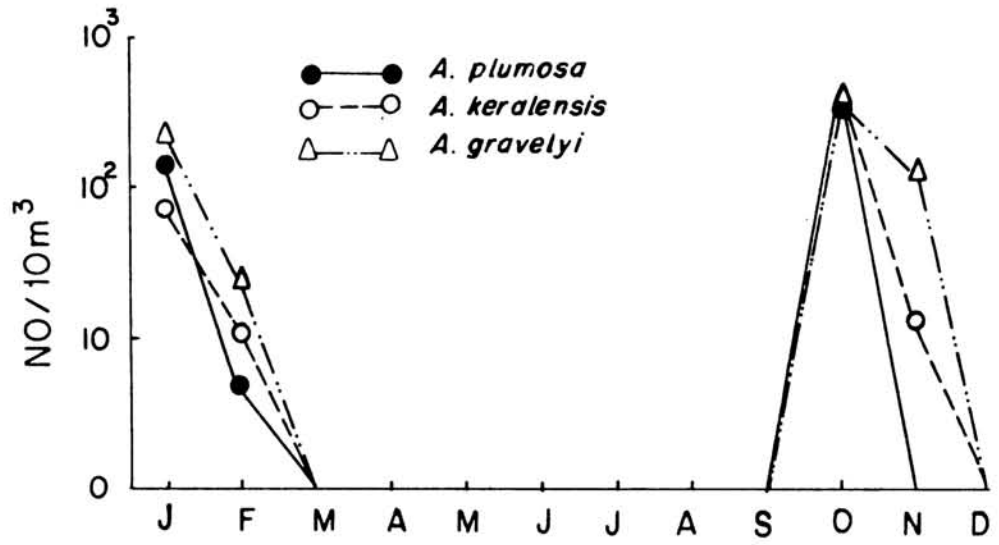
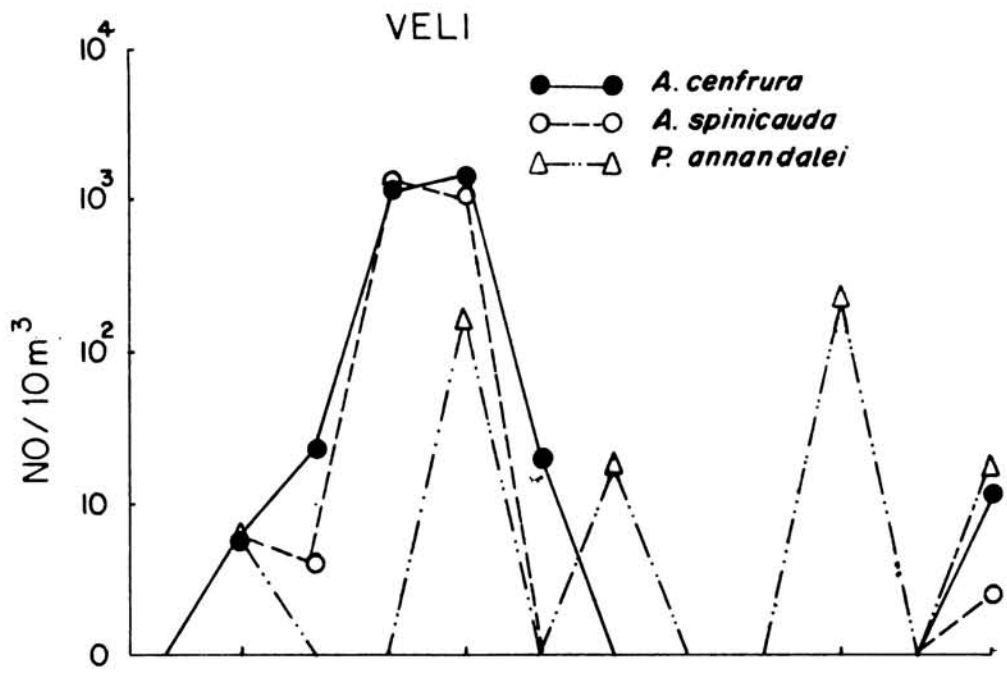
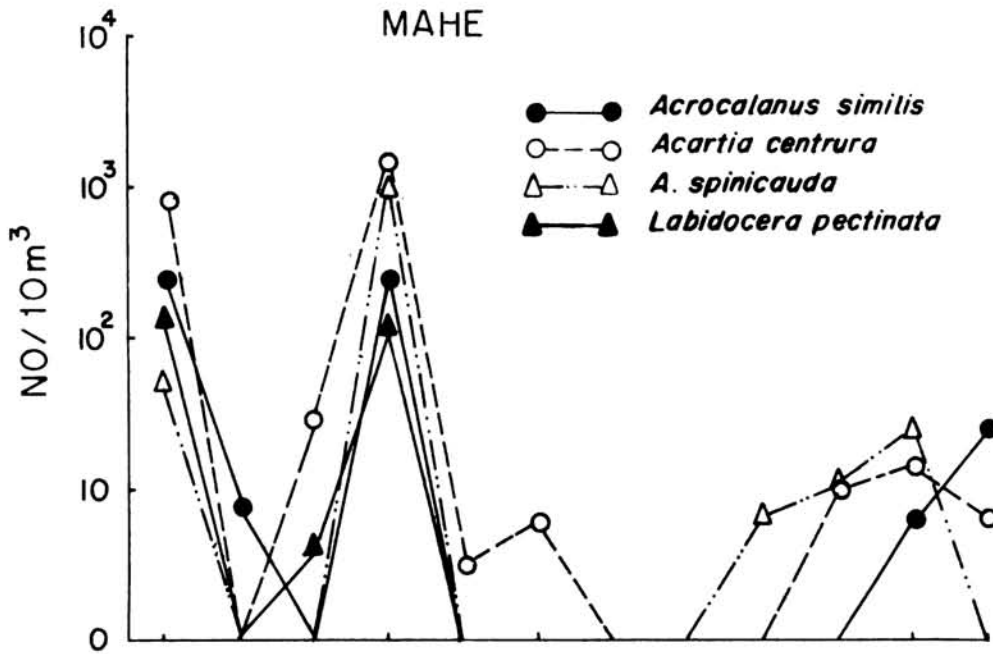


Fig. 27

**Fig. 28 A. Salinity ranges of common neoplankton species.
(Blank space shows the optimum range).**

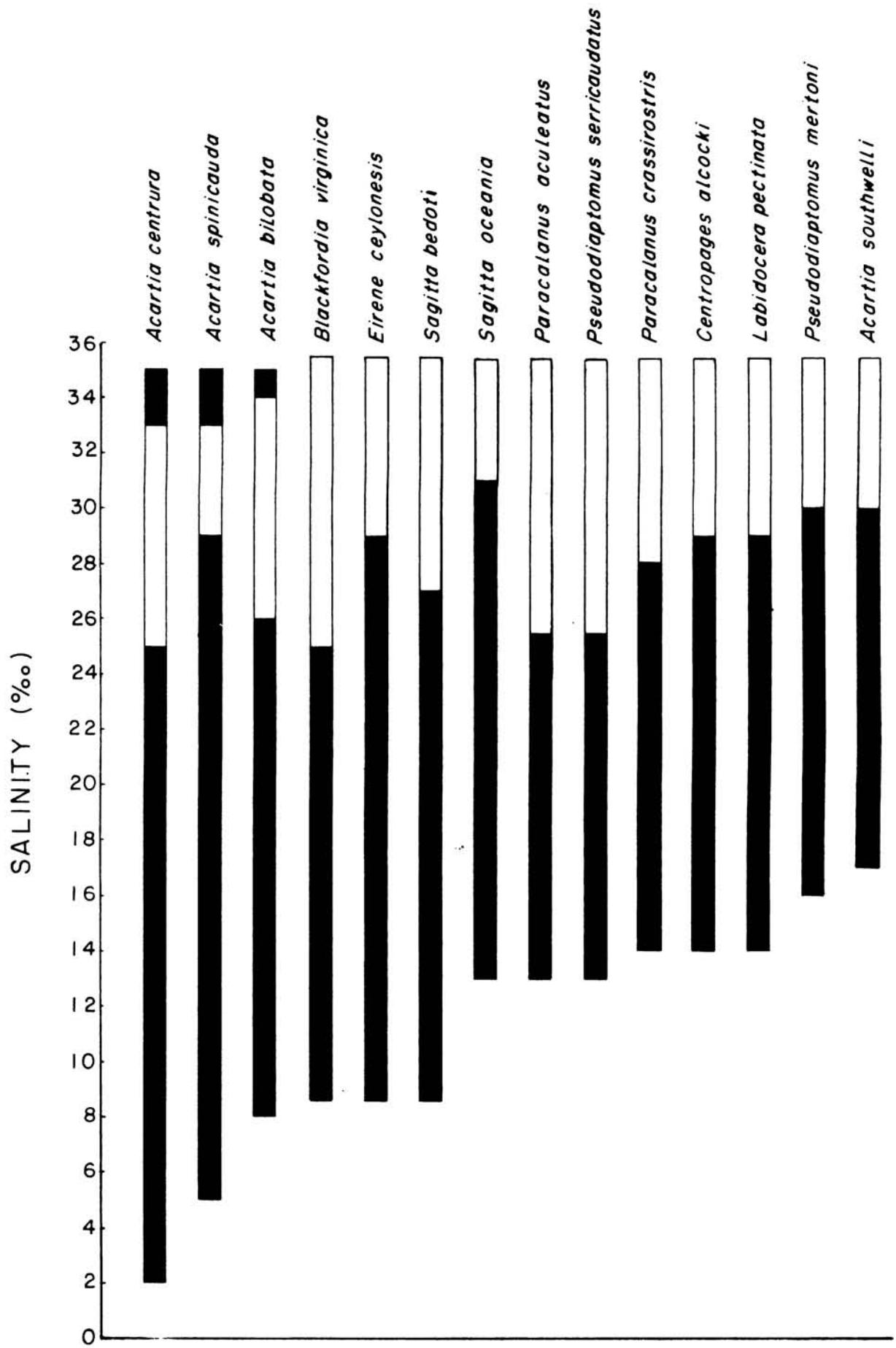


Fig. 28A

**Fig. 20 B. Salinity ranges of common zooplankton species.
(Blank space shows the optimum range).**

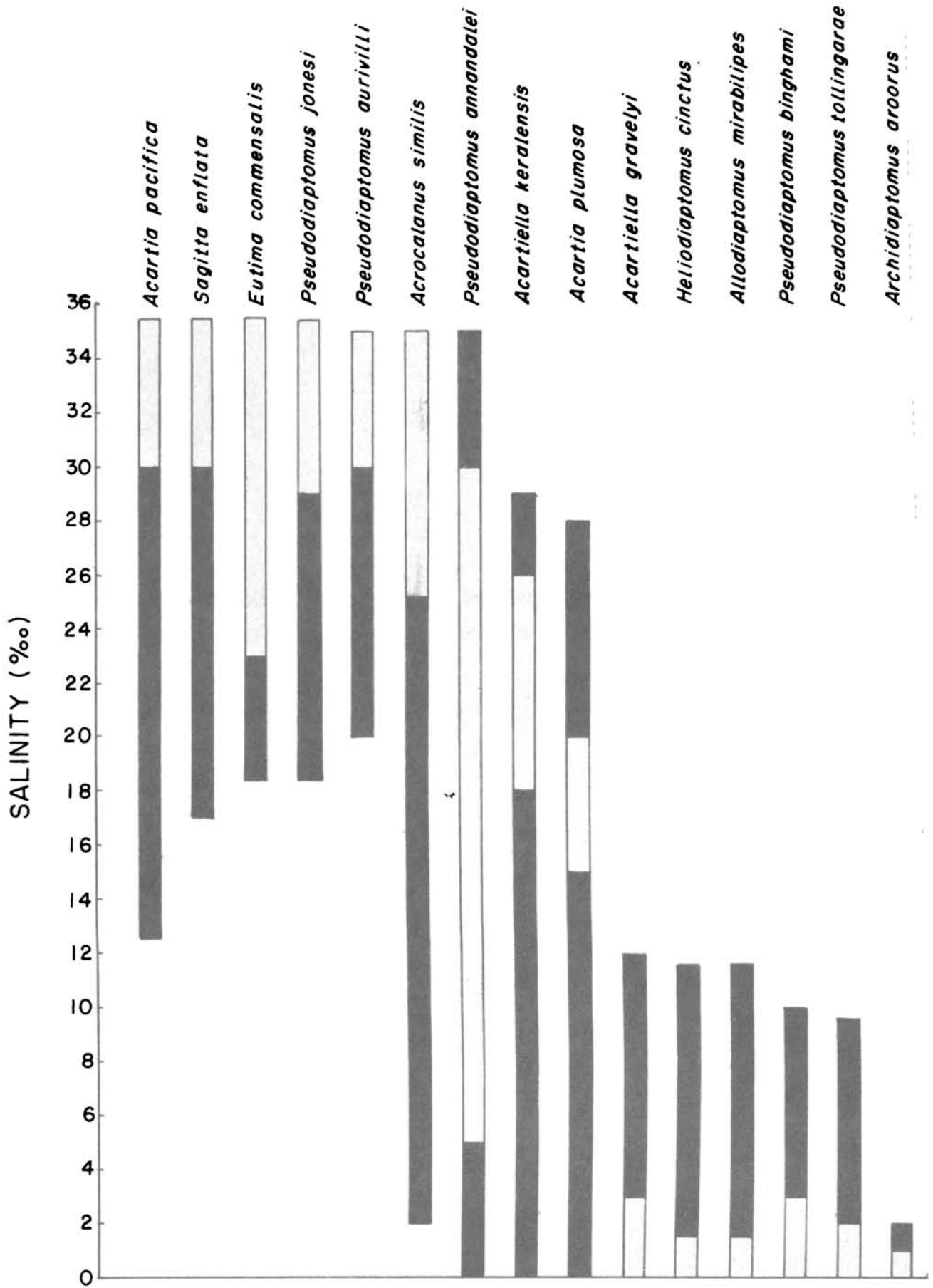


Fig. 28 B

Fig. 29 A. Curves showing gross surface primary production (adapted from Oasin *et al.*, 1969) and zooplankton standing stock in Cochin backwaters. The values are not strictly comparable, but the intention is to show the trends in productions.

Fig. 29 B. Curves showing relation between carnivores and herbivores in Cochin backwaters. The numbers of carnivores are magnified since they never occur in numbers comparable to herbivore abundance. Here also the purpose is only to show the relationship.

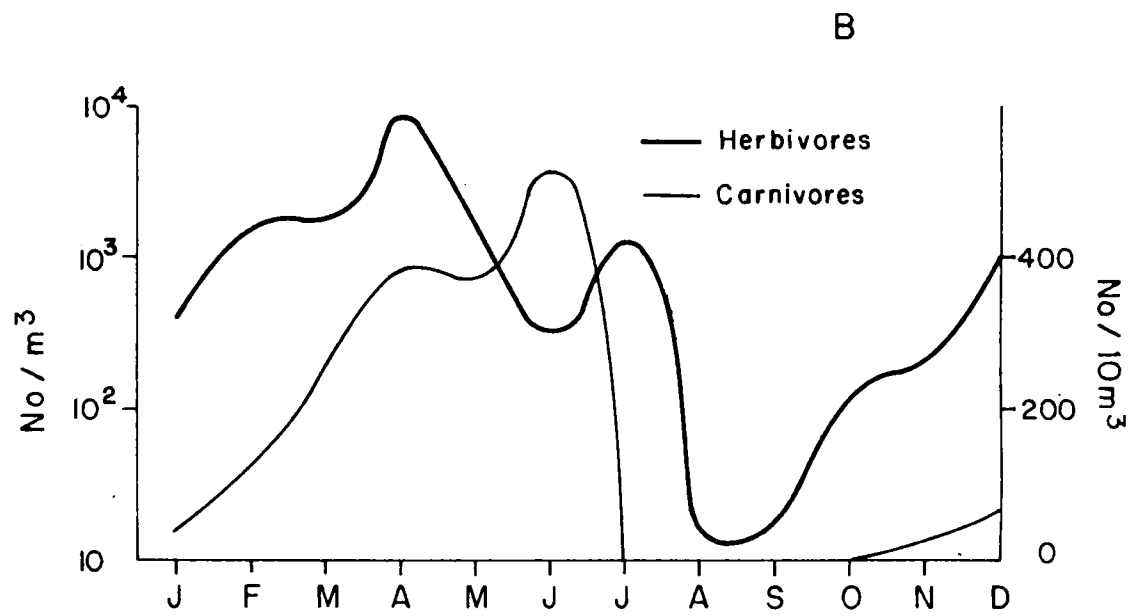
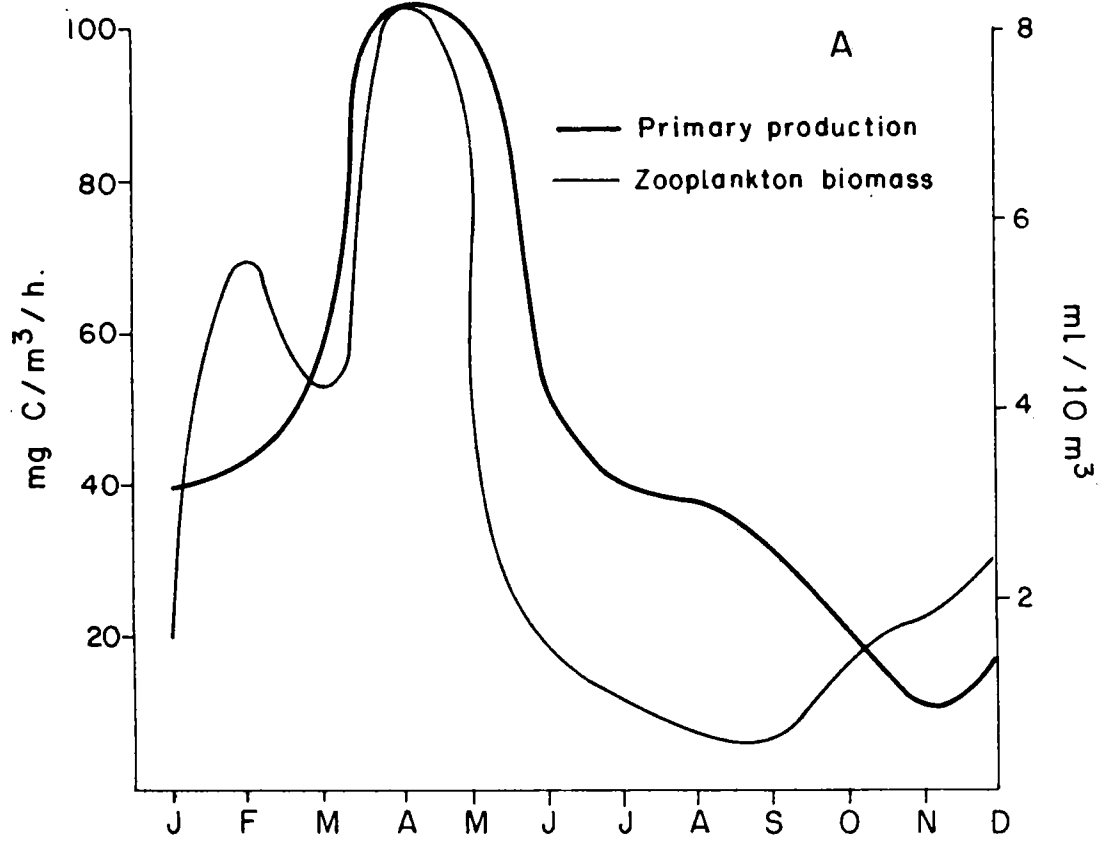


Fig. 29

Fig. 30 A,B & C. Succession of species groups/species in the eight estuaries during 1978.

- A - Acartia centrura
- B - Sagitta badoti
- C - Caridean larvae
- D - Other Decapod larvae
- E - Eutima commensalis
- F - Fish larvae
- G - Acartiella gravelvi
- H - Corophium triaenonyx
- J - Acetes sp.
- K - Acartiella keralensis
- L - Lucifer hansenii
- M - Plurobrechia globosa
- N - Pseudodiaptomus annandalei
- O - Acartia bilobata
- P - Paracalanus aculeatus
- R - P. crassirostris
- S - Evadne terrestina
- T - Oithona nana
- U - Acartia plumosa
- V - Acartia spinicauda
- W - Pseudodiaptomus aurivilli
- X - Heliodiaptomus cinctus
- Y - Allodiaptomus mirabilis
- Z - Zoea larvae

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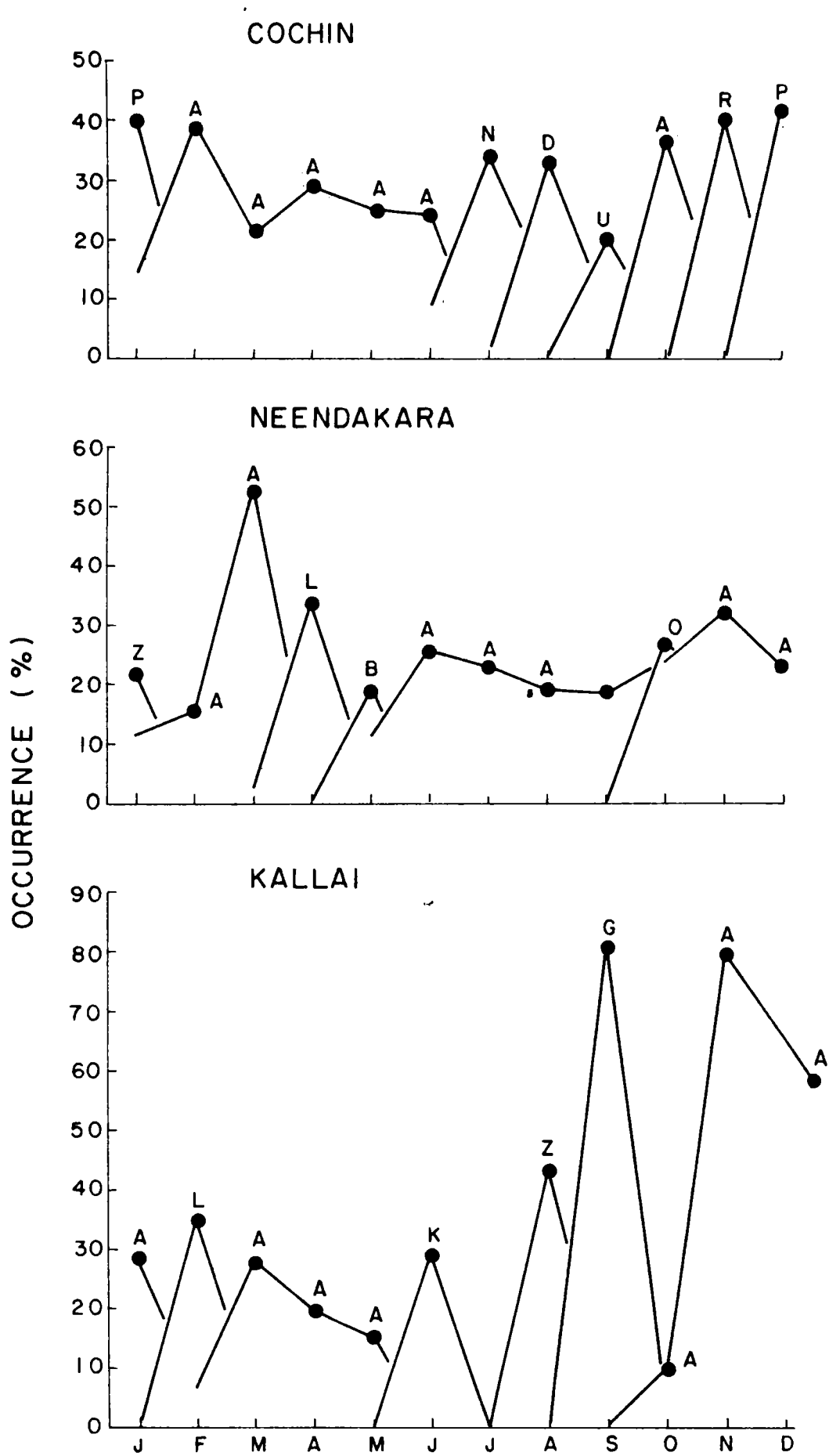


Fig. 30 A

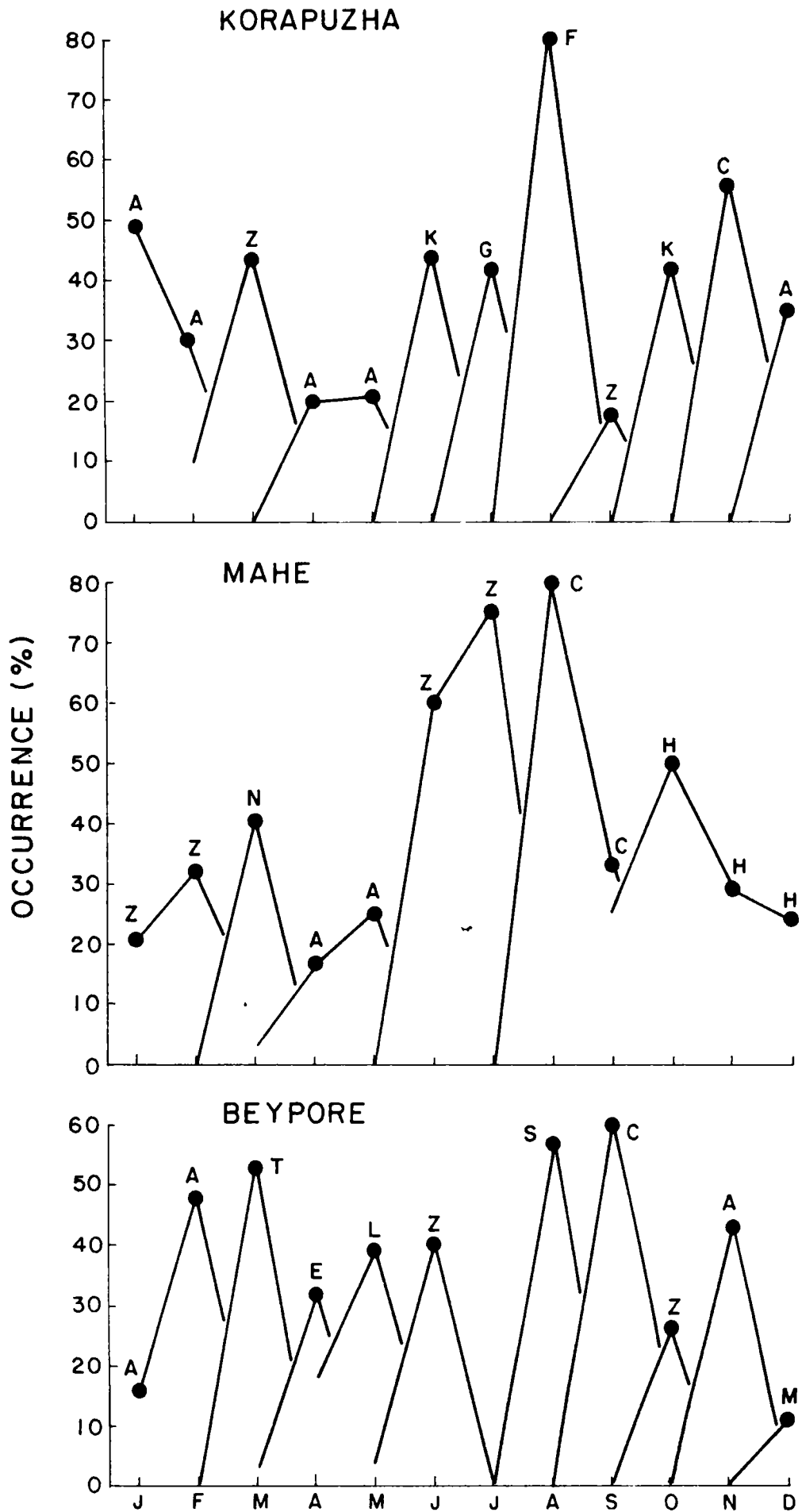


Fig. 30 B

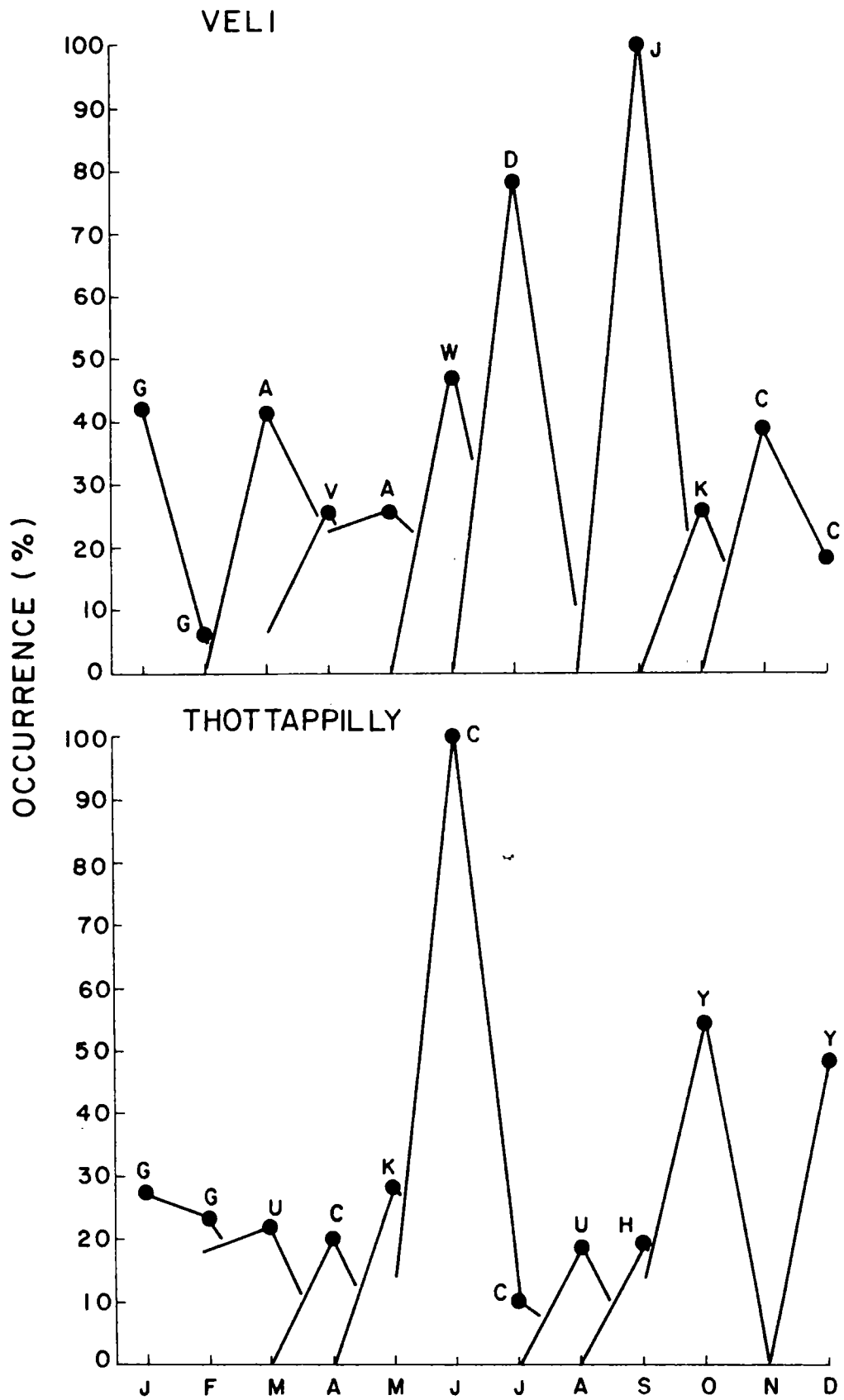


Fig. 30 C

**Fig. 31. Number of species and salinity distribution
in the Cochin bechevater system.**

COCHIN BACKWATERS

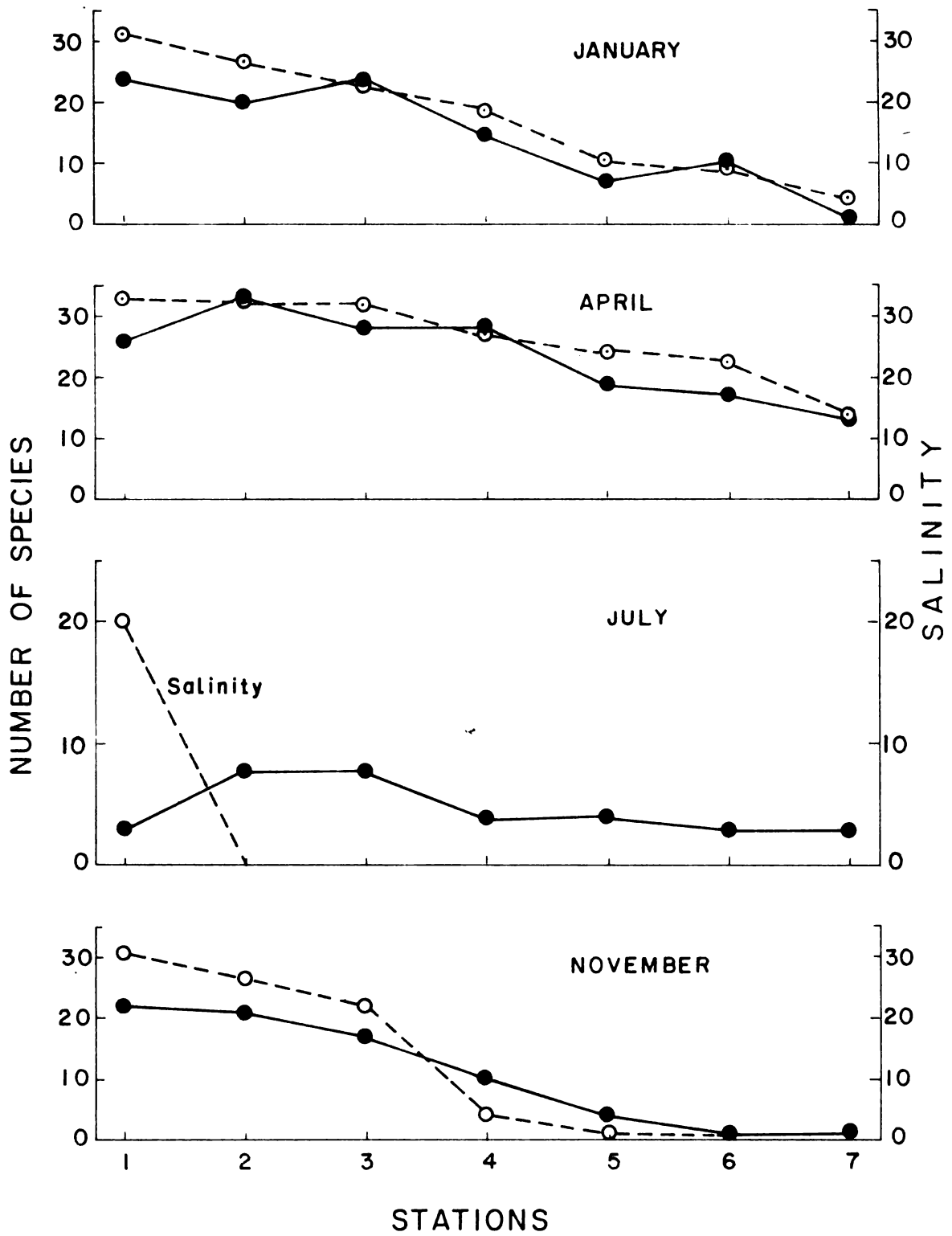


Fig. 31